# First Annual DOE Joint Genome Institute User Meeting

Sponsored By

U.S. Department of Energy Office of Science

March 29–April 1, 2006
Embassy Suites Hotel and DOE Joint Genome Institute
Walnut Creek, California

### **Contents**

Agenda	iv
Speaker Presentations	1
Abtracts in order of presentation according to agenda (p. iv)	
Poster Presentations	9
Posters alphabetical by first author. *Presenting author.	
Attendees	75
Author Index	83

### Agenda

(All events will take place in the Embassy Suites Contra Costa Room unless otherwise noted.)

#### WEDNESDAY - March 29, 2006

WEDNES	DAY – Marc	cn 29, 2000	C : Cl:	
Start Time	End Time	Subject	Session Chair or Speaker	
5:30 PM 6:30 PM Registration, Welcome Reception (Location: Embassy Suites,				
		Atrium)		
6:30 PM	8:00 PM	Keynote Addresses		
		Synthetic Biology	Drew Endy	
		Prochlorococcus: A Model for Systems Biology	Penny Chisholm	
THURSDA	AY – March	30, 2006		
8:15 AM	8:45 AM	Arrival, Registration		
8:45 AM	9:00 AM	Welcome, Introduction	Eddy Rubin	
9:00 AM	12:00 PM	Session I: Large Eukaryotic Genomics	Dan Rokhsar	
9:00 AM	9:30 AM	The Xenopus tropicalis Genome and its Uses	Richard Harland	
9:30 AM	10:00 AM	Chlamydomonas Genome	Arthur Grossman	
10:00 AM	10:30 AM	Plant Genomics	Joe Ecker	
10:30 AM	11:00 AM	Break		
11:00 AM	11:30 AM	Poplar Genome	Jerry Tuskan	
11:30 AM	12:00 AM	Mimulus guttatus	John Willis	
12:00 PM	2:00 PM	Lunch, Poster Session 1 (Location: Embassy Suite	es, Alameda Room)	
2:00 PM	5:00 PM	Session II: Microbial Genomics	Paul Richardson	
2:00 PM	2:30 PM	In Situ Gene Expression and Genomic Analysis	Dawn Holmes	
		of the Geobacteraceae		
2:30 PM	3:00 PM	Shewanella Genomes	Ken Nealson	
3:00 PM	3:30 PM	Post-Genomic Studies in the Facultative	Sam Kaplan	
		Photosynthetic Bacterium, <i>Rhodobacter</i> sphaeroides 2.4.1		
3:30 PM	3:45 PM	Break		
3:45 PM	4:00 PM	Talk Selected from Submitted Abstracts	TBA	
4:00 PM	4:30 PM	To Be Determined	Rachel Whitaker	
4:30 PM	5:00 PM	The White Rot Fungus Phanerochaete	Dan Cullen	
		chrysosporium		
5:00 PM	5:30 PM	Transportation to JGI		
		(Buses leave from the Embassy Suites Hotel Main En	trance)	
5:30 PM	8:00 PM	Reception, Poster Session 2, Tour of JGI (Location: JGI Conference Center)		
8:00 PM	8:30 PM	Transportation from JGI to Embassy Suites		

#### Agenda continued

FRIDAY - Marcl	h 3	1, 2006
----------------	-----	---------

FRIDAY – Start Time	End Time	Subject	Session Chair or Speaker
9:00 AM	12:15 PM	Session III: Microbial Communities	Jim Bristow
9:00 AM	9:30 AM	A Tale of Two Sludges	Phil Hugenholtz
9:30 AM	10:00 AM	Microbial Community Genomics in the	Ed DeLong
).30 / <b>(IVI</b>	10.00 71111	Ocean: From Gene Ecology to Systems	Lu DeLong
		Biology	
10:00 AM	10:30 AM	Ethanol-Producing <i>E. coli</i>	Lonnie Ingram
10:30 AM	11:00 AM	Break	Lonnic Ingram
11:00 AM	11:30 AM	Sequencing the Hindgut Bacterial Community	Jared Leadbetter
11.00 / 11.1	11.50 / 11.1	from a Wood-Feeding Termite Species from	surea Beausetter
		Costa Rica	
11:30 AM	12:00 AM	Bacteriophage DNA Polymerases and Other	David Mead
		Surprises from Superheated Thermal Aquifers	
12:00 AM	12:15 PM	Talk Selected from Submitted Abstracts	TBA
		·	
12:15 PM	1:45 PM	<b>Lunch, Users Business Meeting</b>	
1:45 PM	5:00 PM	Session IV: Technology Development	Sue Lucas
1:45 PM	2:15 PM	Solexa Technology	Gary Schroth
2:15 PM	2:45 PM	454 Technology	Michael Egholm
2:45 PM	3:15 PM	Talk - To Be Announced	TBA
3:15 PM	3:45 PM	Break	
3:45 PM	4:00 PM	Talk Selected from Submitted Abstracts	TBA
4:00 PM	4:30 PM	Using Short Sequencing Reads	Paul Richardson
4:30 PM	5:00 PM	Fly Comparative Genomics	Michael Eisen
CATUDDA	V Annil 1	2004	
9:00 AM	AY – April 1, 2 1:00 PM	Session V: IMG Workshop	Nikos Kyrpides
9:00 AM	9:30 AM	JGI Microbial Genome Analysis Pipeline	Athanasios Lykidis
9:30 AM	10:00 AM	Using IMG	Iain Anderson
10:00 AM	10:30 AM	Using IMG/M (IMG/Metagenomes)	Natalia Ivanova
10:30 AM	10:45 AM	Break	Natana Ivanova
10:45 AM	11:00 AM	Future Developments of IMG	Nikos Kyrpides
11:00AM	11:45 AM	Hands-On IMG	Tarkos Trytpiacs
11:45 AM	12:15 PM	Discussion/Feedback	
12:15 PM	1:00 PM	Hands-On IMG, Continued (Optional)	
12.13 1 101	1.001111	Tiands on 1110, Continuou (Optional)	
	1:00 PM	End of 2006 JGI User Meeting	

#### Speaker Presentations

Abtracts in order of presentation according to agenda (p. iv).

#### Prochlorococcus: A Model for Systems Biology

Penny Chisholm (chisholm@mit.edu)

Massachusetts Institute of Technology, Cambridge, MA

Prochlorococcus is the numerically dominant photosynthetic microbe in the tropical and sub-tropical oceans, often reaching concentrations of 10<sup>8</sup> cells per liter, and contributing a significant fraction of the total photosynthesis in ocean ecosystems. This group is comprised of closely related (less than 3% different in their rRNA sequence), phylogenetically and physiologically distinct 'ecotypes' that display different relative abundances along the light, temperature, nutrient, and predator (including viruses) gradients in the oceans. Genome sequences from a number of cultured Prochlorococcus strains, phages that infect them, and fragments of DNA from wild Prochlorococcus cells are providing exciting new insights into the origins and dynamics of micro-diversity within the Prochlorococcus meta-population, and the self-organization of microbial ecosystems.

*Prochlorococcus* is thus proving to be a useful model for developing cross-scale systems biology, the goal of which is to understand life processes at all scales of organization, from the cell to the biosphere.

## In Situ Gene Expression and Genomic Analysis of *Geobacteraceae*

Dawn Holmes (dholmes@microbio.umass.edu)

University of Massachusetts, Amherst, MA

To be able to optimize and predict the activity of *Geobacteraceae* in subsurface environments, it is necessary to first have an understanding of which *Geobacteraceae* genes are present in the environment and whether they are expressed *in situ*. Three different approaches were used to obtain genomic data from a uranium-contaminated aquifer in Rifle, CO in which the growth of *Geobacteraceae* was stimulated by the addition of acetate:1) multiple displacement amplification (MDA) of genomic DNA from single cells isolated from the environment; 2) small insert clone libraries constructed from genomic DNA extracted directly from the environment; and 3) targeted amplification of *Geobacteraceae* genes with degenerate primers. The genomic information obtained from these studies was then used to construct a number of quantitative RT-PCR primers targeting genes that are diagnostic of a range of metabolic states and highly conserved throughout the *Geobacteraceae*. Rates of metabolism, stress

#### **Speaker Presentations**

levels, chemotaxis, motility, and nutrient requirements of *Geobacteraceae* in groundwater samples collected from the U(VI) contaminated site during bioremediation were monitored with these primers. A direct correlation was observed between expression of a central metabolism gene in *Geobacteraceae*, malate dehydrogenase (*mdh*) and acetate concentrations detected in the groundwater. Expression of *ideR*, an iron dependent repressor remained constant throughout the experiment once *Geobacteraceae* became dominant members of the community, indicating that cells were limited for Fe(II) during the bioremediation process. In addition, the number of transcripts for *feoB* (iron uptake protein), the twitching motility protein (*pilT*), and the heavy metal resistance protein (*cusA*) increased significantly during U(VI) bioremediation. These results demonstrate that monitoring the *in situ* transcript levels of key genes can provide insight into the rates of metabolism and nutrient requirements of *Geobacteraceae* during *in situ* bioremediation.

## Post Genomic Studies in the Facultative Photosynthetic Bacterium, *Rhodobacter sphaeroides* 2.4.1

**Samuel Kaplan** (samuel.kaplan@uth.tmc.edu) University of Texas Health Science Center, Houston, TX

www.rhodobacter.org

Rhodobacter sphaeroides is a member of the  $\alpha$ -3 proteobacteria belonging to the non-sulfur purple photosynthetic group. R. sphaeroides possesses as extensive repertoire of reaction pathways enabling it to obtain energy, reducing power, carbon and nitrogen through a display of physiological diversity unmatched by most living systems. R. sphaeroides possesses a complex genome consisting of two chromosomes whose relationship to one another appears to date to the origin of the species. Of particular note is the ability of R. sphaeroides to undergo significant morphogenetic change when it is shifted from chemoheterotrophic growth in the presence of  $O_2$  to photoheterotrophic growth in the absence of  $O_2$ . Under these conditions, the cell membrane undergoes a series of invaginations to form the intracellular, vesicular membrane system or ICM, which comprises the photosynthetic apparatus whose abundance varies inversely to light intensity. The ICM is comprised of approximately 200-250 unique proteins of which only approximately 40 have been unequivocally identified, and which correspond to approximately 40-60% of the ICM protein. We now have the ability, for the first time, to identify all of the ICM proteins and their possible role(s) in photosynthesis and ICM structure through a combination of proteomic and post-genomic studies. Regulating the induction and synthesis of the ICM are: the PpsR repressor/AppA antirepressor unique to PS gene control, the Prr two component regulatory system which in addition to regulating ICM formation is a truly global regulatory element controlling either directly or indirectly over 800 genes/orfs, FnrL which is highly selective in PS gene

control and tetrapyrrole regulation. The major pathway through which O<sub>2</sub> exercises control over PS gene expression is through the terminal cytochrome c oxidase,  $cbb_3$  which signals the membrane bound PrrB histidine kinase to increase its phosphatase activity relative to its kinase activity which maintains the response regulator PrrA in the phosphate negative mode. The  $cbb_3$  terminal oxidase, in addition to its role in respiratory electron transport, is also shown to conduct electron flow under anaerobic conditions. In conjunction with this anaerobic electron flow the  $cbb_3$  plays a role in the regulation of over 230 genes/orfs and the ultimate electron acceptor for this anaerobic electron flow is thought to be an intermediate involved in carotenoid biosynthesis. The Prr system is shown to regulate either directly or indirectly over 21% of the R. sphaeroides genome. Given the complete DNA sequence of the R. sphaeroides 2.4.1 genome, its annotation and an array of tools, e.g., facile genetics, DNA chip, proteome profiles etc., we are in the unique position of being able to assess the complete composition and function of the ICM and the regulatory processes which accompany the transition of one growth state to another, as environmental conditions change.

This work is supported by grants from the NIH and the DOE.

#### The White Rot Fungus Phanerochaete chrysosporium

Dan Cullen (dcullen@facstaff.wisc.edu)

University of Wisconsin, Madison, WI

The most abundant source of global carbon is plant biomass, composed primarily of cellulose, hemicellulose and lignin. Many microorganisms are capable of degrading and utilizing cellulose and hemicellulose. However, a much smaller group of filamentous fungi have the ability to breakdown lignin, the most recalcitrant component of plant cell walls. Collectively referred to as white rot fungi, they possess the unique ability to efficiently degrade lignin to CO<sub>2</sub>. Common inhabitants of forest litter and fallen trees, these fungi have attracted considerable attention in a wide range of applications including fiber bleaching and hazardous waste remediation. All white rot fungi are members of the Basidiomycotina, a large and diverse phylum.

In 2002, the DOE Joint Genome Institute released the initial genome assembly (v1.0) for *Phanerochaete chrysosporium*, the most widely studied white rot fungus. Analysis of the ~30 Mb haploid genome revealed an impressive array of genes encoding extracellular proteins (Martinez et al 2004 *Nat. Biotechnol.* 22:695-700), many of which are likely involved in lignocellulose degradation. Among the secreted oxidative enzymes, complex families of peroxidases, copper radical oxidases, and FAD-dependent oxidases were identified. The genome also harbors an amazing number of putative carbohydrate-active enzymes, including at least 160 glycoside hydrolases. In addition to extended families of structurally

#### **Speaker Presentations**

related genes, an assortment of class I and II transposable elements (TEs) were found in the *P. chrysosporium* genome. Several TEs have disrupted and inactivated genes potentially involved in lignocellulose degradation.

The JGI's February 2005 release (v2.1) of improved gene models has paved the way for high throughput protein identification by LC-MS/MS. Recent results will be presented.

#### A Tale of Two Sludges

**Phil Hugenholtz** (phugenholtz@lbl.gov), <sup>1</sup> Héctor García Martín, <sup>1</sup> Victor Kunin, <sup>1</sup> S. Brook Peterson, <sup>3</sup> Natalia Ivanova, <sup>1</sup> Falk Warnecke, <sup>1</sup> Linda L. Blackall, <sup>2</sup> and Katherine D. McMahon<sup>3</sup>

<sup>1</sup>DOE Joint Genome Institute, Walnut Creek, CA; <sup>2</sup>Advanced Wastewater Management Centre, University of Queensland, Queensland, Australia; and <sup>3</sup>Civil and Environmental Engineering Department, University of Wisconsin, Madison, WI

Metagenomic datasets hold the allure of providing genome-wide insights into microbial population structure without the potential bias of cultivation. We obtained ca. 150 Mbp of shotgun sequence data from two sludge samples taken from geographically remote lab-scale bioreactors to investigate the structure and biogeography of the dominant and as-yet uncultivated bacterial species, Candidatus Accumulibacter phosphatis (CAP). Each sludge was dominated by a single nearly clonal CAP population that was flanked by a number of other low abundance CAP strains. The dominant strains were closely related sharing >95% sequence identity at the nucleotide level over most of their genomes. Differences were mostly localized in genomic regions encoding cassettes of exopolysaccharide synthesis genes and CRISPR elements that we speculate are highly variable in response to phage predation. We conclude that CAP can be globally dispersed possibly via aquatic reservoirs and that CAP populations in sludge may be inherently vulnerable to catastrophic collapse by virtue of their clonality and absence of detectable homologous recombination.

## Microbial Community Genomics in the Ocean: From Gene Ecology to Systems Biology

Edward F. DeLong (delong@mit.edu)

Massachusetts Institute of Technology, Cambridge, MA

Microbial life predominates in the ocean, yet much remains to be learned about its genomic variability, especially along the depth continuum. We recently conducted genomic analyses of planktonic microbial communities in the Pacific Ocean, from the surface waters to 4000 m deep. Sequence variation of microbial community genes reflected vertical zonation of

taxonomic groups, as well as differential distributions of functional genes. These distributional patterns of microbial genes suggested depth-variable community trends in carbon and energy metabolism, attachment and motility, gene mobility, and host-viral interactions. In addition, lateral gene transfer events that tracked environmental forcing factors and selective pressure were also observed. Our Pacific Ocean genomic depth profiles have revealed several ecological trends that apparently reflect higher order community organization and dynamics.

Despite the dizzying progress in microbial metagenomic sequencing, its important to recognize that a number of critical challenges lay ahead. The fostering of broad-based informatics efforts is a critical, including the integration of genomic datasets with environmental and ecological data. Measurement of microbial diversity and genomic variability, as well an microbial function process, and distributions, are increasing in scope, resolution and frequency. The various "snapshots" of gene content in ocean microbial populations may soon develop into motion pictures, that better describe genomic diversity, function and evolution in complex natural communities. Genome-derived functional hypotheses need to be tested, verified and quantified in the field. Similarly, process-oriented studies of individual "functional groups" need to strive towards simultaneous analyses of many different groups, to understand their dynamics and interactions. Genomic and post genomic tools may soon help bring the resolution and frequency of many biological measurements, on par with the physical and chemical measurements of environmental processes and phenomena.

## Sequencing the Hindgut Bacterial Community from a Wood-Feeding Termite Species from Costa Rica

**Jared Leadbetter** (jleadbetter@caltech.edu)
California Institute of Technology, Pasadena, CA

Termites play critical roles in plant lignocellulose turnover and the biological emissions of CO<sub>2</sub> and CH<sub>4</sub> globally. They are dependent in this endeavor on the activities of a unique microbial community housed within their hindguts. This community can contain on the order of 100 microbial species found no where else in nature. These microbial ecosystems are simple in an unusual sense of the word, because they are tiny, on the order of one microliter in volume, and have a distinct boundary—the gut lining of the insect. Yet because of the huge reach and great abundances of their hosts, these microbial ecosystems become major sites of catalysis that ultimately exert a huge impact on global C & N biogeochemistry, especially in tropical rainforests and similar biomes. Despite their mutually obligate interactions with their host, 80 years of study have revealed only a meager understanding of the roles, functions, and even the identities of the majority of termite gut microbes. This CSP has focused on making major, novel inroads into achieving a better understanding of the

#### **Speaker Presentations**

termite gut microbiota and its lignocellulose processing activities. A metagenomic study on the hindgut microbial community of a representative "higher" wood-feeding termite species, *Nasutitermes corniger*, is now well underway. Ca. 80 Mb of termite gut metagenome has been sequenced and is now being analyzed and annotated. This program involves key collaborations between laboratories at the California Institute of Technology in Pasadena; Diversa Corp. of San Diego; INBio, Costa Rica's National Biodiversity Institute; and the DOE Joint Genome Institute in Walnut Creek.

## **Bacteriophage DNA Polymerases and Other Surprises from Superheated Thermal Aquifers**

**David Mead** (dmead@lucigen.com), Vinay Dhodda, Robert DeFrancesco, Melodee Patterson, Paul Richardson, Mark Young, Phillip Brumm, and Thomas Schoenfeld

<sup>1</sup>Lucigen, Middleton, WI; <sup>2</sup>DOE Joint Genome Institute, Walnut Creek, CA; and <sup>3</sup>Montana State University, Bozeman, MT

Boiling thermal pools present an attractive target for community genomic analysis. They are physically isolated from other ecosystems, are among the most hostile environments known, and little is known about their constituents. The abundance of microbes and phage in thermal aquifers is significantly lower than fresh or salt water habitats, requiring new methods for extracting genetic information. Water column samples were collected from Bath hot spring, a boiling thermal pool in Yellowstone National Park, and adherent cells surrounding the pool. Planktonic organisms in the water column potentially originate from regions of 150-200°C found at depths of 100-300 meters throughout YNP. The microbial community of the water column was distinct from that found in the outflow sediment. Unusual and novel morphologies, some resembling multicellular assemblies, were observed in enrichment cultures. A pure culture demonstrating some of these characteristics has 16S rDNA similar to Thermus.

Waterborne viruses (phages) in thermal springs have been largely unexamined, despite their potential importance in the biosphere. Phages promote microbial diversity by predation of the most abundant microbes and by transfer of genes through transduction and lysogeny. Sequence analysis (>37,000 reads) of thermal aquifer phage libraries provides insight into viral complexity, lifestyles, molecular diversity, and access to coding sequences for useful proteins. Homology to a handful of cultivated viruses from high temperature springs establishes an ecological framework for these environmental samples. Numerous similarities to enzymes associated with temperate phage, such as integrases, suggest that lysogeny is common in thermal environments. Unexpectedly, a *Thermosynechococcus*-like photosynthesis regulatory gene was found next to an integrase gene. Over 200 DNA polymerase genes have been identified with approximately

#### **Speaker Presentations**

58 being full length. Ten active enzymes have been expressed, one of which allows isothermal amplification at elevated temperatures with greater specificity than conventional polymerases. This subset of thermostable phage DNA polymerases appears much more diverse than known microbial or phage enzymes. The fact that no *pol* genes were reisolated suggests the level of diversity seen so far is the tip of a very large iceberg.

#### Poster Presentations

Posters alphabetical by first author. \*Presenting author.

## 1. Industrial, Clinical and Environmental Significance of *Ralstonia pickettii*

**Catherine C. Adley\*** (catherine.adley@ul.ie) University of Limerick, Limerick, Ireland

#### 2. Sequence-Based Identification and Phylogenetic Analysis of Filamentous Fungi Isolated from Extreme Environments

**G. L. Andersen**, E. L. Brodie, T. DeSantis, P. Hu, Y. Piceno, and **T. Torok\*** (ttorok@lbl.gov)

Center for Environmental Biotechnology, Lawrence Berkeley National Laboratory, Berkeley, CA

Extremophillic microorganisms have become a valuable bioprospecting target because they represent a challenging scientific opportunity not only for those interested in microbial diversity and the evolution of life but for researchers searching for clues to extraterrestrial life. Extremophiles also produce unique biocatalysts that function under extreme conditions comparable to those prevailing in various industrial processes. Therefore, taxonomic identification and detailed characterization of these microorganisms provide a greater understanding of their diversity, unique metabolism, and ecological function.

Fungi are of great importance because (1) they are the primary decomposers and biosorbents in all terrestrial ecosystems, (2) they are important symbiotic associates of vascular plants both in mutualistic and parasitic relationships, (3) they constitute the overwhelming majority of plant pathogens and as such have a tremendous economic impact, and thereby, represent the next wave of potential bioterrorism threats, (4) several human pathogens are fungi, (5) they offer several well-developed genetic systems for molecular biologists as eukaryotic model organisms, (6) they are of significant potential for bioremediation, and finally (7) they are crucial to the fermentation and biotechnology industries.

Over a period of almost two decades, large numbers of filamentous fungi were isolated from extreme environments in the USA and the territory of the former Soviet Union. These environmental field sites were considered extreme in terms of their water activity, xenobiotic contamination, temperature, geology or chemistry. Published media and isolation techniques were modified and applied to simulate the eco-physiological conditions at the environmental habitat. The pure cultures of filamentous

#### **Poster Presentations**

fungi were preserved and stored at ultra-low temperature for long-term maintenance.

Currently ongoing work involves a collection of filamentous fungi that were isolated from environmental samples collected at and around the failed nuclear reactor in Chernobyl, Ukraine. The collection contains some 2,500 strains representing 98 genera and over 200 species. Characterization of these fungal organisms was carried out using classical colony- and cell morphology-based techniques, as well as molecular-level protocols. Here we compare the former with genomic DNA sequencing of specific target genes valuable for phylogenetic differentiation: preliminary data obtained by sequencing the D1/D2 domains of the large ribosomal subunit RNAgenes in almost 200 strains allowed us to confirm the classical identification results in over 85% of the cases. Based on that, we discuss our objectives of (1) developing a repository of high quality biomarker gene sequences from this valuable collection of extremophilic filamentous fungi and thereby providing a nucleus for an online database of fungal biomarkers, (2) generating a cohesive fungal biomarker sequence database available online, and (3) using the compiled sequences to select DNA sequences unique to fungal families, genera and species leading to the design of a state-of-the-art Affymetrix-style DNA microarray for fungal detection and identification.

## 3. The Genomes of Five Polycylic Aromatic Hydrocarbon Degrading Mycobacteria

**Anne J. Anderson\*** (anderson@biology.usu.edu) Utah State University, Logan, UT

#### 4. Community Sequencing of Novel Ultra-Small Euryarchaea from an Extremely Acidic Subsurface Mine, Iron Mountain

**Brett J. Baker\*** (bbaker@eps.berkeley.edu), <sup>1</sup> Gene W. Tyson, <sup>2</sup> Richard Webb, <sup>3</sup> Phil Hugenholtz, <sup>4</sup> Eric E. Allen, <sup>2</sup> and Jillian F. Banfield<sup>1, 2</sup>

<sup>1</sup>Dept. of Earth and Planetary Sciences, University of California, Berkeley, CA; <sup>2</sup>Environmental Sciences, Policy, and Management, University of California, Berkeley, CA; <sup>3</sup>Dept. of Microbiology and Parasitology, University of Queensland, Australia; and <sup>4</sup>DOE Joint Genome Institute, Walnut Creek, CA

Culture-independent PCR-based surveys have greatly expanded our knowledge of microbial diversity. However these studies are blind to the subset of microorganisms whose genes (e.g. 16S rRNA) do not contain conserved primer motifs. Random shotgun sequencing of an extremely acidic (pH <0.9) chemolithotrophic subsurface biofilm community

recovered a genome fragment from a novel archaeon, named WTF-1. This novel lineage's 16S rRNA gene has several mismatches with all commonly used broad-specificity PCR primers. The group is deeply branched within a region of the Euryarchaeota that has few cultivated representatives. WTF specific rRNA gene primers revealed two new groups in mine samples, WTF-2 and WTF-3, which are highly divergent from WTF-1 sequences (8% and 17% respectively). Fluorescence in situ hybridization and PCR-based detection of these groups has shown that these organisms are ubiquitous in microbial communities involved in acid mine drainage (AMD) generation at the Richmond Mine field site, but they are relatively low in abundance. Analysis of community sequence (100 Mb) from another site in the mine has revealed another ~38kb of WTF-2 and less of WTF-1. Using GC content, coverage (4X in this new library), and phylogenetic analyses of unassigned archaeal genomic fragments we have been able to identify >70kb of WTF in this new dataset. Comparison of a syntenic region between these two groups (WTF-1 and 2) shows that there are roughly 89% similar at the DNA level. It is difficult to get adequate coverage of organisms that occur in relatively low abundance via community genomic analysis without greatly over sampling the dominant members. Using filtration (0.45 µm), and PCR to confirm it, we have been able to enrich for these WTF groups. It is this filtrate that we are sequencing in order learn more about these novel Archaea.

#### 5. Flagellar Genes of Selaginella moellendorffii

Jody Banks\* (banksj@purdue.edu)
Purdue University, West Lafayette, IN

## 6. The Acidothermus cellulolyticus Genome: A Preliminary Analysis

**Ravi D. Barabote\*** (ravib@lanl.gov), Gary Xie, William S. Adney, Philippe Normand, Pierre Pujic, and **Alison M. Berry\*** (amberry@ucdavis.edu)

<sup>1</sup>Biosciences Division, Los Alamos National Laboratory, Los Alamos, NM; <sup>2</sup>National Renewable Energy Laboratory, National Bioenergy Center, Golden, CO; <sup>3</sup>Ecologie Microbienne, UMR CNRS, Universite Claude Bernard Lyon, Villeurbanne, France; and <sup>4</sup>Dept. of Plant Sciences, University of California, Davis, CA

Acidothermus cellulolyticus is a high GC actinobacteria that dwells in acid hot springs such as Yellowstone National Park. It was isolated in a screen for efficient and complete cellulose degradation, and the glycosyl hydrolases are highly thermotolerant. A. cellulolyticus is also of interest evolutionarily, because its closest phylogenetic neighbor is Frankia, a

nitrogen-fixing plant root symbiont. Recently, the genome of A. cellulolyticus has been sequenced by the JGI. Preliminary genomic characterization of A. cellulolyticus is presented. The 2.4 MB genome is approximately 67% GC-rich and is predicted to encode 2151 proteins. A significant proportion of the A. cellulolyticus proteins show closest homology to proteins from Streptomyces, Frankia, and Thermobifida fusca. Approximately 2% of its proteins do not have any recognizable homologs in other organisms and appear to be unique to A. cellulolyticus. Analyses of the predicted transporters revealed that the genome encodes very few recognizable membrane channel proteins, and approximately equal numbers of permeases and ABC-type transporters. The genome encodes numerous glycosyl transferases and other enzyme activities involved in the complex degradation pathway of cellulose. Although previous studies have shown a bias in relative abundance of certain amino acids in thermophiles, no significant differences were observed in a comparative analysis of the amino acid distributions in A. cellulolyticus and six mesophilic high GC actinobacteria. Analyzing the A. cellulolyticus proteins whose closest homologs are proteins from other thermophiles, such as T. fusca, may reveal insights into common evolutionary adaptations to thermophilic lifestyles of these organisms. The underlying basis for thermotolerance could be exploited to engineer proteins for industrial and environmental applications.

#### 7. Thellungiella halophila

Hans J. Bohnert\* (bohnerth@life.uiuce.edu) University of Illinois, Urbana-Champaign, IL

## 8. *Phaeodactylum tricornutum*: A Model Species for Exploring the Cell Biology of Diatoms

Chris Bowler\* (cbowler@biologie.ens.fr),<sup>1,2</sup> Andrew Allen,<sup>1</sup> Sacha Coesel,<sup>2</sup> Alessandra De Martino,<sup>1</sup> Angela Falciatore,<sup>2</sup> Marc Heijde,<sup>1</sup> Kamel Jabbari,<sup>1</sup> Uma Maheswari,<sup>1</sup> Manuela Mangogna,<sup>2</sup> Florian Maumus,<sup>1,2</sup> Anton Montsant,<sup>1,2</sup> Edda Rayko,<sup>1</sup> Magali Siaut,<sup>2</sup> and Assaf Vardi<sup>1</sup>

<sup>1</sup>Dept. of Biology, Ecole Normale Supérieure, Paris, France and <sup>2</sup>Cell Signalling Laboratory, Stazione Zoologica, Naples, Italy

Diatoms are eukaryotic, photosynthetic microorganisms found throughout marine and freshwater ecosystems that are responsible for around 20% of global primary productivity. A defining feature of diatoms is their ornately patterned silicified cell wall (known as frustule), which display species-specific nanoscale-structures. These organisms therefore play major roles in global carbon and silicon cycles. The marine pennate diatom *Phaeodactylum tricornutum* is the second diatom for which a whole

genome sequence has been generated. It was chosen primarily because of the superior genetic resources available for this diatom (e.g., genetic transformation, 80,000 ESTs), and because it has been used in laboratorybased studies of diatom physiology for several decades. Although not considered to be of great ecological significance, it has been found in several locations around the world, typically in coastal areas with wide fluctuations in salinity. Unlike other diatoms it can exist in different morphotypes, and changes in cell shape can be stimulated by environmental conditions. This feature can be used to explore the molecular basis of cell shape control and morphogenesis. Furthermore the species can grow in the absence of silicon, and the biogenesis of silicified frustules is facultative, thereby providing opportunities for experimental exploration of silicon-based nanofabrication in diatoms. The sequence is 30 mega base pairs and, together with the sequence from the centric diatom Thalassiosira pseudonana (34 Mbp; the first diatom whole genome sequence), it provides the basis for comparative genomics studies of diatoms with other eukaryotes and will provide a foundation for interpreting the ecological success of these organisms.

### 9. Microbial Systems Division: Metabolism of Microbial Communities

**Elbert Branscomb**, **Patrik D'Haeseleer\*** (patrikd@llnl.gov), Eivind Almaas, Jason Raymond, Tomer Altman, Jennifer Pett-Ridge, Steven Singer, Harry Beller, and Michael Thelen

Biosciences Directorate, Lawrence Livermore National Laboratory, Livermore, CA

The Microbial Systems Division at LLNL takes a Systems Biology approach towards understanding the energy metabolism of microbial communities. This includes integrating biochemistry, thermodynamics, metabolite transport and utilization, meta-genomic sequencing, regulatory and metabolic network analysis, and comparative and evolutionary genomics. Our objective is to predict and rationalize the dominant microbially mediated metabolic activities that develop in a given type of environment, and to predict a microbe's behavior and lifestyle directly from its genomic sequence.

#### 10. The JGI EST Pipeline

**Peter Brokstein\*** (pbbrokstein@lbl.gov), Jarrod Chapman, and Erika Lindquist

DOE Joint Genome Institute, Walnut Creek, CA

Expressed Sequence Tags (ESTs) provides a critical counterpart to whole genome shotgun sequence data. ESTs provide a direct evidence of

#### **Poster Presentations**

transcribed regions in the genome, greatly increasing the accuracy of JGI genome annotations. EST projects can also provide a large amount of information about a genome at a relatively low cost when compared to whole genome shotgun projects. ESTs are also used as part of the quality control process for genome assembly.

Over the past 2 years, the JGI EST Pipeline has processed 3,000,000 ESTs from 75 organisms. The pipeline is largely automated, and executes a workflow comprised of 44 distinct states. Tasks executed by the EST workflow include: vector trimming, quality trimming, contaminant screening, ab-initio clustering, cluster consensus sequence generation, and cluster consensus sequence annotation. The JGI EST Pipeline provides important feedback to the EST program laboratory process in the form of library quality and diversity statistics.

EST data is highly error prone and redundant, which necessitates a filtering process whereby ESTs from the same gene are grouped together into clusters. The process of Ab-initio clustering involves the comparison of the ESTs with one another, and the generation of clusters based on similarity between the ESTs. The JGI EST Pipeline uses a simple transitive closure graph based clustering approach derived from technology developed for the JAZZ genome assembler.

## 11. Complete Genome Sequences and Functional Genomics of Green Bacteria

**Donald A. Bryant**\* (dab14@psu.edu), <sup>1</sup> Tao Li, <sup>1</sup> Julia A. Maresca, <sup>1</sup> Aline Gomez Maqueo Chew, <sup>1</sup> Jörg Overmann, <sup>2</sup> Alex Copeland, <sup>3</sup> Susan Lucas, <sup>3</sup> Alla Lapidus, <sup>3</sup> Kerrie Barry, <sup>3</sup> J. Chris Detter, <sup>3</sup> Tijana Glavina, <sup>3</sup> Nancy M. Hammon, <sup>3</sup> Sanjay Israni, <sup>3</sup> Sam Pitluck, <sup>3</sup> Miriam L. Land, <sup>4</sup> Frank W. Larimer, <sup>4</sup> and Paul M. Richardson <sup>3</sup>

<sup>1</sup>Dept. of Biochemistry and Molecular Biology, Pennsylvania State University, University Park, PA; <sup>2</sup>Institut für Genetik und Mikrobiologie, Ludwig-Maximilians-Universität München, München, Germany; <sup>3</sup>DOE Joint Genome Institute, Walnut Creek, CA; and <sup>4</sup>Oak Ridge National Laboratory, Oak Ridge, TN

Green sulfur bacteria (GSB; or *Chlorobi*) and filamentous anoxygenic phototrophs (FAPs; *Chloroflexi*; formerly the green non-sulfur bacteria) are widespread but poorly characterized phototrophic eubacteria. In anoxic environments GSB can account for more than 80% of the carbon fixation and much of the nitrogen fixation and sulfur oxidation. The genome sequences of 8 *Chlorobi* and 7 *Chloroflexi* are being determined as part of an effort to define the metabolic diversity of anoxygenic phototrophic bacteria. Additionally, the two genomes of the phototrophic consortium, "*Chlorochromatium aggregatum*," are being determined. Each consortium consists of ~20 GSB cells attached to one β-proteobacterial cell, the "central rod," which has one polar flagellum. The central rod

provides motility to the entire consortium, which is phototactic due to light signals received by the epibiont GSB cells. It is anticipated that a novel mechanism of cell-to-cell communication may exist between these organisms, and the metabolic basis for their co-dependency is unknown.

Ten sequenced GSB genomes range from 2 to 3 Mb and encode about 1700 to 2800 genes. The FAP genomes are larger, about 5.5 to 6 Mb in size, and they encode more than 5000 genes. Moderately thermophilic Chlorobium tepidum [Eisen et al., PNAS, 2002] has become an important model GSB for functional genomics studies because it is efficiently and naturally transformable. Using various comparative bioinformatics approaches, we have identified and verified all steps in the carotenoid biosynthetic pathway and all but one step in the bacteriochlorophyll (BChl) c biosynthetic pathway. This work has identified more than 10 novel genes and the enzymes they encode. For example, two novel lycopene cyclases have been characterized, which are the first members of a large family of previously unknown lycopene cyclases found in GSB and cyanobacteria. By comparing the genomes of brown-colored GSB, which synthesize BChl e and the important biomarker carotenoid isorenieratene, and green-colored GSB, which produce BChl c and the carotenoid chlorobactene, we have identified a 6-kb gene cluster encoding 4 genes. This cluster includes a gene required for isorenieratene biosynthesis and seems likely to confer the physiologically and ecologically important trait of "brownness," since a radical SAM enzyme encoded in this cluster is probably responsible for the introduction of the C7 formyl group into BChl e. BChl e and isorenieratene synthesis are codistributed, polyphyletic traits, which almost certainly spread among GSB by lateral gene transfer. These pigments confer the ability to grow much deeper in stratified aquatic environments where sulfide concentrations are typically higher. We are currently testing whether these four genes can transform the green-colored C. tepidum to "brownness" by introducing this 6-kb gene cassette by transformation into a non-essential chromosomal gene. Finally, *Chlorobium ferrooxidans* is the only known GSB that cannot oxidize sulfide but instead oxidizes ferrous iron to provide the electrons for carbon fixation via the reverse TCA cycle. Comparative analyses confirm that *C. ferrooxidans* lacks all known genes required for thiosulfate, sulfide, polysulfide, and sulfite oxidation except for sulfide quinone reductase, and candidate genes for iron oxidation have been identified.

## 12. Comparative Genomics and the California Purple Sea Urchin

**R.** Andrew Cameron\* (acameron@caltech.edu) and Eric Davidson Biology Division and Center for Computational Regulatory Genomics, California Institute of Technology, Pasadena, CA

The extensive knowledge base in cell and developmental biology of the California purple sea urchin plus the remarkably complete echinoderm fossil record afford an excellent opportunity to inform the annotation of the gene set predicted from the assembled genome. Two unique echinoderm features are the water vascular system and calcium carbonate skeleton. It is to be expected that the differentiation gene batteries used in these structures are likely to be de novo constructions from ancestral genes. Other unique biological characteristics of the echinoderms such as catch collagen, odorant receptors, the digestive enzymes used in marine herbivory, echinochrome pigments, body wall components and teeth ought to likewise involve distinctive sets of genes. A number of genome-wide comparisons to the sea urchin sequences using the several well annotated bilaterian genomes currently available further highlight some of these unique features. About 10 of the most prevalent domains in the sea urchin gene models are not as abundant in the other genomes and thus constitute expansions that are, at least, specific to sea urchins if not to the complex of echinoderms and hemichordates that form the sister group to the chordate deuterostomes. Obvious among these are the motifs that correspond to genes encoding innate immunity proteins. These analyses show that the specific complement of genes in the sea urchin genome result largely from expansion and contraction of existing families from the common bilaterian "toolkit" of genes

## 13. The Genome of Obligate Methylotroph *Methylobacillus* flagellatus

**Ludmila Chistoserdova\*** (milachis@u.washington.edu), Alla Lapidus, Cliff Han, Lynne Goodwin, Liz Saunders, Roxanne Tapia, Paul Gilna, Susan Lucas, Paul M. Richardson, and Mary E. Lidstrom

<sup>1</sup>University of Washington, Seattle, WA; <sup>2</sup>DOE Joint Genome Institute, Production Genomics Facility, Walnut Creek, CA; and <sup>3</sup>DOE Joint Genome Institute, Los Alamos National Laboratory, Los Alamos, NM

The complete genome of a model obligate methylotroph, *M. flagellatus* (strain KT) was sequenced. The genome is represented by a single circular chromosome of approximately 3 Mbp potentially encoding a total of 2,766 proteins. Based on genome analysis as well as the results from previous genetic and mutational analyses, methylotrophy is enabled by methanoland methylamine dehydrogenases, the tetrahydromethanopterin-linked formaldehyde oxidation pathway, the assimilatory and dissimilatory

branches of the ribulose monophosphate cycle, and a formate dehydrogenase. Some of the methylotrophy genes are present in more than one (identical or non-identical) copy. The obligate dependence on single carbon compounds is likely due to the incomplete tricarboxylic acid cycle, as no genes potentially encoding alphaketoglutarate-, malate- or succinate dehydrogenases are identifiable. The genome of *M. flagellatus* was compared, in terms of methylotrophy functions, to the previously sequenced genomes of three methylotrophs: *Methylobacterium extorquens* (Alphaproteobacterium, 7 Mbp), *Methylibium petroleophilum* (Betaproteobacterium, 4 Mbp), and *Methylococcus capsulatus* (Gammaproteobacterium, 3.3 Mbp). Strikingly, metabolically and/or phylogenetically, methylotrophy functions in *M. flagellatus* are more similar to the ones in *M. capsulatus* and *M. extorquens* than to the ones in the more closely related *M. petroleophilum*, providing the first genomic evidence of polyphyletic origin of methylotrophy in Betaproteobacteria.

## **14.** Environmental Genomic Characterization of a Deep Subsurface Microorganism

**Dylan Chivian\*** (dcchivian@lbl.gov), <sup>1,2</sup> Eric J. Alm, <sup>2,3</sup> Fred J. Brockman, <sup>4</sup> Eoin L. Brodie, <sup>1</sup> David E. Culley, <sup>4</sup> Thomas Gihring, <sup>5</sup> Alla Lapidus, <sup>6</sup> Li-Hung Lin, <sup>7</sup> Duane P. Moser, <sup>8</sup> Paul Richardson, <sup>6</sup> Adam P. Arkin, <sup>1,2,9,10</sup> Terry C. Hazen, <sup>1,2</sup> and Tullis C. Onstott <sup>11</sup> Lawrence Berkeley National Laboratory, Berkeley, CA; <sup>2</sup> Virtual Institute for Microbial Stress and Survival, Berkeley, CA; <sup>3</sup> Massachusetts Institute of Technology, Cambridge, MA; <sup>4</sup> Pacific Northwest National Laboratory, Richland, WA; <sup>5</sup> Florida State University, Tallahassee, FL; <sup>6</sup> DOE Joint Genome Institute, Walnut Creek, CA; <sup>7</sup> National Taiwan University, Taipei, Taiwan; <sup>8</sup> Desert Research Institute, Las Vegas, NV; <sup>9</sup> University of California, Berkeley, CA; <sup>10</sup> Howard Hughes Medical Institute, Berkeley, CA; and <sup>11</sup> Princeton University, Princeton, NJ

The deep subsurface planktonic community found at depths greater than 1.5 km across central South Africa is dominated by a species of *Firmicutes* that has never been successfully cultivated. A metagenome was assembled from DNA obtained from fracture water emanating from a borehole at 2.8 kilometers depth in a South African Gold mine. The draft assembly is consistent with a single microorganism, and implies that this species is capable of living in isolation in this deep subsurface environment. Genes for dissimilatory sulfate reduction were present in the genome. Based the 16S rRNA gene, the most similar described organism is *Desulfotomaculum kuznetsovii* (~91% identity), making this one of the first sequenced genomes of a dissimilatory sulfate reducing Gram-positive bacterium. In addition to sulfate reduction, the genome indicates that the organism is capable of formate oxidation. The genome also contains: 1) the acetogenic acetyl-CoA (Wood-Ljungdahl) pathway, 2) a partial TCA cycle, 3) a N<sub>2</sub> fixation pathway, 4) genes for sporulation and germination,

#### **Poster Presentations**

5) heat shock proteins, 6) genes for pilus formation, and 7) genes for flagellum formation and chemotaxis. The variety of metabolic pathways and chemotactic capability is not suggestive of a streamlined genome for a sulfate reducer in an energy depleted environment, but rather more consistent with a motile sulfate reducer in an energy rich environment that actively seeks a specific subsurface niche when present and is capable of surviving long periods of time when that niche is absent. The fracture zone geochemistry is consistent with these inferences. The apparent absence of O<sub>2</sub> tolerance genes indicates the organism is an obligate anaerobe consistent with an indigenous origin.

## 15. Sequencing *Arabidopsis lyrata* and *Capsella rubella*: What We Can Learn from the Genomes of *A. thaliana*'s Close Relatives

**Richard M. Clark\*** (richard.clark@tuebingen.mpg.de), <sup>1</sup> Yalong Guo, <sup>1</sup> Joy M. Bergelson, <sup>2</sup> Justin O. Borevitz, <sup>2</sup> Brandon S. Gaut, <sup>3</sup> Anne E. Hall, <sup>4</sup> Charles H. Langley, <sup>5</sup> Barbara Neuffer, <sup>6</sup> Klaus F. X. Mayer, <sup>7</sup> Magnus Nordborg, <sup>8</sup> Outi Savolainen, <sup>9</sup> Yves van de Peer, <sup>10</sup> Stephen I. Wright, <sup>11</sup> and Detlef Weigel<sup>1</sup>

<sup>1</sup>Dept. of Molecular Biology, Max Planck Institute for Developmental Biology, Tübingen, Germany; <sup>2</sup>Dept. of Ecology and Evolution, University of Chicago, Chicago, IL; <sup>3</sup>Dept. of Ecology and Evolutionary Biology, University of California, Irvine, CA; <sup>4</sup>Dept. of Molecular Genetics and Cell Biology, The University of Chicago, Chicago, IL; <sup>5</sup>Section of Evolution and Ecology, University of California, Davis, CA; <sup>6</sup>Dept. of Systematic Botany, Universität Osnabrück, Osnabrück, German; <sup>7</sup>Munich Information Center for Protein Sequences, Institute for Bioinformatics, Gesellschaft fur Strahlenforschung Research Center for Environment and Health, Neuherberg, Germany; <sup>8</sup>Molecular and Computational Biology, University of Southern California, Los Angeles, CA; <sup>9</sup>Dept. of Biology, University of Oulu, Oulu, Finland; <sup>10</sup>Dept. of Plant Systems Biology, Flanders Interuniversity Institute for Biotechnology (VIB), Ghent University, Ghent, Belgium; and <sup>11</sup>Dept. of Biology, York University, Toronto, Canada

Arabidopsis thaliana has many advantages that make it a powerful system for genetic and genomic studies. These include a short generation time, small genome size (~125 Mb), easy DNA-mediated transformation, and extensive intraspecific polymorphism datasets. While these resources rival those available in other multicellular reference organisms such as C. elegans, D. melanogaster and M. musculus, reference genome sequences from close relatives of A. thaliana have been lacking. In order to leverage genomic information available from A. thaliana, as well as to better understand plant genome evolution, we have proposed two close relatives for genome sequencing (A. lyrata and Capsella rubella). Both genomes are now scheduled for ~8-fold shotgun genomic sequencing by

the Department of Energy Joint Genome Institute in 2006. From a large number of close relatives, *A. lyrata* and *C. rubella* were selected for sequencing because of their comparatively small genome sizes (~230 and 250 Mb, respectively), interesting life histories (*e.g.*, selfing versus outcrossing), and existing genetic and genomic resources (*e.g.*, genetic maps and BAC libraries are available for both species). Comparing the *A. thaliana*, *A. lyrata*, and *C. rubella* genome sequences will provide an unprecedented opportunity to understand dynamic mutational processes, such as genome rearrangements and gene duplication/deletions, that appear to occur in plants at much higher rates than in animals. Given the recent divergence times for these species (~5-10 million years), a particular interest will be to understand the basis for the approximately 2-fold difference in genome size between *A. thaliana* and both *A. lyrata* and *C. rubella*. An overview of the project and planned analyses will be presented.

## 16. The *Daphnia* Genomics Consortium Partnership with the DOE Joint Genome Institute: Towards a Model System for Ecological Genomics

**John Colbourne\*** (jcolbour@cgb.indiana.edu), <sup>1</sup> Michael Pfrender, <sup>2</sup> Kelley Thomas, <sup>3</sup> Justen Andrews, <sup>1</sup> Michael Lynch, <sup>1</sup> and Jeffrey Boore <sup>4</sup> <sup>1</sup> Indiana University, Bloomington, IN; <sup>2</sup> Utah State University, Logan, UT; <sup>3</sup> University of New Hampshire, Durham, NH; and <sup>4</sup>DOE Joint Genome Institute, Walnut Creek, CA

The freshwater crustacean *Daphnia* has attracted attention from biologists for over two centuries because of its sentinel role within nearly all freshwater ecosystems and its unique habits when responding to environmental challenges. For instance, within just a short time (~4-6 million years) members of the D. pulex species complex have adapted to the numerous physical and biological factors that impact life in ephemeral pools, eutrophic lakes, oligotrophic reservoirs, UV radiated ponds and saline habitats. Within yet shorter periods (~100 years), many populations have faced novel stressors from industrial activities. In many cases, the response of *Daphnia* populations involves phenotypic plastic traits that rapidly modify their morphology, physiology and even their mode of reproduction, which are triggered by cues from the environment. Although the Daphnia system is ideal for genetic studies on the potential and limits of adaptive evolution, only recently have we produced the necessary tools that can link the structure and expression of the genome to individual fitness and population-level responses of *Daphnia* to environmental stress.

In partnership with the Joint Genome Institute, the *Daphnia* Genomics Consortium has created (1) a microsatellite-based genetic map to identify ecologically important genes by quantitative trait locus (QTL) analyses, (2) microarrays for the functional annotation of *Daphnia* genes and (3) a genome sequence. Expressed sequence tags (ESTs) were also created from

#### **Poster Presentations**

Daphnia exposed to 12 distinct ecological stressors ranging from toxic metals and UV radiation to hypoxia, starvation and predation. So far, 12,600 unique genes from 71,000 high quality ESTs were recovered. Clear patterns exist among the different libraries that highlight classes of loci transcribed under various ecological conditions. Yet overall, the majority of sequenced cDNA from Daphnia (>55%) show no similarity to genes from other organisms. Although the candidate gene approach will prove to be useful for ecological studies – by selecting loci that share sequence conservation with functionally well characterized genes of other model species – microarrays that probe for all gene transcripts will undoubtedly be a more valuable tool for finding ecologically important loci.

Acknowledgements: Elizabeth Bohuski, Jeong-Hyeon Choi, Melania Cristescu, Brian Eads, Jelena Radivojac, (Indiana University), Erika Lindquist, Peter Brokstein, Harris Shapiro (JGI), Darren Bauer (University of New Hampshire), Joseph Shaw, Jennifer Davey, Joshua Hamilton, Carol Folt, Celia Chen (Dartmouth College).

This project is supported by the Department of Energy's Office of Science and the National Science Foundation.

# 17. An Overview of the Capillary Electrophoresis Process at the DOE Joint Genome Institute Production Genomics Facility: The Dual Operation of the AB 3730xl and GE MegaBACE 4500 DNA Sequence Analyzers

Christopher Daum\* (daum1@llnl.gov), Lena Philip\*, Danielle Mihalkanin, Cailyn Spurrell, Don Miller, Susan M. Lucas, Alex Copeland, Damon Tighe, Eric Abbott, Marlon Arcaina, Melanie Lafrades, Albert Linkowski, Adrienne Loero-Pequignot, Andy Yuen, and Mathew Zane U.S. DOE Joint Genome Institute, Walnut Creek, CA

At the center of the Department of Energy's (DOE) Joint Genome Institute (JGI) Production Genomics Facility (PGF), lies a highly efficient and automated production line devoted to the generation of high-quality genomic DNA sequence. The JGI utilizes a dual platform of DNA sequence analyzers within its Production Capillaries group: seventy Applied Biosystems 3730xl and thirty-six GE Healthcare MegaBACE 4500 instruments. The Capillaries group is comprised of eleven employees that are responsible for operating and maintaining both platforms; the group is also involved in the daily monitoring of performance stats and the troubleshooting of DNA sample and instrument related issues. The operation of these high-throughput fluorescence-based DNA sequence analyzers will be assessed on the strengths and benefits of each platform, including instrument overviews of operational parameters and mechanical/component specifications. In addition, instrument setups for production operation, operation schedules, loading, and maintenance strategies as well as the various sequencing strategies for each platform

will be compared. Throughput numbers and sequencing quality results will be presented.

This work was performed under the auspices of the U.S. Department of Energy's Office of Science, Biological and Environmental Research Program, and by the University of California, Lawrence Livermore National Laboratory under Contract No. W-7405-Eng-48, Lawrence Berkeley National Laboratory under contract No. DE-AC02-05CH11231 and Los Alamos National Laboratory under contract No. W-7405-ENG-36.

## 18. Phylogenomic Analyses of Cytoskeletal Genes in the Free-Living Basal Eukaryotes *Spironucleus vortens* and *Naegleria gruberi*

**Scott Dawson\*** (sdawson@nature.berkeley.edu), **Lillian Fritz-Laylin\*** (fritz-laylin@berkeley.edu), and W. Z. Cande

Molecular and Cell Biology, University of California, Berkeley, CA

Molecular phylogenetic analyses indicate that the majority of the over seventy proposed "kingdom-level" lineages of eukaryotes are actually microbial, and understanding the origins and evolution of the eukaryotes requires genomic data from diverse eukaryotic microbes. Most of our knowledge of biological processes, such as the cell cycle and cell division, and their functional diversity derive from studies of more recently evolved, and phylogenetically-limited, macroscopic eukaryotic model organisms such as fungi and metazoans. For obvious medical reasons, most basal microbial eukaryotes currently studied are parasites (e.g. Trypanosoma brucei, Giardia intestinalis). This is our rationale for genome sequencing of a free-living diplomonad, Spironucleus vortens, and a heterolobosean, Naegleria gruberi. Using pulsed-field gel electrophoresis, we estimated the genome size of *Spironucleus vortens* to be 18Mb, and *Naegleria gruberi* to be about 40Mb. Additionally, as compared to Giardia, an intestinal parasite, we have identified more complete biosynthetic pathways of vitamins and amino acids in Spironucleus – supporting the notion that Giardia's gene losses in the adaptation to parasitism are mainly metabolic. In terms of the cytoskeleton, the microtubule system in *Spironucleus* appears to contain a complete array of component proteins, including many all tubulin isoforms, and many kinesins and dyneins. Yet in stark contrast, the actin or microfilament system is missing the majority of known proteins other than actin itself. Specifically, we find no evidence for myosin, arps 1, 2, 3, profilin, cofilin, or numerous proteins involved in actin assembly, disassembly and stability. Thus our analyses of two diplomonads – the free-living Spironucleus and the parasitic Giardia – indicates a lack of actin-associated proteins, implying a later evolution of an elaborated actin cytoskeleton. Further, although Naegleria has some cytoskeletal components, there is far less than in other more recently evolved amoebae like Dictyostelium. Additional analyses will allow us to identify core features of the eukaryotic cytoskeleton in Spironucleus and Naegleria.

### 19. Two Rounds of Whole Genome Duplication in the Ancestral Vertebrate Genome

**Paramvir S. Dehal\*** (psdehal@lbl.gov)<sup>1</sup> and Jeffrey L. Boore<sup>1,2</sup>
<sup>1</sup>DOE Joint Genome Institute, Lawrence Berkeley National Laboratory, Walnut Creek, CA and <sup>2</sup>Dept. of Integrative Biology, University of California, Berkeley, CA

The hypothesis that the relatively large and complex vertebrate genome was created by two ancient, whole genome duplications has been hotly debated, but remains unresolved. We reconstructed the evolutionary relationships of all gene families from the complete gene sets of a tunicate. fish, mouse, and human, then determined when each gene duplicated relative to the evolutionary tree of the organisms. We confirmed the results of earlier studies that there remains little signal of these events in numbers of duplicated genes, gene tree topology, or the number of genes per multigene family. However, when we plotted the genomic map positions of only the subset of paralogous genes that were duplicated prior to the fish-tetrapod split, their global physical organization provides unmistakable evidence of two distinct genome duplication events early in vertebrate evolution indicated by clear patterns of 4-way paralogous regions covering a large part of the human genome. Our results highlight the potential for these large-scale genomic events to have driven the evolutionary success of the vertebrate lineage.

## 20. "PhIGs" (Phylogenetically Inferred Groups): A Tool for Whole Genome Phylogenetic Analysis

**Paramvir Dehal**<sup>1</sup> and **Jeffrey L. Boore**\*<sup>1,2</sup> (jlboore@berkeley.edu)

<sup>1</sup>DOE Joint Genome Institute, Lawrence Berkeley National Laboratory, Walnut Creek, CA, and <sup>1,2</sup>Dept. of Integrative Biology, University of California, Berkeley, CA

We present the PhIGs database, a phylogenomic resource for sequenced genomes. Although many methods exist for clustering gene families, very few attempt to create truly orthologous clusters sharing descent from a single ancestral gene across a range of evolutionary depths. Although these non-phylogenetic gene family clusters have been used broadly for gene annotation, errors are known to be introduced by the artifactual association of slowly evolving paralogs and lack of annotation for those more rapidly evolving. PhIGS automates the clustering of genes into families with a method that respects the known evolutionary relationships among the organisms, creation of evolutionary trees for each cluster using maximum likelihood, the most sophisticated method of evolutionary reconstruction, determination of orthologous and paralogous relationships, and the correlation of this with information on gene function and relative position. We plan further work to link other information, such as gene expression patterns.

The PhIGs database (http://phigs.org) currently contains 23 completely sequenced eukaryotic genomes of fungi and metazoans, containing 409,653 genes that have been grouped into 42,645 gene clusters. Clusters can be searched using keywords on annotations, such as GO and InterPro assignments, and by sequence similarity using BLAST and HMM. Results include graphics that show the evolutionary relationships of all genes within each gene family, color-coded multiple sequence alignments, comparisons of exon-intron structures, links to databases on gene function, and multi-species maps showing the relative positions of sets of large numbers of orthologous genes.

#### 21. The Genome of Halorhodospira halophila

Ratnakar H. Deole\* and Wouter D. Hoff\*

Oklahoma State University, Stillwater, OK

#### 22. High-Density Universal 16S rRNA Microarray Analysis Reveals Broader Diversity than Typical Clone Library When Sampling the Environment

**Todd DeSantis**, **Eoin Brodie\*** (elbrodie@lbl.gov), Jordan Moberg, Ingrid Zubietta, Yvette Piceno, and Gary Andersen

Molecular Microbial Ecology Group, Ecology Department, Earth Sciences Division, Lawrence Berkeley National Laboratory, Berkeley, CA

Molecular approaches aimed at broad prokaryotic environmental detection routinely rely upon classifying heterogeneous 16S rRNA genes amplified by universal PCR. The general method of sampling DNA types has been to clone and sequence the PCR products. However, the number of clones required to adequately catalogue the majority of taxa in a sample is unwieldy. Alternatively, hybridizing target sequences to a universal 16S rRNA microarray may provide a more rapid and comprehensive view of prokaryotic community composition. This study investigated the breadth and accuracy of a microarray in detecting diverse 16S rRNA gene sequence types compared to clone-and-sequencing using three environmental samples: urban aerosol, subsurface soil and subsurface water. PCR products generated from universal 16S primers were classified using either the clone-and-sequence method or by hybridization to a novel high-density microarray of 297,851 probes complementary to 842 prokaryotic sub-families. The three clone libraries comprised 1,391 highquality sequences. Approximately 8% of the clones could not be placed into a known sub-family and were considered novel. The microarray results confirmed the majority of clone-detected sub-families and additionally demonstrated greater amplicon diversity extending into phyla missed by the cloning method. Sequences matching OTUs within the phyla Nitrospira, Plantomycetes, and TM7, which were uniquely detected

#### **Poster Presentations**

by the array, were verified with specific primers and subsequent amplicon sequencing. Clone-observed richness appeared to be an underestimate when compared to the non-parametric richness predictions, however array-observed richness corresponded well. It was concluded that although the microarray is unreliable in classifying novel prokaryotic taxa, it is more adept at identifying greater diversity in environmental samples than sequencing a typically-sized clone library. Furthermore, the microarray allowed samples to be rapidly evaluated with replication, a significant advantage in studies of microbial ecology.

## 23. A Multiple Sequence Alignment Server for Comparative Analysis of 16S rRNA

**Todd DeSantis**, <sup>1,4</sup> Phil Hugenholtz, <sup>2</sup> Keith Keller, <sup>5,4</sup> Eoin Brodie, <sup>1</sup> Niels Larsen, <sup>3</sup> **Yvette Piceno\*** (ympiceno@lbl.gov), <sup>1</sup> Richard Phan, <sup>1,4</sup> and Gary Andersen <sup>1,4</sup>

<sup>1</sup>Center for Environmental Biotechnology, Lawrence Berkeley National Laboratory, Berkeley, CA; <sup>2</sup>Microbial Ecology Program, DOE Joint Genome Institute, Walnut Creek, CA; <sup>3</sup>Danish Genome Institute, Aarhus, Denmark; <sup>4</sup>Virtual Institute for Microbial Stress and Survival, Lawrence Berkeley National Laboratory, Berkeley, CA; and <sup>5</sup>Quantitative Biomedical Research, University of California, Berkeley, CA

Microbiologists conducting surveys of bacterial and archaeal diversity often require comparative alignments of thousands of 16S rRNA genes collected from a sample. The computational resources and bioinformatics expertise required to construct such an alignment has inhibited highthroughput analysis. It was hypothesized that an online tool could be developed to efficiently align thousands of 16S rRNA genes via the NAST (Nearest Alignment Space Termination) algorithm for creating multiple sequence alignments (MSA). The tool was implemented with a webinterface at http://greengenes.lbl.gov/cgi-bin/nph-NAST\_align.cgi. Each user-submitted sequence is compared to Greengenes' "Core Set", comprising approximately 10,000 aligned non-chimeric sequences representative of the currently recognized diversity among bacteria and archaea. User sequences are oriented and paired with their closest match in the Core Set to serve as a template for inserting gap characters. Non-16S data (sequence from vector or surrounding genomic regions) is conveniently removed in the returned alignment. From the resulting MSA, distance matrices can be calculated for diversity estimates and organisms can be classified by taxonomy. The ability to align and categorize large sequence sets using a simple interface has enabled researchers with various experience levels to obtain prokaryotic community profiles.

## 24. Coral Reef Genomics: A Genome-Wide Approach to the Study of Coral Symbiosis

**Michael DeSalvo\*** (mdesalvo@ucmerced.edu), <sup>1</sup> Jodi Schwarz, <sup>2</sup> Peter Brokstein, <sup>2</sup> Cindy Lewis, <sup>3</sup> Shruti Lal, <sup>1</sup> Constance Rogers-Lowery, <sup>4</sup> Chitra Manohar, <sup>5</sup> Carol Tang, <sup>6</sup> Alina Szmant, <sup>4</sup> Mary Alice Coffroth, <sup>3</sup> and Mónica Medina <sup>1</sup>

<sup>1</sup>University of California, Merced, CA; <sup>2</sup>DOE Joint Genome Institute, Walnut Creek, CA; <sup>3</sup>State University of New York, Buffalo, NY; <sup>4</sup>University of North Carolina, Wilmington, NC; <sup>5</sup>Lawrence Livermore National Laboratory, Livermore, CA; and <sup>6</sup>California Academy of Sciences, San Francisco, CA

Coral reefs are among the most beautiful and diverse ecosystems on Earth, upon which 500 million people depend for food, coastal protection, and other resources. Yet coral reefs are threatened by the declining health of the marine environment: coral bleaching and coral disease are causing the world's reefs to suffer drastic declines in coral cover. At the heart of the reef ecosystem is a symbiosis between coral hosts and endosymbiotic dinoflagellates (Symbiodinium spp.). The importance of this symbiosis to reef-building corals and reef nutrient and carbon cycles is well documented, but little is known about the cellular and molecular mechanisms by which the partners establish and regulate the symbiosis. We have developed cDNA libraries for different life stages of the Caribbean corals *Montastraea faveolata* and *Acropora palmata*, and these libraries have been used to print cDNA microarrays. cDNA libraries and gene expression microarrays are also being developed for two Symbiodinium strains identified to infect the two coral hosts. By experimentally infecting aposymbiotic coral larvae with dinoflagellates, we will see gene expression changes that are correlated with the onset and maintenance of the symbiosis. These observations will provide new knowledge and avenues of research to help protect these endangered organisms. We have also started to design experiments to address: 1) the molecular basis of coral bleaching; 2) the mechanisms of calcification in corals; 3) the molecular basis of coral diseases; 4) speciation genes in Montastraea and Acropora Caribbean species and their hybrids; and 5) differences in gene expression within and between populations.

## 25. The Genome Sequence of *Alkaliphilus metallireducens*, an Alkaliphilic, Metal-Reducing Bacterium

**M. W. Fields\*** (fieldsmw@muohio.edu), <sup>1</sup> J. Zhou, <sup>2</sup> A. Lapidus, <sup>3</sup> C. S. Han, <sup>4</sup> O. Chertkov, <sup>4</sup> and P. M. Richardson <sup>3</sup>

<sup>1</sup>Dept. of Microbiology, Miami University, Oxford, OH; <sup>2</sup>Institute for Environmental Genomics, University of Oklahoma, Norman, OK; <sup>3</sup>Genomics Facility, DOE Joint Genome Institute, Walnut Creek, CA; <sup>4</sup>DOE-Joint Genome Institute, Los Alamos National Laboratory, Los Alamos, NM

Alkaliphilus metalliredigens is an alkaliphilic, iron- and metal-reducing bacterium that was isolated from borate leachate ponds, and the draft genome sequence is now available. A. metalliredigens is a low G+C Gram-positive bacterium and is distantly related to other metal-reducing bacteria. The genome is approximately 4.4 Mb in size, and has a 36.6% G+C content. The genome is predicted to contain 4,090 protein-coding ORFs, and approximately 30% do not have function prediction. In the laboratory, growth is supported only in the presence of a metal as a terminal electron acceptor; however, the genome contained few ORFs annotated as cytochromes. Only four flavocytochromes were identified as possible fumarate reductase subunits. An ammonia-forming nitrite reductase and a nitric oxide reductase were predicted, and attempts to cultivate the organism on nitrate or nitrite were not successful. Multiple Ftype, V-type, and P-type ATPases were predicted, and the organism is moderately halophilic and was isolated from a site that contained multiple heavy metals. Genes annotated as pyruvate dehydrogenase, pyruvateferredoxin oxidoreductase, and pyruvate-formate lyase were observed. In relation to intermediary metabolism, an incomplete glycolysis, an incomplete reductive TCA, and a complete pentose phosphate pathway were detected. Annotated genes involved in butanoate production and possibly propanoate production were observed. Annotated genes for mannitol and fructose-specific PTS genes were observed, and mechanisms for the utilization of mannitol, fructose, sorbose, sorbitol, melibiose, and mannose could be predicted. In addition, the genome was predicted to contain two polysaccharide-degrading enzymes: arabinanase and mannanase. Almost 50 genes were predicted to be involved in sporulation, and spore-formation was detected. A ferrichrome system (enterochelin) was predicted, both the binding protein and permease. In accordance with different metabolisms and dynamic environment, A. metallireducens was predicted to have approximately 25 ORFs with PAS domains, 7 of which also contained HTH motifs.

## **26.** Phylogenomics with Single Copy Universal Genes (SCUGs)

**M. Pilar Francino\*** (mpfrancino@lbl.gov), Monica Medina, Jenna Morgan, and Paramvir Dehal

<sup>1</sup>Evolutionary Genomics Department, DOE Joint Genome Institute, Walnut Creek, CA, and <sup>2</sup>School of Natural Sciences, University of California, Merced, CA

The duplicability, or tendency of a gene family to proliferate within a genome, is very variable. Some genes, mostly those involved in information processing and other complex mechanisms requiring many molecular interactions, are nearly universally found in single copy across all genomes, others tend to form small gene families, and a few appear greatly duplicated in many organisms. We suggest that the set of Single Copy Universal Genes (SCUGs), or genes present in all organisms of a given clade in single copy, is likely to be the most reliable set of genes for phylogenetic purposes.

The utilization of SCUGS will avoid several of the basic problems commonly confounding phylogenetic reconstruction. In addition to the obvious advantage of avoiding hidden paralogy, SCUGs generally evolve at slower rates and are less prone to horizontal gene transfer. Also, genes whose histories involve bursts of duplication in different lineages would be more variable in evolutionary rate through time and across species, because gene duplication is most often followed by accelerations of evolutionary rate in some of the resulting duplicates.

Here we present phylogenetic analyses based on SCUGs for two deep clades of the tree of life: the Bacteria and the Opisthokonta, the eukaryotic lineage that contains animals and fungi. We show that substantial levels of phylogenetic resolution can be obtained for each clade by concatenating alignments of their respective SCUGs. We also present a comparative analysis of the gene constituency of the two SCUG sets, and a study of phylogenetic congruence among different genes.

## 27. Over and Under-Representation of Sigma-70 Binding Sites in Different Bacterial Genomic Regions

**Jeff Froula\*** (jlfroula@lbl.gov) and M. P. Francino Evolutionary Genomics, DOE Joint Genome Institute, Walnut Creek, CA

The  $\sigma^{70}$  subunit of the bacterial RNA polymerase holoenzyme is responsible for specific binding to the -10 and -35 promoter motifs. It has been shown that selection acts to remove -10 and -35 consensus sequences in both coding and noncoding regions, implying that it is disadvantageous to maintain misplaced sites that can strongly bind  $\sigma^{70}$  and interfere with proper gene expression. Here we analyze 47 bacterial

genomes and show that the numbers of potential  $\sigma^{70}$  binding sites in noncoding regions deviate significantly from the random expectations based on base composition, di- and tri-nucleotide contents in a majority of eubacteria. This implies that not only is there selection for maintaining high densities of potential  $\sigma^{70}$  binding sites signals in *regulatory* DNA, there is also selection against these sites in *non-regulatory* DNA. This result suggests that these often overlapping binding sites confer some subtle survival advantage although experimental evidence has shown only one or two of these sites are actually transcription initiation sites. Additional results showing that faster growing bacteria have more selection against potential  $\sigma^{70}$  binding sites in non-regulatory DNA is evidence that the efficiency needed for faster growing bacteria can only be achieved by reducing spurious RNA polymerase binding to false sites.

#### 28. Cloning the Alvinella pompejana XPD Homolog

**Jill O. Fuss\*** (jfuss@lbl.gov), David S. Shin, Quen J. Cheng, S. Craig Cary, and John A. Tainer<sup>1,2</sup>

<sup>1</sup>Lawrence Berkeley National Laboratory, Berkeley, CA; <sup>2</sup>Scripps Research Institute, La Jolla, CA; and <sup>3</sup>University of Delaware, Lewes, DE

Alvinella pompejana are one of the earth's most thermophilic multicellular eukaryotes. These polychaete annelids, or segmented bristle worms, live in hydrothermal vents 2500 meters below the surface of the Pacific Ocean, where they thrive in an environment of high temperature (20-80°C), low pH, and rich in heavy metals, carbon dioxide, and hydrogen sulfide. Despite living in an environment that is vastly different from our own, our initial analyses of EST sequences generated by the JGI suggest that A. pompejana and human genes share high homology. One such example is the A. pompejana homolog of the human XPD gene.

XPD is a 5'-3' helicase that is required for the efficient repair of damaged DNA. Mutations in the XPD gene result in three distinct hereditary diseases: Xeroderma pigmentosum (XP), Cockayne's syndrome (CS) and Trichothiodystrophy (TTD). XP is characterized by extreme sunsensitivity and susceptibility to skin cancer due to inability to repair UV and chemical DNA damage. CS is a post-natal developmental disorder characterized by severe progressive neurological deterioration. TTD patients have severe neurological deterioration and developmental abnormalities that are distinct from similar symptoms in CS. The spectrum of XPD mutations sheds little light as to the mechanistic cause of XP, CS, or TTD. Disease causing point mutations are spread throughout the length of the protein and even adjacent mutations can give rise to different phenotypes. Efforts to understand XPD structural biology have been hindered by difficulties in expressing recombinant human XPD in sufficient quantities for structural analysis. In addition, XPD has been difficult to model in thermophilic archaea due to poor sequence conservation.

Using EST sequences generated by our FY2005 JGI collaboration, we have cloned the *A. pompejana* homolog of human XPD. A BLAST search of *A. pompejana* ESTs revealed a 700 bp fragment with high homology to the human XPD protein sequence. Using primers complementary to this fragment, we amplified specific 5' and 3' RACE products that spanned the length of the *A. pompejana* XPD gene. The protein sequence of ApXPD is highly conserved to human XPD: 75% identity and 89% similarity. Most importantly for understanding human disease, residues that are mutated in human disease are largely conserved in ApXPD.

Understanding the biology of *A. pompejana* will be important not only for understanding human disease, but will also provide insight into how eukaryotes survive in a toxic environment of high temperature, carbon dioxide, and heavy metal stresses.

## 29. An Overview of the Sequencing Prep Process at the DOE Joint Genome Institute's Production Genomics Facility

**Tijana Glavina del Rio\*** (glavinadelrio1@llnl.gov), Arshi Khan, Megan Kennedy, Zalak Trivedi, Michael Philips, David Robinson, Terri Jackson, and Michael Pintor

DOE Joint Genome Institute, Walnut Creek, CA

The sequencing prep process is the second step of the production process at the DOE Joint Genome Institute (JGI) Production Genomics Facility (PGF). The goal of the process is to prepare labeled fragments in 384-well plates for loading onto the capillary sequencers. The process begins by alliquoting 2µl of glycerol stock from the Library Support Process using Matrix PlateMate Plus. The 384-well plates, containing glycerol stock and buffer, are placed on the PE 9700 thermocyclers and heated to 95°C for 5 minutes in order to lyse the cells and release the plasmids. The amplification of plasmid DNA is performed using Templiphi, a kit made by GE Healthcare in order to generate large quantities of template. TempliPhi is added with a multidrop micro instrument and samples are incubated at 30°C for 20 hours to allow the amplification to occur. The amplified template is then heated to 95°C to inactivate the enzyme. Small amounts of amplified DNA are transferred using a Robbins Hydra Twister robot into two 384-well plates to set up dual sequencing reactions. The Chemistry cocktail is added by the Cavro Dispensing instrument and then cycled. For high GC templates we utilize a 5% final concentration of DMSO that is added to our TempliPhi and chemistry reactions. Before DNA can be loaded onto the capillary sequencer, the leftover reagents, cell debris, buffers, and salts must be removed from the sample. The last step in the Sequencing prep process is the cleanup step. The process uses a modified magnetic bead protocol to purify DNA fragments from the sequencing reaction and get it ready for sequencing. This step is performed using the Beckman Coulter Biomek FX robots. The reactions

are then ready to load on either the ABI 3730xl or the MegaBACE 4500 for capillary electrophoresis. This poster will present a detailed overview of the sequencing prep process reviewing each step and the instruments that are used.

#### 30. Annotation of Eukaryotic Genomes at JGI

**Igor V. Grigoriev\*** (ivgrigoriev@lbl.gov)

DOE Joint Genome Institute, Walnut Creek, CA

#### 31. Automated Microbial Genome Annotation

**Loren J. Hauser\*** (hauserlj@ornl.gov)
Oak Ridge National Laboratory, Oak Ridge, TN

## 32. The Genome Sequences of the Ethanol Producing Bacteria *Thermoanaerobacter ethanolicus* 39E and *Thermoanaerobacter ethanolicus* X514

**C. L. Hemme\*** (hemmecl@ou.edu), <sup>1,2</sup> M. W. Fields, <sup>3</sup> E. H. Saunders, <sup>4</sup> M. L. Land, <sup>1</sup> L. J. Hauser, <sup>1</sup> A. L. Lapidus, <sup>5</sup> C. S. Han, <sup>4</sup> D. C. Bruce, <sup>4</sup> P. Richardson, <sup>5</sup> and J. Zhou<sup>2</sup>

<sup>1</sup>Oak Ridge National Laboratory, Oak Ridge, TN; <sup>2</sup>The Institute for Environmental Genomics, University of Oklahoma, Norman, OK; <sup>3</sup>Miami University, Oxford, OH; <sup>4</sup>Los Alamos National Laboratory, Los Alamos, NM; and <sup>5</sup>DOE Joint Genome Institute, Walnut Creek, CA

Recent volatility in the global energy markets has sparked renewed interest in the development of alternative fuel technologies to reduce American reliance on fossil fuels. One promising technology that has generated much interest is the generation of bio-produced ethanol from cellulosic materials such as switch grass. To this end, the genomes of two strains of the thermophilic ethanol-producing bacteria Thermoanaerobacter ethanolicus have been sequenced. Strain 39E was originally isolated from thermal springs in Yellowstone National Park while the metal-reducing strain X514 was obtained from a deep subsurface location geographically isolated for ~200 mya. The preliminary annotations for these two strains were compared to the annotation for the completed genome of T. tengcongensis. All species studied encode systems for the breakdown of cellulose and glycogen. The two T. ethanolicus strains also encode systems for the uptake and fermentation of xylose and other pentose sugars derived from xylan and hemicellulose. A β-xylosidase gene implicated in degradation of xylan and hemicellulose was previously cloned from T. ethanolicus JW200 and this gene was found to be highly conserved in strain 39E but significantly diverged in

strain X514. The *T. ethanolicus* strains also encode components of the Entner-Doudroff pathway which are lacking in *T. tengcongensis*. Interestingly, strain X514 lacks a methylglyoxal shunt that is present in the other two *Thermoanaerobacter* species. In many fermenting bacteria, the methylglyoxal shunt is employed to limit energy production when carbohydrates are plentiful. Thus, the loss of the shunt in X514 may represent an adaptation of the strain to the presumably energy-poor conditions of the deep subsurface. The preliminary results suggest that all three organisms utilize similar carbon metabolism pathways but that differences in the carbon metabolism and energy flux pathways may have evolved in response to particular environmental pressures.

### **33.** Fish *Hox* Cluster and the Evolution of Regulatory Elements

**Simone Hoegg**, <sup>1</sup> Jeffrey L. Boore, <sup>2</sup> and **Axel Meyer\*** <sup>1</sup> (axel.meyer@unikonstanz.de)

<sup>1</sup>Dept. of Biology, LS Evolutionary Biology, University of Konstanz, Konstanz, Germany and <sup>2</sup>Evolutionary Genomics Program, DOE Joint Genome Institute, Walnut Creek, CA

While tetrapods, as human, mouse and frog (*Xenopus tropicalis*), have a constant number of four Hox clusters, teleost fish have undergone an lineage-specific genome duplication event and, subsequently, experienced gene and cluster losses. Based on current genomic projects, we know the Hox cluster composition of four teleost fish, *Danio rerio* (zebrafish), *Oryzias latipes* (medaka), *Takifugu rubripes* (torafugu) and *Tetraodon nigroviridis* (spotted green pufferfish). All these species possess seven clusters; however, zebrafish has lost the *HoxDb* cluster, while the other species described have lost the *HoxCb* cluster. From this we can conclude that the loss of these clusters happened independently in the Ostariophysii, to which *Danio rerio* belongs and in the Neoteleostei, which are represented by the other three species. Since Hox gene setup in ray-finned fish show much higher variability than in tetrapods further studies for a denser sampling are required.

With ongoing sequencing projects and the development of large insert size libraries (BAC, PAC), research on Hox clusters has moved from counting genes to comparative analyses of the non-coding proportions of these clusters. Hox cluster provide an ideal genomic model region for research on conserved non-coding elements, since the gene clusters are highly condensed, contain relatively few repetitive elements (e.g. SINEs, LINEs, LTRs [long terminal repeats] and DNA transposons) and are conserved over a large evolutionary scale. Therefore, homologous DNA stretches are easy to identify between species and gain and loss of conserved non-coding elements can be traced easily on a phylogeny. Also the content of conserved non-coding sequences seems to be especially high in this genomic region.

#### **Poster Presentations**

Ongoing research projects on e.g. bichir (*Polypterus senegalus*) and coelacanth (*Latimeria menadoensis*) are providing data for basal lineages for which data usually is difficult to obtain. Another sampling gap so far is the order Perciformes, the largest order of ray-finned fish with approximately 10,000 species in 150 families and morphologically highly diverse. So far, coding sequences for *Hox* genes have been described and the complete *HoxAa* cluster for *Oreochromis niloticus* (tilapia). A BAC (Bacterial Artificial Chromosome) library of *Astatotilapia burtoni*, a haplochromine cichlid with a basal position relative to the Lake Victoria radiation, has been constructed in our lab. This enables us to sequence the complete *Hox* clusters. East African cichlids with their high morphological diversity and low genetic distance provide a fascinating study field for possible influence of regulatory evolution on morphology.

#### 34. Genomic Sequencing of the Wood-Rotting Mushroom Schizophyllum commune

Stephen Horton\* (hortons@union.edu), <sup>1</sup> Tom Fowler\* (tfowler@siue.edu), <sup>2</sup> Vanessa Bailey, <sup>3</sup> Scott Baker, <sup>3</sup> Allen Gathman, <sup>4</sup> Erika Kothe, <sup>5</sup> Walt Lilly, <sup>4</sup> Jon Magnuson, <sup>3</sup> and Han Wosten <sup>6</sup> <sup>1</sup>Union College, Schenectady, NY; <sup>2</sup>Southern Illinois University, Edwardsville, IL; <sup>3</sup>Pacific Northwest National Laboratory, Richland, WA; <sup>4</sup>Southeast Missouri State University, Cape Girardeau, MO; <sup>5</sup>Friedrich-Schiller University, Jena, Germany; and <sup>6</sup>Utrecht University, Untrecht, The Netherlands

The filamentous fungus *Schizophyllum commune* belongs to the group of gilled mushrooms that includes the commercially valuable species *Agaricus bisporus* (white button mushroom) and *Pleurotus ostreatus* (oyster mushroom). The worldwide market for these and other mushrooms for use as food, as dietary supplements and nutriceuticals, and in medicine was estimated to be over U.S. \$13 billion per year in 1995. Mushrooms from many of these fungi are also the source of important medicinal compounds with activities against tumors, viruses, and other microbes.

S. commune has been utilized extensively as a model system for studying mating-type gene function and mushroom development. It is a genetically tractable system that has well-developed methodologies for both targeted and random gene transformation. The S. commune genome is about 36-38 Mb in size, with a G+C content of 57%. 14 chromosomes have been discerned from CHEF gel/hybridization experiments, ranging in size from 1.6 Mbp to 4.7 Mbp. Taken together, these molecular characteristics make S. commune an excellent sequencing target. The ability of S. commune to degrade complex plant biomass, including lignin, also makes the organism attractive for the DOE-JGI genomic sequencing program. We expect that its' genome will yield an interesting catalog of secreted enzymes that play a role in ligno-cellulose degradation. The organism has demonstrated bioremediation potential with respect to heavy metal uptake. A genome

sequence, combined with already established molecular techniques, would make *S. commune* an ideal model organism for additional studies in metal uptake by fungi.

We expect that the availability of the *S. commune* genome will attract considerable interest in the fungal research community. Genomic comparisons between wood-rotters *S. commune* and *Phanerochaete chrysosporium*, and the dung-fungus *Coprinopsis cinerea* would potentially allow identification of specializations for wood-rotting, as distinct from general saprophytic mechanisms. The comparison of two distinct model organisms for the study of mushroom development (*S. commune* and *C. cinerea*) should distinguish elements common to this process apart from those elements that are species-specific. The *S. commune* genome sequence, in conjunction with these other fungal genomes and the wealth of genetic information about *S. commune*, will be a valuable resource to promote continued and vigorous research in mushroom biology.

### 35. *Pichia stipitis*: A Native Xylose Fermenting Yeast that Inhabits Beetle Gut

**Thomas W. Jeffries\*** (twjeffri@wisc.edu)<sup>1</sup>, Igor Grigoriev,<sup>2</sup> Jane Grimwood,<sup>3</sup> Andrea Aerts,<sup>2</sup> Asaf Salamov,<sup>2</sup> Erika Lindquist,<sup>2</sup> Yong-Su Jin,<sup>4</sup> Volkmar Passoth,<sup>5</sup> José M. Laplaza,<sup>6</sup> and Paul Richardson<sup>2</sup>

<sup>1</sup>USDA, Forest Service, Forest Products Laboratory and University of Wisconsin, Madison, WI; <sup>2</sup>DOE Joint Genome Institute, Walnut Creek, CA; <sup>3</sup>Stanford Human Genome Center, Palo Alto, CA; <sup>4</sup>Dept. of Food Science and Biotechnology, Sungkyunkwan University, Suwon, Korea; <sup>5</sup>Dept. of Microbiology, Swedish University of Agricultural Sciences (SLU), Uppsala, Sweden; and <sup>6</sup>BioTechnology Development Center, Cargill, Minneapolis, MN

Pichia stipitis Pignal (1967) is an predominantly haploid, heterothallic yeast related to Candida shehatae and other pentose metabolizing ascomycetous species. It belongs to a clade that uses an alternative nuclear genetic code in which CUG codes for serine rather than leucine. Its closest relatives are found in the intestines of passalid beetles, which in turn are found grazing on white-rotted hardwood. P. stipitis has the highest native capacity for xylose fermentation of any known microbe. Strains of P. stipitis are among the best xylose-fermenting yeasts in type culture collections. Fed batch cultures of P. stipitis produce up to 47 g/L of ethanol from xylose at 30°C under low aeration conditions. In addition to fermenting D-xylose to ethanol, P. stipitis can assimilate cellobiose and will oxidize the lignin-related compounds veratraldehyde and vanillin to their respective alcohols and acids. As such it is an ideal organism for lignocellulose bioconversion.

#### **Poster Presentations**

The entire 15.44 Mbp of the *P. stipitis* genome was shotgun sequenced, finished to high quality, and annotated. *P. stipitis* has 8 chromosomes ranging from 3.5 to 0.97 mbp. The genome assembly was annotated using the JGI Annotation Pipeline. *P. stipitis* release v2.0 includes 5841 gene models supported by EST, cDNA and protein evidence. Average gene, transcript and CDS lengths are 1.6 kb, 1.5 kb and 493 amino acids, respectively. Average gene density is 56% with 4204 single exon genes. The genome size, number of genes and CDS lengths are comparable to the numbers found in other sequenced yeast genomes. A set of 19,635 ESTs was sequenced and clustered: 3839 (94%) of the clusters mapped to the genome; 2252 (40%) genes were supported by ESTs. An absolute majority of predicted genes were supported by protein homology including 4879 (84%) with strong homology in other fungi. 4083 (70%) of all predicted genes have a predicted protein domain.

More than 2600 genes have been manually curated using the JGI Genome Portal Tools, and there appear to be major differences between P. stipitis and other yeasts in oxidative phosphorylation, fatty acid metabolism and fatty acid synthesis. The sequenced genome (CBS 6054) includes seven family 3  $\beta$ -glucosidases, a family 10 endo xylanase, one  $\beta$ -mannanase, two exo-1,3  $\beta$ -glucanases, five cinnamyl alcohol dehydrogenases and 57 transporters in the major facilitator superfamily – including 5 putative xylose transporters. Aside from genes for assimilating a wide variety of lignocellulosic polymers, P. stipitis codes for several proteins that enable it to ferment xylose with the production of very little xylitol. These include four NADP-dependent alcohol dehyrogenases, two primary alcohol dehydrogenases and three pyruvate decarboxylases.

In preparing ESTs for annotation, cells were grown on glucose or xylose under aerobic or oxygen limited conditions. Each of these four pools was clustered separately and the resulting hit frequencies provided estimates of gene expression.

### **36.** Comparative Genomics of Two Biomedically Important Marine Actinomycetes

**Paul R. Jensen\*** (pjensen@ucsd.edu) and Bradley S. Moore Scripps Institution of Oceanography, Center for Marine Biotechnology and Biomedicine, University of California, San Diego, La Jolla, CA

Actinomycetes are the single most significant source of microbial natural products, accounting for more than one half of all known antibiotics. We have recently discovered a new actinomycete genus (*Salinispora*) whose members are proving to be an important source of novel secondary metabolites including a new treatment for cancer. These actinomycetes reside in marine sediments and display fundamental physiological differences from those that occur on land including an obligate requirement of seawater for growth. JGI is in the process of sequencing

the type strains for the species *S. tropica* and *S. arenicola*. These will be the first marine actinomycetes and, more generally, the first marine Grampositive bacteria to be sequenced thus providing important new information about an environmentally significant yet poorly studied group of prokaryotes. These two species produce completely different suites of secondary metabolites with a preliminary evaluation of the *S. tropica* draft sequence indicating that as much as 10% of the genome is devoted to secondary metabolite biosynthesis. This includes pathways for the clinically relevant enediyne antitumor agents and salinosporamide A, a potent proteasome inhibitor. The genome sequences will advance our understanding of how secondary metabolites are produced and provide new information about the evolutionary significance of secondary metabolite production.

#### 37. Functional Metagenomics of Methylotrophy

**Marina G. Kalyuzhnaya\*** (mkalyuzh@u.washington.edu), David C. Bruce, Kerrie W. Barry, Paul M. Richardson, Mary E. Lidstrom, and Ludmila Chistoserdova<sup>1</sup>

<sup>1</sup>University of Washington, Seattle, WA; <sup>2</sup>DOE Joint Genome Institute, Los Alamos National Laboratory, Los Alamos, NM; and <sup>3</sup>DOE Joint Genome Institute, Production Genomics Facility, Walnut Creek, CA

Microorganisms form a foundation of Earth's biosphere and are key drivers of global biogeochemical processes. Therefore understanding, monitoring and prediction of environmental changes require comprehensive studies of microbial communities that lay beyond the reach of tradition microbiology. Rapid advances in modern molecular methods and high-throughput genomics, such as the whole genome shotgun (WGS) community sequencing approach, open new ways to study microbial ecology. However, WGS sequencing often results in low sequence coverage, not allowing for sequence assembly, due the high complexity and/or dynamics of natural communities. Here we present a modified WGS sequencing approach (functional metagenomics) that should allow for higher sequence coverage, and potentially for assembling complete genomes from environmental samples. In this approach, a chosen functional group is specifically enriched for, to reduce the community complexity. To test this approach, we focused on the methylotroph community in Lake Washington sediment and employed Stable Isotope Probing (SIP) for enrichment, followed by WGS sequencing. Sediment samples were exposed separately to five different <sup>13</sup>C-labeled C<sub>1</sub> compounds: methane, methanol, methylamine, formate and formaldehyde. Total DNA was extracted from each microcosm, and the <sup>13</sup>C-labeled fractions were separated from unlabeled DNA by isopycnic centrifugation. <sup>13</sup>C-labeled DNA was used to construct five separate shotgun libraries, and these were sequenced at the JGI facility. 42 to 89 Mb of sequence was produced from each microcosm, totaling 372 Mb. These were assembled, and preliminary analysis of community richness was conducted. First

#### **Poster Presentations**

genomic insights into a broad variety of bacterial groups actively metabolizing  $C_1$  compounds in Lake Washington were obtained, and specific shifts in populations in response to specific  $C_1$  compounds were detected. In each microcosm, a finite number of phylotypes were detected, and some of these were present in more than one type of enrichment. Those enrichment patterns potentially reflect the substrate repertoires of methylotroph populations in Lake Washington and their adaptation to specific  $C_1$  compound fluxes in the environment. The results of this project also pointed toward the role in local  $C_1$  cycling for species previously not implicated in this role, such as Betaproteobacteria belonging to *Rhodocyclales* and *Burkholderiales*, or for species with poorly understood ecological function, such as Planctomycetes. In conclusion, the functional metagenomics approach we present is designed to link the phylogenetic diversity of a chosen functional group to its genetic and metabolic potential, and ultimately to its ecological function.

#### 38. Whole Genome Analysis of MTBE-Degrading Beta-Proteobacterium *Methylibium petroleiphilum* Strain PM1

Staci Kane\* (kane11@llnl.gov), <sup>1</sup> Anu Chakicherla, <sup>1</sup> Patrick Chain, <sup>1,4</sup> Radomir Schmidt, <sup>2</sup> Maria Shin, <sup>1</sup> Kate Scow, <sup>2</sup> Frank Larimer, <sup>3,4</sup> Susan Lucas, <sup>4</sup> Paul Richardson, <sup>4</sup> and Krassimira Hristova <sup>2</sup> <sup>1</sup>Lawrence Livermore National Laboratory, Livermore, CA; <sup>2</sup>Dept. of Land Air and Water Resources, University of California, Davis, CA; <sup>3</sup>Genome Analysis Group, Oak Ridge National Laboratory, Oak Ridge, TN; and <sup>4</sup>DOE Joint Genome Institute Production Genomics Facility, Walnut Creek, CA

Methylibium petroleiphilum strain PM1 was one of the first pure culture strains isolated for its ability to completely degrade the noxious fuel oxygenate, methyl tert-butyl ether (MTBE). Strain PM1 can also degrade other gasoline components that are often co-contaminants with MTBE in groundwater, including benzene, toluene, and xylenes (BTX). The whole genome of strain PM1 was sequenced in order to elucidate the novel pathways that this bacterium uses for contaminant biodegradation as well as for its methylotrophic lifestyle. The sequence has revealed some surprising results including the presence of a large ~600 kb plasmid, presence of metal resistance/homeostasis genes and differences in predicted pathways for methylotrophy. Preliminary analysis reveals that the 4.0 Mb circular chromosome contains 3822 predicted genes and has a G+C sequence composition of 69.3% while the plasmid contains 632 predicted genes and has a G+C of 66.2%. This strain contains a single 16S rRNA gene. There are >10 repeat elements in the genome, repeated from 2 to >10 times, some as large as 30-40 kb in size. Furthermore, the plasmid has been characterized in terms of relative representation of functional groups of genes, IS elements, tRNAs and partitioning/origin of replication.

We have performed comparative genomic analysis with both phylogenetic and functional near neighbors of PM1. An overview of strain PM1 and its genome content, together with whole genome analysis and comparative analyses will be presented. The PM1 genomic sequence is available at http://genome.ornl.gov/microbial/rgel/. Strain PM1 serves as a model bacterium for petroleum contaminant degradation in methylotrophic MTBE-degrading bacteria in the environment.

## 39. Sequencing *Mycosphaerella* and *Cercospora* Species Will Revolutionize the Control of Major Global Threats on Wheat, Banana and Maize

**Gert H. J. Kema**\* (gert.kema@wur.nl), <sup>1</sup> Larry D. Dunkle, <sup>1</sup> Alice C. L. Churchill, <sup>2</sup> Jean Carlier, <sup>3</sup> Andy James, <sup>4</sup> Manoel T. Souza, Jr., <sup>5</sup> Pedro Crous, <sup>6</sup> Nicolas Roux, <sup>7</sup> Theo A. J. van der Lee, <sup>8</sup> Cees Waalwijk, <sup>8</sup> Erika Lindquist, <sup>9</sup> Jim Bristow, <sup>9</sup> and **Stephen B. Goodwin** <sup>8</sup>\*

<sup>1</sup>Plant Research International B.V., Wageningen, The Netherlands; <sup>2</sup>Dept. of Plant Pathology, Cornell University, Ithaca, NY; <sup>3</sup>UMR BGPI, CIRAD, Montpellier, France; <sup>4</sup>CICY, Merida, Mexico; <sup>5</sup>EMBRAPA Genetic Resources and Biotechnology, Brasília, Brazil; <sup>6</sup>Fungal Biodiversity Centre, Utrecht, The Netherlands; <sup>7</sup>INIBAP, Montpellier, France; <sup>8</sup>USDA-ARS/Purdue University, West Lafayette, IN; and <sup>9</sup>DOE Joint Genome Institute, Walnut Creek, CA

Mycosphaerella is one of the largest genera of plant pathogenic fungi with more than 1,000 named species, many of which are important pathogens causing leaf spotting diseases in cereals, citrus, banana, and eucalypts, soft fruits such as strawberry, and horticultural crops including many Brassica species. A few species of Mycosphaerella cause disease in humans and other vertebrates. The major Mycosphaerella plant pathogens include M. graminicola of wheat and M. fijiensis of banana, which require global annual fungicide inputs of \$400 million and \$2.5 billion, respectively. Both fungi are very important to the world economy. For example, M. graminicola causes more than \$275 million in losses annually to U.S. wheat growers, and more than 70% of the fungicides sprayed on wheat in Europe are to combat this pathogen. The International Mycosphaerella Genomics Consortium was started with the goal to sequence the genomes of M. graminicola and M. fijiensis. The former was chosen due to its genetic tractability, availability of extensive genomic resources, large worldwide research community and phylogenetic distinctiveness from other fungi being sequenced. The latter was chosen because of its worldwide impact on banana production. A joint project between the USDA-ARS/Purdue University and Plant Research International B.V. was initiated to sequence both genomes, along with 40,000 ESTs from both M. fijiensis and the related maize pathogen Cercospora zeae-maydis. The work was conducted through the Community Sequencing Program sponsored by the U.S. DOE Joint Genome Institute. The initial goals of

the projects are to: assemble 8× genomic shotgun sequences of M. graminicola strain IPO323 and M. fijiensis strain CIRAD 86; perform automated annotations of these genomic sequences and directed annotations using the ~80,000 ESTs from both *Mycosphaerella* species (the M. graminicola set will be made available by Syngenta); and make these sequences available publicly through a series of linked web sites for comparative analyses. Currently, the 8.9× M. graminicola sequence (518,271 traces) is available at NCBI. The 289,742 reads were organized into 2962 contigs spanning 37.65 Mb. These contigs were assembled into 187 scaffolds covering 39.05 Mb, giving a revised genome size of 41.2 Mb, slightly larger than estimated previously. Using a combination of gene prediction methods integrated into the JGI annotation pipeline we predicted a total of 11,395 gene models, with 80% of models supported by homology to other proteins. In addition, a draft mitochondrial assembly yielded a 43,962-base scaffold that appears to cover the complete genome. A community-wide annotation effort culminating in an annotation jamboree is anticipated for later during 2006 and will be open to all interested participants.

#### 40. CSP/USDA/NSF-Sponsored Sequencing of Phytophthora capsici and Benchmarking of de Novo Genome Sequencing with 454 Technology

Stephen Kingsmore\* (sfk@ncgr.org)

National Center for Genome Resources, Santa Fe, NM

## 41. Determination of the Baseline Proteome of the Halophilic Archaeon *Haloferax volcanii*

**P. Aaron Kirkland**, Stanley Stevens, Jr., Jennifer A. C. Busby, and Julie A. Maupin-Furlow\* (jmaupin@ufl.edu)

<sup>1</sup>Dept. of Microbiology and Cell Science, University of Florida, Gainesville, FL; <sup>2</sup>Interdisciplinary Center for Biotechnology Research, University of Florida, Gainesville, FL; and <sup>3</sup>Translational Research Institute, Dept. of Proteomics, Scripps Florida, Jupiter, FL

The genome sequence of the hemophilic archaeon *Haloferax volcanii* is nearly complete. To facilitate our understanding of this genome, we are determining the baseline proteome of *Hf. volcanii* and comparing this with the proteome deduced from the genome sequence. To this end, we have characterized the proteome of cells as they transition from log to stationary phase. In addition to a parent strain DS70 which is cured of plasmid pHV2, an isogenic proteasome mutant (GG102 *panA*) was analyzed based on the increased number of phosphoproteins detected in the proteome of this mutant strain compared to its parent, as determined by Pro-Q Diamond stained 2D-gels. Proteins of these two strains were

digested with trypsin, and peptides were separated by reverse phase and multidimensional protein identification technology (MudPIT), which incorporates back to back strong cation exchange and reverse phase chromatography steps. Peptide masses were determined by electrospray ionization quadrupole time-of-flight mass spectrometry (ESI-QTOF MS) with the Analyst OS data system (OSTAR) as well as LTO linear ion trap tandem mass spectrometry (LTQ MS/MS). SEQUEST and MASCOT were used to correlate the mass spectra of peptides with amino acid sequences deduced from the *Hf. volcanii* genomic database. Using these methods, we identified 1,678 proteins out of 4,083 predicted proteins with conservative filtering constraints. Peptides detected included those of proteins ranging in calculated molecular mass from 5 to 243 kDa and were cytosolic, membrane-associated and integral membrane proteins. Many of the proteins (over 250) were predicted to be involved in energy production and metabolism; however, a large number of hypothetical and conserved hypothetical proteins were also detected.

## **42.** Nitrifier Genomics – New Insights into a Simple Catabolic Lifestyle

Martin G. Klotz\* (jgiusermeeting@mgklotz.com), Daniel J. Arp, David J. Bergmann, Paul Berube, Peter J. Bottomley, Patrick S. G. Chain, Serad Elmore, Amal F. El-Sheikh, Loren J. Hauser, William J. Hickey, Norman G. Hommes, Alan B. Hooper, Miriam L. Land, Frank W. Larimer, Stephanie A. Malfatti, Selanette M. Norton, Manisha T. Poret-Peterson, Luis A. Sayavedra-Soto, Shawn R. Starkenburg, Lisa Y. Stein, Lisa M. Vergez, Selas B. Ward, Martin M. Xueming Wei University of Louisville, Louisville, KY; Oregon State University, Corvallis, OR; Black Hills State University, Spearfish, SD; University of Washington, Seattle, WA; Lawrence Livermore National Laboratory, Livermore, CA; DOE Joint Genome Institute, Walnut Creek, CA; University of Minnesota, St. Paul, MN; Oak Ridge National Laboratory, Oak Ridge, TN; University of Wisconsin, Madison, WI; Utah State University, Logan, UT; University of California, Riverside, CA; and Princeton University, Princeton, NJ

Nitrification, denitrification and nitrogen fixation, are the major processes moving inorganic nitrogen through the biogeochemical N cycle, which is carried out predominantly by bacteria. Nitrification is a two-step dissimilative process and involves the transformation of reduced inorganic nitrogen (i.e., ammonia) to more oxidized forms of nitrogen (i.e., nitrate). Ammonia is first oxidized to nitrite by ammonia-oxidizing bacteria (AOB), then nitrite is oxidized to nitrate by nitrite-oxidizing bacteria (NOB). The genome sequence of the AOB *Nitrosomonas europaea* 19718 was obtained by the DOE Joint Genome Institute as one of its first complete genomes and published in 2003. The complete first genome sequence of an NOB, *N. winogradskyi*, was just recently obtained as one

#### **Poster Presentations**

of five additional genomes of nitrifiers that are characteristic for their respective habitats. Of these, three genomes have been completely sequenced and closed, and their analysis is in progress: (A) Nitrobacter winogradskyi Nb-255, a facultative chemotrophic NOB that usually resides in consortia with respective AOB in soil, wastewater and freshwater/sediment environments. Highlights: an aerobic-type CO dehydrogenase, a Type I RuBisCO, 2 nitrite oxidoreductases and a glyoxylate shunt; lacks inventory needed for a complete glycolysis pathway (EMP or ED); (B) Nitrosospira multiformis ATCC 25196, the typical strain of AOB isolated from agricultural, forest, and grassland soils under both acid and neutral pH conditions in various regions of the world. Highlights: multiple copies of the gene clusters involved in ammonia and hydroxylamine oxidation, single copies of gene clusters encoding rRNAs, a Type I RuBisCO, urea hydrolase, urea amidolyase and a [Ni-Fe] hydrogenase; inventory to express complete central pathways (EMP, TCA, PPP); (C) Nitrosococcus oceani ATCC 19707, a worldwide-distributed marine AOB that is a representative of the gammaproteobacterial purple sulfur bacteria. Highlights: inventory to express complete central pathways (EMP, TCA, PPP); lacks a glyoxylate shunt; 2 functional rrn operons; one gene cluster each that encodes the capacity to oxidize ammonia and hydroxylamine; two copies each of the genes necessary to assemble functional complexes I and IV as well as ATP synthase (one H<sup>+</sup>dependent  $F_0F_1$ -type, one Na<sup>+</sup>-dependent V-type). Two genome projects are still in the closing phase: (D) Nitrosomonas eutropha C-91, a betaproteobacterial AOB isolated from sewage and Nitrobacter hamburgensis X14, an alphaproteobacterial NOB that was isolated from corroded limestone.

The ongoing analysis of these nitrifier genomes have initiated further experimentation and comparative analyses, most of which have led to significant revisions of prior paradigms, which include that: (i) nitrifiers do have a complete tricarboxylic acid cycle; (ii) nitrifiers are capable of glycogen and sucrose synthesis; (iii) nitrifiers have polyphosphate integrated in their energy and detox metabolism; (iv) some but not all nitrifying bacteria have specific catabolic capacity such as hydrogenase, urea hydrolase and amidolyase and CO-dehydrogenase; (v) the extent of nitrification-specific genome inventory has grown beyond AMO and HAO activity-related inventory; (vi) inference into the molecular evolutionary history of nitrification-specific genome inventory points toward its origins in denitrification and sulfur oxidation capacity; and (vii) nitrifier genomes have large regions of repeat elements and are living documents of rampant invasions by bacteriophages.

## 43. The Compositae Genome Project and Development of a Genechip for Massively Parallel Genotyping and Gene Expression Analysis in Lettuce

Alexander Kozik\* (akozik@atgc.org),¹ David G. Caldwell,¹ María José Truco,¹ Dean O. Lavelle,¹ Leah K. McHale,¹ Jason M. Argyris,¹ Marta Matvienko,¹ Oswaldo E. Ochoa,¹ Fallon Chen,¹ Smitha Mathrakott,¹ Kent J. Bradford,¹ Loren Rieseberg,³ Steve J. Knapp,⁴ Rick Kesseli,² David W. Still,⁵ Jeffrey L. Boore,⁶ Allen Van Deynze,¹ and Richard W. Michelmore¹¹ Genome Center, University of California, Davis, CA;²Dept. of Botany, University of Massachusetts, Boston, MA;³Dept. of Biology, Indiana University, Bloomington, IN; ⁴CAGT, University of Georgia, Athens, GA;⁵Dept. of Horticulture/Plant and Soil Science, California State Polytechnic University, Pomona, CA; and ⁶DOE Joint Genome Institute, Walnut Creek, CA

The initial phase of the Compositae Genome Project (CGP; http://compgenomics.ucdavis.edu) generated over 68,000 ESTs for lettuce (Lactuca sativa and L. serriola) and 67,000 ESTs for sunflower (Helianthus annuus, H. paradoxus, H. argophyllus) resulting in ~19,000 and ~18,000 unigenes respectively. Tools for visualization of the data were developed (http://cgpdb.ucdavis.edu) and the lettuce sequence information was subsequently used for mapping candidate genes using our core mapping population (L. sativa cv. Salinas x wild L. serriola, RIL F7:8). Candidate genes coincident with loci determining important agricultural traits, particularly disease resistance, are being investigated further. The second phase of the CGP is generating an expanded set of ESTs from five different Lactuca species (L. sativa, L. serriola, L. saligna, L. virosa and L. perennis) as well as twelve other Compositae species from five different genera. This expanded set of over 500,000 ESTs will be used for comparative genomics studies of rapidly evolving genes involved in domestication and weediness. In parallel, we have been evaluating the phenotypic diversity within and between the targeted species. Using the EST information, we are developing a 6.6 million feature Affymetrix Genechip for robust, massively-parallel genotyping based on Single Feature Polymorphisms (SFPs) as well as gene expression analysis in lettuce. Approximately 200 probes per unigene will be tiled across polymorphic coding and non-coding regions in 2 bp increments for at least ~30,000 unigenes. The design and the analysis tools we are developing should provide greater than a 95% chance of scoring each SFP correctly. In addition, probes will be designed to conserved regions of each gene and therefore may be useful for analysis of heterologous species in the Compositae.

## **44.** Improving Methods and Strategies for Sequencing Complex Genomes

**Gerard R. Lazo\*** (lazo@pw.usda.gov), Yong Q. Gu, Frank M. You, Curt C. Crossman, Devin Coleman-Derr, and Olin D. Anderson USDA-ARS Western Regional Research Center, Albany, CA

Sequencing technology and the tools by which to process information have improved tremendously over recent years. It is the combination of these technologies which help in adapting protocols by which to sequence complex genomes. Methods for sequencing range from random shotgun sequencing approaches using now traditional capillary electrophoresis methods to full genome sequencing using picolitre pyrosequencing approaches. Regardless of the methods used, highly repetitive regions of the genome can make it difficult to close gaps in attempts to build contiguous sequence data. A now common method used to prepare sequencing of the genome is to map out a sequencing scaffold from largeinsert genome libraries using fingerprint profiling techniques, sometimes referred to as high information content fingerprinting (HICF), and assembly of the scaffolds by software (Luo et al., 2003). The assembled scaffolds present a directed guide for the sequencing efforts of the genome. A similar methodology was used in the design of a sequencing strategy for the individual clones of the genome library, applying the fingerprint strategy to smaller subclones and using labels to restriction endonuclease fragments cut with enzymes of four base paired recognition sites. The fingerprint profile data was extracted with the software GenoProfiler (You et al., 2004), and assembled with the software FPC (Soderlund et al., 2000). The method helped to identify and reduce the number of sequences needed to complete sequencing of a BAC clone, normally sequenced using random shotgun sequencing approaches until complete coverage was apparent. In instances where genomes contain a high content of repetitive sequences, this approach helped to leverage efficiency in the sequencing of clones where routine random shotgun sequencing yielded difficult to close gaps. We have tested this method on BAC clones derived from the wheat genome, well known for its complex genome structure.

### **45.** A Functional Genomics Approach to Early Ascidian Development

Patrick Lemaire\* (lemaire@ibdm.univ-mrs.fr) CNRS

## 46. A Metagenomic Approach to Uncover the Genetic Diversity of Bacterial Degradation of the Plant Hormone Indole 3-Acetic Acid

**Johan Leveau\*** (j.leveau@nioo.knaw.nl) and Saskia Gerards Netherlands Institute of Ecology (NIOO-KNAW), Heteren, The Netherlands

Microbial manipulation of a host's hormone system represents an intriguing type of interorganismal interaction. Pseudomonas putida 1290 has previously been proposed as a model organism for the study of bacterial degradation of the plant hormone indole 3-acetic acid (IAA). In the past, bacteria that can destroy IAA have received little attention. despite their ubiquitous association with plants, their likely impact on plant functioning and health, and their potential as biocontrol agents for the suppression of plant diseases associated with excess IAA production by microbial pathogens. This lack of interest is perhaps best illustrated by the fact that no bacterial genes for the degradation of IAA have yet been identified. Here, we report on the cloning and sequencing of IAA degradative genes from P. putida 1290, and on the function-based recovery of similar genes from metagenomic DNA libraries. Transposon mutagenesis of P. putida 1290 revealed the involvement of a CbrAB twocomponent regulatory system in IAA catabolism and confirmed the role of catechol as an intermediate in the IAA degradation pathway. Complementation of P. putida KT2440, which is not able to degrade IAA, with any one of a group of 4 overlapping fosmid clones from a genomic DNA library of P. putida 1290 identified a 16-kb DNA fragment that conferred the ability to grow on IAA as a sole source of carbon. Sequence analysis of this DNA fragment exposed several candidate genes coding for the conversion of IAA to catechol, some of which were highly similar to genes with no function assigned yet on the genomes of fully sequenced bacteria including Burkholderia xenovorans LB400 and Novosphingobium aromaticivorans DSM12444. We recognized that our complementation strategy which exploits IAA P. putida KT2440 as a host strain could easily be adapted as a functional (meta)genomics approach to recover IAA catabolic genes from (meta)genomic DNA. To this end, we screened several large-insert libraries with DNA from various sources (e.g. from newly isolated bacterial IAA degraders belonging to the species Pseudomonas, Burkholderia, or Arthrobacter, from grassland bulk soil, or from the Festuca rubra rhizosphere) for clones that conferred an IAA<sup>+</sup> phenotype to P. putida KT2440. Already, we have identified >40 such clones, and analysis of their DNA inserts will greatly advance our understanding of the types and diversity of genes underlying bacterial IAA metabolism and will provide the currently non-existing but much needed framework for a genetic approach to the phenomenon of bacterial IAA degradation and its role in plant-microbe interactions.

1. Leveau and Lindow (2005). Applied and Environmental Microbiology 71: 2365-2371.

## 47. Preliminary Insights from Comparison of Five Newly Sequenced Hyperthermophile Crenarchaea

**Todd M. Lowe\*** (lowe@soe.ucsc.edu) University of California, Santa Cruz, CA

#### 48. Toward Resolution of Green Plant Phylogeny

**Dina F. Mandoli\*** (mandoli@u.washington.edu), <sup>1</sup> Jeffrey L. Boore, <sup>2</sup> Karin Everett, <sup>1</sup> Michael J. Donoghue, <sup>6</sup> Kenneth G. Karol, <sup>1</sup> Jennifer Kueh<sup>1,2</sup> Brent Mishler, <sup>3</sup> Charlie O'Kelley, <sup>7</sup> Richard Olmstead, <sup>1</sup> Alan R. Smith, <sup>3</sup> Karen Renzaglia, <sup>4</sup> and Paul Wolf<sup>5</sup>

<sup>1</sup>Dept of Biology, University of Washington, Seattle, WA; <sup>2</sup>DOE Joint Genome Institute Production Center, Walnut Creek, CA; <sup>3</sup>University Herbarium, University of California, Berkeley, CA; <sup>4</sup>Dept of Plant Biology, Southern Illinois University, Carbondale, IL; <sup>5</sup>Dept of Biology, Utah State University, Logan, UT; <sup>6</sup>Dept. of Ecology and Evolutionary Biology, Yale University, New Haven, CT; and <sup>7</sup>Bigelow Laboratories for Ocean Sciences, West Boothbay Harbor, ME

We are funded to resolve the primary pattern of evolutionary diversification among green plants, and to establish a model for doing so that will be applicable to other groups of organisms with long evolutionary history. To achieve this goal we are 1) completing a matrix of whole genome sequences for chloroplast and mitochondria; 2) producing a comprehensive set of comparable morphological and ultrastructural data for these same taxa; and 3) incorporating inferences from across the phylogenetic hierarchy in green plants using methods designed to permit scaling across studies. We shall indicate how this work will link to other research being conducted on green plants at various scales, especially the concatenation of our data sets with theirs. We will share the methods we are using, primarily FACS and RCA, to isolate and clone genomes, show the genomes we have assembled and relay the implications of these genomes to the phylogeny of the Chlorophytes. We will list genomes we are working on and introduce new taxa we have found. We will summarize our morphological analyses and discuss advances incorporating inferences from across the phylogenetic hierarchy. Funded by NSF's Tree of Life Program, 2002-2006, DEB #0228655 (lead institution), #228432, #0228679, #0228729, #0228660, #0228576.

### **49.** High-Throughput Protein Production via Cell-Free Methods

**Meng L. Markillie\*** (meng.markillie@pnl.gov) and Eric Ackerman Pacific Northwest National Laboratory, Richland, WA

Developments in disparate fields now enable high-throughput protein expression via custom and commercially-available robots for cell-free, protein expression using wheat-germ or E. coli extracts. We have a robot capable of producing 384 proteins per ~20 hour run in quantities of ~50 to 500 ug/protein using PCR templates or plasmids. The major expense per run is incurred by oligonucleotides and PCR reagents — not the wheatgerm cell free extracts producing proteins directly from transcribed PCR templates, or if desired, from more expensive plasmids. Disparate sets of proteins can be targeted for expression, characterization, and optimization. The overall goal of this project is to develop and deploy scaleable capabilities for high-throughput production of active proteins utilizing cell-free methods. Protein targets will be chosen based on several criteria, but a dominant theme will be to support and extend our discovery that enzymes immobilized in properly chosen and configured nanoporous materials display remarkable properties of enhanced activity and stability of use for DOE missions in remediation, energy production, and security.

#### 50. Sugarbeet Breeding, Genetics, and Genomics Progress

Mitch McGrath\* (mitchmcg@msu.edu)

USDA-ARS and Crop and Soil Sciences, Michigan State University, East Lansing, MI

Sugarbeet is an industrial crop whose main product is sucrose, which constitutes 50 - 70% of the biomass produced during its cool temperate growing season. This simple, easily digestible, carbon source is primarily used as a sweetener, and increasingly to provide a concentrated carbon source during fermentation (e.g. ethanol fuel production). New uses for beets are being sought globally, and all parts of the plant already have an economic outlet. Expanding beet breeding to energy uses was a target during the first energy crisis, and remains a viable option today. The key will be to improve biomass production. Preliminary analyses suggest genetic variability for biomass production is available, controlled by relatively few QTL. Identifying these genes is a goal, however the tools available to dissect traits in beets to the molecular level are somewhat rudimentary. Current genomic efforts are geared to build into a complete genome sequence from which genetic markers, gene identification, and genome organization as a representative of the important and widely adaptable Chenopod plant family can be developed.

## 51. Genome of *Syntrophus aciditrophicus*: Lifestyle of the Thermodynamically Challenged

**Michael J. McInerney\*** (mcinerney@ou.edu), <sup>1</sup> Robert Gunsalus, <sup>2</sup> John Campbell, <sup>3</sup> Lars Rohlin, <sup>2</sup> Housna Moutakki, <sup>1</sup> UnMi Kim, <sup>2</sup> Rebecca Krupp, <sup>2</sup> Luis Rio-Hernandez, <sup>1</sup> Jessica Sieber, <sup>1</sup> and Christopher G. Struchtemyer <sup>1</sup>

<sup>1</sup>Dept. of Botany and Microbiology, University of Oklahoma, Norman, OK; <sup>2</sup>Dept. of Microbiology, Immunology, and Molecular Genetics, University of California, Los Angeles, CA; and <sup>3</sup>Integrated Genomics, Chicago, IL

The completed genome sequence of the fatty- and aromatic-degrading, syntrophic bacterium, Syntrophus aciditrophicus, provides the first glimpse of the composition and architecture of the electron transfer and energy transducing systems needed to exist on marginal energy economies by a syntrophic lifestyle. Syntrophy is a thermodynamically based interaction, where the degradation of the primary substrate is thermodynamically unfavorable unless a second species maintains H<sub>2</sub> and/or formate concentrations produced by the first species at very low levels. Syntrophic metabolism proceeds close to thermodynamic equilibrium and requires reverse electron transfer with large negative changes in electrical potential ( $\Delta E$ ) to form H<sub>2</sub> and formate. The genome of S. aciditrophicus contains 3,179,300 base pairs in a single circular chromosome. Approximately 3,100 genes were identified and annotated functions were assigned to 1,618 genes (51%). Two potential regions of horizontal transfer were identified on the basis of deviation from the 51% average GC composition. One region corresponds to a Mu-like prophage; the other anomalous region includes a potential conjugative element. The genes were organized into pathways based on the functional assignments and all pathways considered essential for viability of a typical gramnegative organism appear to be present in S. aciditrophicus, with a few notable exceptions. The genome does not appear to encode any recognizable carbohydrate transporters for mono-, di- or even oligo saccharides. Identifiable transporters for amino acids and small peptides are also extremely limited. The lack of transport of carbohydrates, amino acids and peptides is reflected in the paucity of degradation pathways associated with life on these substrates. Interestingly, no genes homologous to the multifunctional isomerase-hydrolase-dehydrogenase (fadB) subunit, nor to the thiolase (fadA) subunit of the common bacterial fatty acid degradation pathway, are recognizable in the genome. This is particularly odd since fatty acids and aromatic substrates are growth substrates for the organism. The genome also lacks homologs for key reactions in known pathways for 2-oxoglutarate synthesis and it may be made by a novel pathway involving glutaconyl-CoA and 2-hydroxyglutaryl-CoA. Proton and sodium gradients can be formed by membrane-bound pyrophosphatases, glutaconyl-CoA decarboxlase, and proton- and sodium-dependent ATP synthetases. The ion gradients can be coupled to electron transfer by a variety of integral membrane components such as menaquinone, Na<sup>+</sup>-translocating NADH:quinone oxidoreductase, and a putative Na<sup>+</sup>-translocating channel. The genome has five gene clusters with homologs to heterodisulfide reductase complexes found in Archaea; four of which also contain homologs to archaeal aldehyde oxidoreductases. Two of these clusters may be involved in aromatic ring reduction based on similarity to the newly detected, aromatic ring reductase genes in *Geobacter metallireducens*. Surprisingly, for a fermentative anaerobe, the genome of *S. aciditrophicus* lacks gene homologous to those for acetate kinase. Nine genes predicted to encode for ADP-forming acetyl-CoA synthetases, which synthesize ATP from acetyl-CoA, ADP and P<sub>i</sub> are present. The genome sequence of *S. aciditrophicus* suggests a unique strategy for growth with fatty and aromatic acids by a syntrophic lifestyle.

#### 52. The Genomes of Five PAH Degrading Mycobacteria

**Charles Miller\*** (cdmiller@biology.usu.edu), <sup>1,2</sup> Ronald Sims, <sup>2</sup> and Anne Anderson<sup>1,2</sup>

<sup>1</sup>Dept. of Biology and <sup>2</sup>Dept. of Biological and Irrigation Engineering, Utah State University, Logan, UT

Polycyclic aromatic hydrocarbons (PAHs) are organic compounds consisting of two or more fused benzene rings. Because of their toxic, mutagenic, and carcinogenic properties, 16 PAHs are listed on the U.S. Environmental Protection Agency's priority-pollutant list. We have identified five fast growing mycobacterium isolates that degrade high molecular weight PAHs to water and carbon dioxide. Such isolates have potential in remediation of PAH-polluted sites. Mycobacterium isolates JLS, KMS and MCS were from a PAH-contaminated land-farm soil at the EPA superfund site at Libby Montana, where active remediation was being demonstrated. M. vanbaalenii isolate PYR-1 and M. flavescens were from Dr. Carl Cerniglia, being isolated from a PAH-contaminated watershed at Redfish Bay near Port Aransas, and from the Grand Calumet River sediment in northwest Indiana, respectively. Pulse field gel electrophoresis identified linear plasmids in M. KMS, M. MCS and M. flavescens. Draft level sequencing completed in Fall 2005, of the five PAH-degrading mycobacteria by the Joint Genomes Institute revealed genomes sizes ranging from 5.9 to 6.4 Mb and in 16 to 45 contigs. These draft level sequences were approximately 68% GC and contained between 5559 and 6012 potential open reading frames. The size of the PAHdegrading genomes is larger than the genomes for pathogenic mycobacterium isolates (about 4.4 Mb for *M. tuberculosis* H37 RV, M. tuberculosis CDC1551, M. bovis AF2122/97, M. microti, and 3.7 Mb M. leprae) but similar to M. avium (6.6 Mb). M. smegmatis, a nonpathogen, sequenced because it has been used as a surrogate for the pathogenic mycobacteria, has a faster growth time like the PAH-degraders and a similar genome size (6.9 Mb). We are initiating categorizing the sets of genes that contribute to survival of the PAH-degrading mycobacterium

#### **Poster Presentations**

in soils/water versus the genes responsible for the pathogenic niche of pathogenic mycobacteria. Additionally, we will investigate the genes involved in the PAH-degradative pathways for comparison to pathways in other microbial genera.

## 53. Genomic Diversity of *Oenococcus oeni* Strains Isolated from Different Winemaking Regions

**David A. Mills\*** (damills@ucdavis.edu) and Angela Marcobal Dept. of Viticulture and Enology, University of California, Davis, CA

Oenococcus oeni is a unique lactic acid bacteria isolated solely from fruitmash and winery environments. Unlike many other lactic acid bacteria, O. oeni is tolerant to both high levels of alcohol and low pH conditions making it ideal for performing the malolactic conversion, a key secondary fermentation in the production of wine. In collaboration with DOE Joint Genome Institute, the genome sequence of O. oeni PSU-1 has been determined. The complete PSU-1 genome is 1,780,517 nucleotides with a GC content of 38%. 1701 ORFs could be predicted from the sequence of which 75% were functionally classified. Consistent with its classification as an obligately heterofermentative lactic acid bacteria PSU-1 genome encodes all the enzymes for the phosphoketolase pathway. Genes and pathways involved to production of flavor compounds in wine, such as the malolactic conversion, citrate utilization, glycoside activity and catabolism of amino acids were readily identified. Array-based comparisons of ten O. oeni isolates obtained from diverse winemaking regions indicate a high level of genome conservation with all strains possessing at least 96% of the genes present in PSU-1. Strains appeared to cluster into three groups, with no relationship between geographic locale and cluster membership. Interestingly, PSU-1, an isolate obtained from a spontaneous malolactic fermentation in Pennsylvania was shown to be nearly identical to MCW, a well-known O. oeni isolate obtained from a spontaneous malolactic fermentation at Matanzas Creek Winery in California. The lack of diversity apparent in the O. oeni species suggests a wide distribution of strains coincident with winemaking practice. The completion of the O. oeni genome marks a significant new phase for research on the malolactic fermentation in wine whereby the physiology, diversity and performance of O. oeni starter cultures can be more rigorously examined.

#### 54. The Green Tree of Life

**Brent D. Mishler\*** (bmishler@calmail.berkeley.edu) University of California, Berkeley, CA http://ucjeps.berkeley.edu/TreeofLife/

## 55. The Maltose Transporter Genes of *Thermotoga maritima* are Unusually Close Homologs of those from Gamma Proteobacteria

**Dhaval M. Nanavati**, Pascal Lapierre, J. Peter Gogarten, and **Kenneth M. Noll\*** (kenneth.noll@uconn.edu)

Dept. of Molecular and Cell Biology, University of Connecticut, Storrs, CT

The corresponding proteins encoded by the two mal ABC transporter operons of the hyperthermophilic bacterium *Thermotoga maritima* show very high sequence similarities with one another. To determine the origin of these two operons, we carried out phylogenetic analyses of the sequences of the homologous MalE, MalF, and MalG proteins. MrBayes, parsimony and distance matrix methods showed each of these genes are most closely related to their paralog in T. maritima suggesting that the entire operon duplicated or that an ortholog was inherited via horizontal transfer. These phylogenetic analyses grouped MalF and MalG sequences into nearly identical phylogenetic relationships as those found with MalE suggesting that these genes evolved together. A phylogenetic analysis of all MalE homologs showed that the T. maritima homologs are most closely related to one another and represent only one of two major lineages of MalE proteins. An apparent ancient duplication of malE occurred and some members of the Thermococcales now harbor a representative of each lineage. The malF and malG lineages show no evidence of an analogous duplication event. Our analyses show that the T. maritima Mal protein sequences group with their respective homologs from enteric bacteria. In contrast to the MalEFG homologs, an apparent T. maritima MalK homolog (the ATP-binding protein) is more closely related to archaeal MalK homologs from the *Thermococcales*.

Putative structural features observed in the *T. maritima* MalF and MalG protein sequences support the phylogenetic placements for these proteins. The sequence of the *T. maritima* MalF1 homolog suggests it contains eight transmembrane helices and a large periplasmic loop similar to one found uniquely in the MalF proteins of *E. coli* and its close relatives. Hydrophilic domains of both *T. maritima* MalG proteins are predicted to have coiled coil secondary structures similar to those domains in chromosomal maintenance proteins (COG1196). This domain is unique to the *T. maritima* and *Thermotoga neapolitana* MalG homologs indicating that the *mal* genes were acquired only once by their ancestor.

We also have identified both *mal* operons in *Thermotoga sp.* strain RQ2 indicating that these *mal* operons appeared in the *Thermotogales* at least by the time the ancestor of the *Thermotoga* branch emerged. Our data suggest that the *T. maritima malEFG* genes are only distantly related to their homologs in archaeal hyperthermophiles and that they have been shared among a small number of bacterial lineages including the enteric bacteria.

## 56. The *Calyptogena magnifica* Symbiont Draft Genome: An Obligate, Maternally Transmitted Endosymbiont with Extensive Metabolic Capabilities

**I. L. G. Newton\*** (irene.newton@gmail.com), <sup>1</sup> F. J. Stewart, <sup>1</sup> T. Woyke, <sup>2</sup> P. M. Richardson, <sup>2</sup> K. W. Barry, <sup>2</sup> J. C. Detter, <sup>3</sup> D. C. Bruce, <sup>3</sup> S. Sullivan, <sup>4</sup> J. A. Eisen, <sup>4</sup> and C. M. Cavanaugh <sup>1</sup>

<sup>1</sup>Organismic and Evolutionary Biology Department, Harvard University, Cambridge, MA; <sup>2</sup>DOE Joint Genome Institute, Production Genomics Facility, Walnut Creek, CA; <sup>3</sup>DOE Joint Genome Institute, Los Alamos National Laboratory, Los Alamos, NM; and <sup>4</sup>The Institute for Genomic Research, Rockville, MD

Chemoautotrophic endosymbionts form the metabolic foundation for communities at deep-sea hydrothermal vents and cold seeps. These  $\gamma$ proteobacteria use reduced sulfur compounds as an energy source for carbon fixation, providing their hosts (e.g., the vestimentiferan Riftia pachyptila) the majority of their carbon requirements. At hydrothermal vents, the giant clam, Calyptogena magnifica, harbors chemoautotrophic bacteria in its gills. These bacterial symbionts are transmitted via the eggs each generation, and the mutualism between clam and bacterium is obligate — neither has been found alone in the natural environment and the symbionts have yet to be cultured. Given that this symbiosis is maternally transmitted and is relatively old (between 50 and 100 MY), we hypothesized that the C. magnifica symbiont would have a reduced genome with skewed base composition, as has been found in insect endosymbiont genomes as well as most obligate intracellular species. The genome of the C. magnifica symbiont is being sequenced from a mixed gDNA sample (host and symbionts) by the DOE Joint Genome Institute and currently constitutes 3-5 contigs at 14x coverage. The draft of the symbiont genome is AT rich (35% GC) and estimated to be relatively small (~1.2Mb). Although reduced in size, the *C. magnifica* genome contains not only the chemoautotrophic pathways thought to be central to the symbiosis (e.g., Calvin cycle enzymes and sulfur oxidation pathways) but also many of the metabolic pathways of free-living chemoautotrophs (e.g., biosynthetic pathways for essential amino acids). This is the first description of a chemoautotrophic symbiont genome and our analyses reveal the extent of host-symbiont metabolic dependency and the evolutionary pressures on the symbiont genome. The study of extant bacterial-eukaryote symbioses will allow us to better understand how metabolic symbioses form and evolve and provide insights into the evolution of eukaryotic organelles such as mitochondria and chloroplasts.

## 57. Gene Expression and Genomic Analysis of the *in situ* Geobacteraceae during Uranium Bioremediation

Regina O'Neil\* (rtarallo@microbio.umass.edu), Dawn Holmes, Lorrie Adams, Juliana Larrahando, Joy Ward, Helen Vrionis, Kelvin Gregory, Kelly Nevin, Lucie N'Guessan, and Derek Lovley University of Massachusetts, Amherst, MA

Geobacteraceae have been shown to be the dominant microorganisms during bioremediation of a U(VI) contaminated aquifer in Rifle, CO when Fe(III) and U(VI) reduction are stimulated by the addition of acetate. Molecular studies have shown that the diversity of 16S rRNA gene sequences detected in the groundwater and sediments during the U(VI) bioremediation process is extremely low. For example, Geobacteraceae with 16S rRNA gene sequences closely related to the subsurface isolate Geobacter bemidiiensis accounted for 98% of the microbial community. and these sequences were 97-100% similar to each other. Four methods are currently being used to obtain genomic information from the dominant Geobacteraceae at this site: 1) shotgun sequencing of environmental genomic DNA, 2) sequencing of the genome from a single cell amplified by multiple displacement amplification, 3) gene specific sequencing using targeted degenerate Geobacteraceae primers, and 4) sequencing of isolates which have an exact 16S rDNA match to the predominant Geobacteraceae. Significant progress has been made with these methods. Shotgun sequencing of environmental genomic samples obtained during the peak of U(VI) bioremediation, when Geobacteraceae accounted for 99% of the microbial population, is currently underway. In addition, methods to isolate and amplify the genome of a single Geobacteraceae cell from the environment have also been developed, resulting in the ability to amplify over 80% of the genome from this single environmental cell. Information obtained from the targeted gene amplification approach has also been valuable, and has indicated that a number of genes that are conserved among the seven sequenced Geobacteraceae genomes are also found in the environment. Techniques have also been developed to isolate environmentally relevant Geobacter species that have 16S rRNA gene sequences that are identical to the dominant Geobacteraceae during uranium bioremediation.

The genomic information obtained from these approaches is currently being used to analyze the metabolic state of *Geobacteraceae* during uranium bioremediation. Quantitative RT-PCR primers targeting specific genes from the dominant *Geobacteraceae* were designed from genomic sequences obtained from the environment and used to monitor gene expression during the process of uranium bioremediation. The results to date suggest that there is a correlation between increased reduction of Fe(III) and U(VI) in the subsurface and the expression of specific *Geobacteraceae* genes.

#### 58. The rRNA-Based Map of Biological Diversity

**Norman R. Pace\*** (nrpace@colorado.edu), J. Kirk Harris, Jeffrey J. Walker, John R. Spear, Ruth E. Ley, Phillip Hugenholtz, and Falk Warnecke

<sup>1</sup>University of Colorado, Boulder, CO; <sup>2</sup>Colorado School of Mines, Golden, CO; <sup>3</sup>Washington University School of Medicine, St. Louis, MO; and <sup>4</sup>DOE Joint Genome Institute, Walnut Creek, CA

Our understanding of the extent of microbial diversity has expanded dramatically over the past decade. In large part this expansion has resulted from analysis of ribosomal RNA sequences from uncultured environmental microbes. The number of known main phyla, phylogenetic divisions, of bacteria has increased from 12 in 1987 to currently ~100. Only about one-quarter of these kingdom-level clades have cultured representation and it is common that the members of these uncultured clades are dominant in their respective environments. Thus, the breadth of diversity represented by environmental sequences far outreaches that of biological experience through cultivation. The number of environmental sequences in the databases continues to expand exponentially and now exceeds the numbers from cultivars. Results with Bacteria, Eucarya and Archaea have refined substantially our view of the large-scale Tree of Life. A challenge at this time is to learn the phenotypic natures of uncultured microorganisms.

Although molecular phylogeny with rRNA gene sequences provides some perspective on the evolutionary course of the genetic machinery, rRNA does not necessarily speak to phenotype as manifest in the environment. Much overtly expressed phenotype depends on genes that come and go through lateral gene transfer or propagation through gene families. Thus, for correlations of phenotype (ecotype) with phylotype, as seen by rRNA sequence, it will be necessary to cull phenotypic information from studies such as metagenomic sequencing efforts. Current efforts of the laboratory with JGI involve sequencing deeply into the community rRNA genes of the Guerrero Negro hypersaline microbial mat and shotgun sequencing community DNA of this complex, laminated ecosystem.

#### 59. High-Throughput Assembly

Jasmyn Pangilinan\* (jlpangilinan@lbl.gov) DOE Joint Genome Institute, Walnut Creek, CA

# 60. Genomic Analysis of the Aromatic Catabolic Pathways of *Cupriavidus necator* (ex *Ralstonia eutropha*) Provides Clues for Adaptation to Aromatic Compound Catabolism in Bacteria

**D. Perez-Pantoja**, R. De la Iglesia, J. E. Gonzalez-Pastor, V. de Lorenzo, D. H. Pieper, and **B. Gonzalez** (bgonzalez@bio.puc.cl) Depto. Genetica Molecular y Microbiologia y EMBA Millennium Nucleus. FCB. P. Universidad Catolica de Chile, Chile; Centro de Astrobiologia (CSIC-INTA), Spain; and Division of Microbiology, GBF, Braunschweig, Germany

The complete genome sequence of C. necator JMP134 (ex R. eutropha), the best known (halo) aromatic compounds degrading beta proteobacterial strain, is available. We performed the metabolic reconstruction of aromatic compounds degradation, correlating the catabolic abilities as determined in silico with a complete study of the range of compounds that support growth of this bacterium. More than 50 out of 160 tested aromatic compounds were used as a sole carbon source. Catabolic genes encoding the degradation pathways for aromatics were identified by BLAST using published gene sequences as in silico probes. We observed a strong correlation between the catabolic abilities found in silico with those determined in vivo. At least ten central ring-cleavage pathways for aromatics, described so far in the class Proteobacteria, were encoded in this genome. Functionality of most of the predicted catabolic pathways was verified with a 50-mer oligonucleotide DNA microarray including 350 catabolic genes of *C. necator*. On the other hand, strict hierarchy of use in aromatics mixtures was observed for most chromosomally encoded functions. Gene redundancy expressed as repeated complete operons, gene encoding peripheral or central reactions, and gene regulators, is a key feature in catabolic versatility. This work confirmed the potential of complete genome sequencing to predict the metabolic abilities of microorganisms and the utility of microarrays to confirm metabolic predictions.

Work supported by a FONDECYT grant (1030493), the MIFAB Millennium Institute, the EMBA Millennium Nucleus, the DOE-JGI initiative and the ICA4-CT-2002-10011 (ACCESS) Contract of the European Union.

#### 61. Soybean Rust Genome Project

**Martha L. Posada-Buitrago\*** (mlposada-buitrago@lbl.gov), <sup>1</sup> Jeffrey L. Boore, <sup>1</sup> and Reid D. Frederick<sup>2</sup>

<sup>1</sup>DOE Joint Genome Institute, Walnut Creek, CA, and <sup>2</sup>USDA-ARS Foreign Disease-Weed Science Research Unit, Fort Detrick, MD

Soybean rust, the most devastating foliar disease responsible for significant losses of soybean crop in Africa, Asia, Australia and South

#### **Poster Presentations**

America, is caused by the fungal pathogen *Phakopsora pachyrhizi*. The pathogen was found for the first time in the continental U.S.A. in November 2004 and has spread to more than 130 counties in the Southeastern States, becoming a major threat to U.S. soybean production.

Here, we present an update on the *Phakopsora pachyrhizi* Genome Sequencing Project funded by the U.S. Department of Agriculture, Agricultural Research Service; the U.S. Department of Defense; and the U.S. Department of Energy.

#### 62. Jazz – The JGI Genome Assembler

**Nik Putnam**, <sup>1</sup> Jarrod Chapman, <sup>1</sup> Dan Rokhsar, <sup>1</sup> **Isaac Ho\*** (iyho@lbl.gov), <sup>2</sup> Serge Dusheyko, <sup>2</sup> and Craig Furman <sup>3</sup> <sup>1</sup>DOE Joint Genome Institute, Dept. of Physics, University of California, Berkeley, CA; <sup>2</sup>DOE Joint Genome Institute, Lawrence Berkeley National Laboratory, Berkeley, CA; and <sup>3</sup>DOE Joint Genome Institute, Lawrence Livermore National Laboratory, Livermore, CA

Jazz, originally developed by graduate students Nik Putnam and Jarrod Chapman under the direction of Dr. Dan Rokshar, is a whole-genome shotgun assembler designed to handle paired-end sequencing data sets both small and large. It has been used in several high-profile assemblies, notably *Ciona intenstinalis*, *Fugu rubripes*, and *Xenopus tropicalis*.

Jazz relies on the overlap-layout-consensus paradigm made prominent by the Celera assembler; several algorithmic innovations distinguish Jazz from similar products, including evaluation of sister pairing early in the layout process, a hybrid depth/percent-id alignment scoring system, and a classical spring model for resolving complex scaffold topologies. Future directions of research include explicit handling of polymorphism, adaptation to cutting-edge prototype sequencing technologies, and the seamless integration of various forms of linking data such as restriction maps.

## 63. Genome Sequence and Comparative Analysis of the Solvent-Producing Bacterium *Clostridium beijerinckii*

Flavia A. Ramirez\* (framirez@uiuc.edu) University of Illinois, Urbana-Champaign, IL

### 64. Massive Horizontal Gene Transfer in the *Amborella* Mitochondrial Genome

**Aaron O. Richardson\*** (arichard@bio.indiana.edu) Indiana University, Bloomington, IN

## 65. Genotypic Analysis of European and U.S. Populations of *Phytophthora ramorum* by Microsatellite Markers Derived from Genome Sequence

**Noah Rosenzweig\*** (noahr@nature.berkeley.edu) and Matteo Garbelotto Dept. of ESPM-ES, University of California, Berkeley, CA

Sudden oak death (SOD) caused by a newly described *Phytophthora* species (*Phytophthora ramorum*), has recently become a major disease of concern in western U.S. coastal forest ecosystems and European (EU) nurseries and private estates. Genetic analysis was used to elucidate the genetic structure of the pathogen in its known worldwide range, with the aim of understanding its population biology and its migration patterns. Microsatellite analysis of twelve polymorphic simple sequence repeats (SSR) identified from the genome sequence of *P. ramorum*, revealed three distinct clades among 151 isolates. Populations recovered from U.S. forest and EU nursery isolates clustered into two well-supported separate clades, while isolates from a U.S. nurseries belonged to a third novel clade. Phytophthora ramorum genotypic diversity from populations recovered from nurseries and forests was significantly higher 91% compared to 18% respectively. Multilocus SSR analysis determined populations of P. ramorum collected from U.S. forests may reproduce clonally and are likely descendants of a single introduced individual. The genetic structure of populations recovered from EU nurseries displayed higher complexity, including multiple, closely related genotypes. All three clades were identified in some U.S. nurseries, including genotypes that perfectly matched the U.S. wild and the EU nursery SSR genotypic profiles, emphasizing the role of commercial plant trade in the introduction and migration of this pathogen. The combined SSR or microsatellite, sequencing and morphological analyses suggest the three clades represent distinct evolutionary lineages, all exotic to the U.S. and probably Europe and of unknown origin. Analysis using two hyper-variable tetra-repeat microsatellite loci of over 200 additional isolates, allowed for genotyping of asexually generated individuals. Results from a statewide monitoring study indicated different genotypes arise and dominate locally. In one instance, evidence of selection against old genotypes by newer ones was obtained. Genotyping allowed for the elucidation of possible migration patterns of the pathogen. Our study documents the local and regional adaptation of an introduced pathogen through the creation of novel genotypes via the accumulation of favorable mutations and/or somatic recombination.

## 66. Comparison of Four Microbial Genomes: Desulfovibrio vulgaris, Syntrophobacter fumaroxidans, Methanosarcina barkeri, and Methanospirillum hungatei

**Johannes C. M. Scholten\*** (johannes.scholten@pnl.gov), Fred J. Brockman, and Weiwen Zhang

Biological Sciences Department, Pacific Northwest National Laboratory, Richland, WA

Our project team at Pacific Northwest National Laboratory (PNNL) is developing a method to predict biochemical behavior in a simple, fourmicroorganism community.

A set of four microorganisms form together a "simple" quad-culture community that represent a versatile sulfate-reducer that is also a syntroph (Desulfovibrio vulgaris), a syntrophic specialist that prefers not to be a sulfate-reducer (Syntrophobacter fumaroxidans), a versatile methanogen that can live acetoclastically or hydrogenotrophically (Methanosarcina barkeri), and a specialist hydrogenotrophic methanogen (Methanospirillum hungatei). The syntrophic and methanogenic guilds interact intimately with each other, their physiologies are very well known, the genomes are sequenced, the two foundational dual-cultures are well understood, and we believe this quad-culture is the most constrained microbial system available to develop an approach for identifying, understanding, and modeling dynamic and coupled inter-species molecular networks within a "simple" microbial community. Thus, the quad-culture model system is a relatively highly constrained microbial community and represents an optimal system to learn how to identify inter-species metabolic networks and interactions. Through this study, we will characterize phenotypic traits important, for example, to the sulfatereducing and/or syntrophic lifestyles of *D. vulgaris* and *S. fumaroxidans*. The system we are using allows us to study different microbial interactions in a four-microorganism or less complex community.

The organisms *D. vulgaris* and *S. fumaroxidans* are closely related to each other as are *M. barkeri* and *M. hungatei* given that they belong to the *delta-Proteobacteria* and *Archaea*, respectively. The goal of our research is to describe carbon and energy flow in the microbial community using transcriptomic, proteomic and macroscopic process data. In collaboration with The Nimblegen Systems, we have developed customer oligonucleotide microarrays for each of these four species. In order to minimize the possible cross-hybridization during microarray analysis, extra efforts have been made to compare the genome sequences of all four species, and only the probes unique to each genome were picked and used in the microarrays. In addition, we also used the Integrated Microbial Genomes website (http://img.jgi.doe.gov/cgi-bin/pub/main.cgi) to compare the genomes of *D. vulgaris*, *S. fumaroxidans*, *M. barkeri* and *M. hungatei*. In our paper we present some preliminary results we obtained by comparing two, three of four genomes at the same time.

Furthermore we list the genes that might be of concern for cross-hybridization.

## 67. Next Generation DNA Sequencing: Clonal Single Molecule Array Technology and its Application to Genetic Analysis

**Gary P. Schroth\*** (gschroth@solexa.com) Solexa, Hayward, CA

## **68.** High-Throughput Eukaryotic Whole-Genome Shotgun Assembly

**Harris Shapiro\*** (hjshapiro@lbl.gov), Jasmyn Pangilinan, and Hank Tu DOE Joint Genome Institute, Walnut Creek, CA

Historically, the amount of time needed to run, analyze, and release whole-genome-shotgun (WGS) assemblies of eukaryotic genomes has been on the order of months or (usually) years. This has allowed for detailed human inspection and correction of assembly results, as well as extensive customization of WGS assemblers, sometimes on a per-release basis.

As the scale of WGS sequencing has increased, the amount of time available for such pre- and post-assembly analysis has been drastically reduced. The DOE Joint Genome Institute presently sequences 20 - 30 eukaryotic genomes per year, ranging in size from 15 MB to more than 1 GB. Keeping up requires running, assessing, and releasing an assembly every 1-2 weeks. The situation is further complicated by the fact that many of the organisms are relatively novel. There has been little prior characterization of their genomes, and even basic measures such as the genome size have large uncertainties associated with them.

For most organisms, the main bottleneck is the person-hours needed to make sense of the results. Even at the current level of sequencing throughput, it is no longer feasible to address this by simply assigning additional people. We describe the automated pre- and post-assembly quality control (QC) checks that we have implemented, to try to identify potential assembly problems as soon as possible, and to speed their resolution. We also discuss some of the initial results of these processes, e.g. that most eukaryotic sequencing projects have some amount of potentially symbiotic prokaryotic contamination associated with them.

## 69. Utilization and Characterization of cDNA and Protein from the Deep-Sea Thermal Vent Worm A. pompejana for Structural Biology

**David S. Shin\*** (davess@scripps.edu), <sup>1</sup> Michael DiDonato, <sup>2</sup> David P. Barondeau, <sup>3</sup> Greg Hura, <sup>3</sup> Chiharu Hitomi, <sup>3</sup> J. Andrew Berglund, <sup>3</sup> S. Craig Cary, <sup>3</sup> and John A. Tainer <sup>3</sup>

<sup>1</sup>Skaggs Institute for Chemical Biology and Scripps Research Institute, La Jolla, CA; <sup>2</sup>Protein Sciences Genomics Institute, Novartis Research Foundation, San Diego, CA; <sup>3</sup>Advanced Light Source, Lawrence Berkeley National Laboratory, Berkeley, CA; <sup>4</sup>Institute of Molecular Biology, University of Oregon, Eugene, OR; and <sup>5</sup>College of Marine Studies, University of Delaware, Lewes, DE

Structural biology is used to visualize protein conformations and complex interfaces. Three-dimensional protein structures not only aid in our understanding of protein function but also guide mutational analyses and drug design. Many key human protein sample preparations have proven difficult for atomic level structural analyses, particularly those for X-ray crystallography and small-angle X-ray scattering (SAXS) in solution, due to their inherent instability. Therefore, more stable proteins derived from thermophilic unicellular organisms are often characterized in lieu of their less stable counterparts. For example, we have used the archaeaons Pyrococcus furiosus and Archaeoglobus fulgidus for structure determination of fundamental DNA repair proteins, whose defects produce genomic instability phenotypes. Although we aim to extend our analyses of additional DNA repair proteins, as well as downstream signaling and cell-cycle control partners, many are missing from unicellular thermophiles. Alvinella pompejana, a worm that inhabits deep-sea hydrothermal vents, may provide a missing resource of thermostable eukaryotic proteins.

A. pompejana is a polychaetous annelid that inhabits the deep-sea thermal vents that extend along 9°N to 32°S within the axial summit caldera of the East Pacific Rise. Adult worms are found within self-constructed tubes in the hottest sections of black smoker chimneys. The mixing of iron- and sulfide-rich vent fluids with ambient seawater generates solid FeS (black smoke) within a local area that is nearly devoid of detectable oxygen. The water temperature in the worms' environment reaches up to 84°C and chemical reactions at the vents drive the pH of the water to ~5.5. The water also contains a toxic soup of heavy atoms (Ag, Cd, Co, Cu, Fe, Mg, Mn, Ni, Pb, Sr, V, Zn) at levels up to 1000x to that of ambient waters.

A. pompejana samples were collected at a depth of ~2.5 km (1.55 miles) using the deep sea submersible, DSV Alvin. An *in situ* stabilization chamber that bathes the worms in a nucleic acid stabilization solution was used to preserve mRNA. cDNA sets for sequencing and cloning recombinant genes were then generated.

Our preliminary validation of *A. pompejana's* utility for structural biology was initiated using copper zinc superoxide dismutase (SOD), whose human homolog is involved in familial amyotrophic lateral sclerosis or Lou Gehrig's disease, a very common neurological disorder that affects roughly 1 in 200,000 people. *A. pompejana's* SOD protein resulted in X-ray diffraction to 0.99 Å resolution using protein crystallography and also was stable enough to generate a *de novo* low resolution structure using SAXS. These results set the stage for *A. pompejana's* use in structural biology and formed the basis of our FY2005 CSP project proposal to sequence *A. pompejana* cDNA.

#### 70. Preliminary Analysis and Comparison of the Pseudomonas fluorescens Pf0-1 Genome with P. fluorescens Strains SBW25 and Pf5

**Mark W. Silby\*** (mark.silby@tufts.edu), Ana M. Cerdeño-Tárraga, and Stuart B. Levy

<sup>1</sup>Dept. of Molecular Biology and Microbiology, Tufts University School of Medicine, Boston, MA, and <sup>2</sup>Pathogen Sequencing Unit, Wellcome Trust Sanger Institute, Hinxton-Cambridge, UK

Pseudomonas fluorescens are a diverse group of bacteria commonly found in a wide range of terrestrial and aquatic habitats. Like other member of the genus *Pseudomonas*, they are metabolically versatile and highly adaptable. A number of P. fluorescens strains are being investigated for use in biocontrol or bioremediation. The DNA sequence of strain Pf0-1 was recently completed by the DOE Joint Genome Institute, and strain SBW25 has been sequenced at the Sanger Institute. We are in the process of annotation and preliminary analysis of these two genome sequences. The *P. fluorescens* genomes will provide us clues on how this species successfully colonizes a range of environments. Detailed comparison with the published *Pseudomonas* genomes should improve understanding of the differences between pathogenic and non-pathogenic members of the genus. It appears that *Pseudomonas* sp. share conserved 'core' genes, but significant differences exist in other genomic regions even between strains of the same species. For example, non-ribosomal peptide synthetases responsible for siderophore production have only limited similarity, possibly indicating functionally important domains. Our initial comparison aims to highlight some of these differences between Pf0-1, SBW25, and the published genome sequence of P. fluorescens Pf5. The Pf0-1 genome sequence and preliminary gene predictions can be found at http://genome.jgi-psf.org/finished\_microbes/psefl/psefl.home.html, and the SBW25 sequence with preliminary predictions can be found at http://www.sanger.ac.uk/Projects/P\_fluorescens.

### 71. Unlocking the Genomic and Ecophysiological Diversity of *Verrucomicrobia*

**Hauke Smidt\*** (hauke.smidt@wur.nl), <sup>1</sup> Erwin Zoetendal, <sup>1</sup> Muriel Derrien, <sup>1</sup> Caroline M. Plugge, <sup>1</sup> Peter H. Janssen, <sup>2</sup> and Willem M. de Vos <sup>1</sup> Laboratory of Microbiology, Wageningen University, Wageningen, The Netherlands, and <sup>2</sup>Dept. of Microbiology and Immunology, University of Melbourne, Victoria, Australia

Isolates of the bacterial phylum *Verrucomicrobia* have in common that they are specialized to the utilization of sugars. Their abundance and activity in a large variety of aquatic and terrestrial ecosystems, as well as in the intestine of vertebrates, indicates that this until recently unknown group of microorganisms plays a major role in the global carbon cycle. This calls for whole-genome sequencing of representatives of the six different monophyletic subdivisions. (*Chthoniobacter flavus*, *Opitutus terrae*, *Akkermansia muciniphila*, and soil isolate "Ellin514"), plus a deep evolutionary relative from the candidate phylum *Lentisphaerae* (*Victivallis vadensis*).

The Verrucomicrobia have until recently been overlooked by conventional cultivation-based approaches. The group is an excellent example of bacteria that are hardly detected by culturing but frequently found in 16S rRNA gene libraries from a wide variety of ecosystems, including aquatic and terrestrial environments as well as the mammalian gastrointestinal tract. On average, 5% of all surveyed sequences can be assigned to this phylum. In addition to being abundant, members of the phylum Verrucomicrobia also appear to be active members of the soil microbial community. They produce 1 to 9.8% of the bacterial 16S rRNA in soils. The recently obtained human intestinal isolates, Akkermansia muciniphila and Victivallis vadensis, have been identified as normal inhabitants of the human intestinal ecosystem. Using genus-specific 16S rRNA-targeted FISH analysis, Akkermansia muciniphila was found to have a mean abundance of 1% of the total fecal bacterial community. This interaction could be related to the ability of A. muciniphila to degrade mucin. However, only little or nothing is known about the roles of these bacteria, in terms of their metabolic properties and, in case of intestinal isolates, their interaction with the host environment.

During the past five years, the application of improved approaches for cultivation has yielded the isolation of several members of this phylum, now for the first time allowing for a thorough intra-phylum comparison. However, although this phylum is of similar phylogenetic depth and environmental relevance as the *Proteobacteria*, only 2 genome projects are currently ongoing. Therefore, we propose whole genome sequencing of representatives of 4 of the 6 monophyletic subdivisions, and a deep evolutionary relative from the candidate phylum *Lentisphaerae*.

The availability of a comprehensive set of genomes, which for the first time will allow for sound comparative genomic approaches to organisms from this phylum, can be expected to be of high relevance in the fields of taxonomy and evolution. Whole-genome sequencing of the proposed organisms will provide a wealth of information on their genetic potential and hence will allow for the further understanding and comparison of their metabolic capacities and interaction with their environments, including the mammalian host environment and GI tract microbiota, through comparative and functional genomics.

## 72. Toward a Comprehensive Overview of the *Bacillus* cereus Group Genomes

**Alexei Sorokin\*** (sorokine@jouy.unra.fr), Sandrine Auger, Nathalie Galleron, S. Dusko Ehrlich, Eugene Goltsman, Paul Richardson, and Alla Lapidus

<sup>1</sup>Génétique Microbienne, CRJ INRA, Jouy-en-Josas, France, and <sup>2</sup>DOE Joint Genome Institute, Walnut Creek, CA

The *Bacillus cereus* group consists of gram-positive, spore-forming bacteria with an impact on human activities due to their pathogenic properties. The most famous pathogens of animals and insects, *B. anthracis* and *B. thuringiensis*, carry their toxins in large plasmids. However, the bacteria of this group are also involved in mild food poisoning, causing rather noxious, although not fatal, vomiting or diarrhea. The emetic group strains appear to represent a very narrow phylogenetic group, and the corresponding toxin is encoded by a plasmid (Ehling-Schulz et al, 2005). Contrary to that, the characterized diarrheic toxins (Nhe, Hbl and CytK) are always present in the chromosome.

Three different strains were selected for complete sequencing, in addition to those reported earlier, to provide the comprehensive genomic characterization of the group. The work is being done as a collaboration of INRA (France), DOE Joint Genome Institute (Walnut Creek, USA) and CNS Génoscope (Evry, France). One of the strains, NVH398-91, is now completely sequenced. It was isolated from a severe food poisoning case in France (Lund et al, 2000). Surprisingly, this strain contains a gene for only one of known diarrheic toxins — CytK, which was shown to be synthesized in elevated amounts and to be particularly effective (Brillard and Lereclus, 2004; Fagerlund et al, 2004). The strain is rather different from other representatives of the *B. cereus* group and has a compact chromosome of 4.1 Mb, compared to 5.2-5.4 Mb genomes of other sequenced strains. This strain can also be regarded as the first bacterium of the group in which a highly effective toxin is chromosome-encoded. Divergence of the strain from others and its phenotypic particularities inspires to consider it as a representative of a new species, for which we propose a name *Bacillus cytotoxis*. However, only one such strain was described so far.

#### **Poster Presentations**

The second strain being sequenced, *Bwe* KBAB4, isolated from forest soil near Versailles (France), is a typical representative of recently recognized species — *B. weihenstephanensis* (Lechner et al, 1998; Sorokin et al, 2006). It contains a total of 5.6 Mb of genetic material, which includes genes to encode Nhe and Hbl, but lacks genes for CytK and HlyII. Although closely related strains are frequently reported to be food contaminants, they are in almost all cases not poisonous. However, the presence of Nhe and Hbl encoding genes suggests that such strains can also become toxic.

Finally, the third strain, *B. cereus* F0837/76, can be regarded as one of typical strains that can cause a severe diarrheic food poisoning. It synthesizes at least two characteristic toxins, Nhe and Hbl, in elevated amounts and was isolated from a contaminated surgical wound in a hospital (Turnbull et al, 1979; Beecher and MacMillan, 1990; Lund and Granum, 1997). Genomically the strain appeared to be very closely related to *B. anthracis* and this is probably the closest to it *B. cereus* strain known. It does not however contain the relevant pathogenic plasmids.

## 73. Methanosarcina barkeri Genome: Comparative Analysis with M. acetivorans and M. mazei Reveals Extensive Rearrangement within Methanosarcinal Genomes

**Kevin R. Sowers\*** (sowers@umbi.umd.edu)
University of Maryland Biotechnology Institute, Rockville, MD

### 74. *Hydra* as a Model Organism for Studying Evolution of Metazoan Genomes

Rob Steele\* (resteele@uci.edu)

Dept. of Biological Chemistry, University of California, Irvine, CA

A complete understanding of the evolution of metazoan genomes will require genome sequences from taxa at all levels of the metazoan phylogenetic tree. Of particular interest are organisms from such early-diverging phyla as Cnidaria, since they will be informative with regard to the nature of the earliest metazoan genomes. The cnidarian *Hydra* has been used as a model organism for experimental studies, particularly studies of developmental processes, for nearly 250 years. Genomic resources are now being developed for *Hydra*. These include a draft genome sequence that is nearing completion at the J. Craig Venter Institute and a dataset of approximately 150,000 ESTs. Analyses of these data have revealed a number of interesting evolutionary phenomena. As has been deduced from studies of EST datasets of limited size from two other cnidarians (*Acropora* and *Nematostella*), the *Hydra* data support the conclusion there has been substantial gene loss from the model bilaterians

Drosophila and C. elegans. Hydra carries out trans-spliced leader addition to a subset of its mRNAs, an interesting biological process and a useful one for identifying the 5' ends of mRNAs. Surprisingly, trans-spliced leader addition is absent from Acropora and Nematostella, suggesting that it was acquired relatively late during the cnidarian radiation. We have identified a *Hydra* gene which is a homologue of the *flp* genes in the parabasalid protist *Trichomonas vaginalis*. This gene has not been found in any other genome sequence or EST dataset, including the genome sequence of Nematostella, a member of the same phylum as Hydra. These results are consistent with lateral transfer of the flp gene from a parabasalid protist into *Hydra*. Using sequences from the *Hydra* genome project, we have assembled the region containing the flp gene. Immediately upstream of the flp gene is a gene encoding the 140 kDa subunit of replication factor C (RFC140). The distance between the end of the RFC140 gene and the beginning of the flp gene is about 400 nucleotides. This short distance and the fact that the Hydra flp mRNA undergoes trans-spliced leader addition suggests that the flp gene forms an operon with the RFC140 gene. Trans-splicing is predicted to facilitate functional incorporation of laterally transferred genes into the *Hydra* genome since it would allow expression of such genes without the need for them to have their own promoters. Recently, the Bosch lab at the University of Kiel has developed a method for generating stably transgenic *Hydra*. This will allow functional studies of the many interesting genes being identified from the *Hydra* EST and genome projects.

## 75. Comparative Genomic Analysis of Freshwater, Groundwater, and Marine *Caulobacter* Strains

**Craig Stephens\*** (cstephens@scu.edu) Santa Clara University, Santa Clara, CA

## 76. Functional Genomics and Thermal Biology of the Porcelain Crab, *Petrolisthes cinctipes*

Jonathon Stillman\* (stillmaj@sfsu.edu) San Francisco State University, San Francisco, CA

## 77. Roseobacter denitrificans: A Ubiquitous Marine Phototroph with a Peculiar Carbon Fixation Pathway

Wesley D. Swingley\* (wswingley@msn.com), Sumedha Gholba, Steven D. Mastrian, Heather J. Matthies, Jicheng Hao, Hector Ramos, Chaitanya R. Acharya, Amber L. Conrad, Heather L. Taylor, Liza C. Dejesa, Maulik K. Shah, Maeve E. O'Huallachain, Michael T. Lince, Robert E. Blankenship, J. Thomas Beatty, and Jeffrey W. Touchman School of Life Sciences, Arizona State University, Tempe, AZ; Dept. of Computational Biosciences, Arizona State University, Tempe, AZ; Translational Genomics Research Institute, Phoenix, AZ; Dept. of Chemistry and Biochemistry, Arizona State University, Tempe, AZ; and University of British Columbia, Vancouver, BC

Purple aerobic phototrophic bacteria (APBs) are the only bacteria that perform photosynthesis in the presence of oxygen, but do not produce oxygen as a result. These highly adaptive APB compose greater than 10% of the microbial community in some euphotic upper ocean waters and are potentially major contributors to the fixation of the greenhouse gases CO and CO<sub>2</sub>. We describe genomic sequence features of the APB *Roseobacter* denitrificans that reveal clues to its physiology. The genome does not contain any genes that code for known carbon-fixation pathways and most notably missing is a gene for the Calvin cycle enzyme RubisCO. However, we describe the presence and potential importance of a mixotrophic CO<sub>2</sub>-fixation pathway analogous to the C<sub>4</sub> cycle found in many plants. We suggest that this pathway functions to fix much of the CO<sub>2</sub> for the formation of cellular components, but does not allow for autotrophic growth. While some genes that code for the redox-dependent regulation of photosynthetic machinery are present, some light sensors and transcriptional regulatory motifs found in other anoxygenic photosynthetic bacteria are absent.

## 78. Novel Proteins from an Acidic, Hot, Metal-Rich Ecosystem

Michael P. Thelen\* (mthelen@llnl.gov), <sup>1</sup> Jill Banfield, <sup>3</sup> Steven Singer, <sup>1</sup> Christopher Jeans, <sup>1</sup> Mona Hwang, <sup>1</sup> Nathan VerBerkmoes, <sup>2</sup> Manesh Shah, <sup>2</sup> Robert Hettich, <sup>2</sup> Clara Chan, <sup>3</sup> Vincent Denef, <sup>3</sup> and Gene Tyson <sup>3</sup> <sup>1</sup> Biosciences Directorate, Lawrence Livermore National Laboratory, Livermore, CA; <sup>2</sup> Chemical Sciences Division, Oak Ridge National Laboratory, Oak Ridge, TN; and <sup>3</sup> Dept. of Earth and Planetary Sciences, University of California, Berkeley, CA

Although a large portion of proteins deduced from microbial genomes do not match any annotated sequences, it is likely that many of these "hypothetical proteins" are important, in some cases essential, for microbial fitness under natural conditions. The void in our knowledge regarding this dark side of genomics, representing 30 – 40% of protein sequences, is a formidable obstacle to nearly every investigation enabled by full genome sequence information. In our study of acidophilic biofilm communities, we are focused directly on the problem of hypothetical proteins within a well-defined system in one of the most extreme limits of habitability. These microbes assemble in very acidic streams of underground mine tunnels to form floating, matrix-bound, self-sustaining communities that derive electrons from iron (II) while in turn catalyzing the dissolution of pyrite (FeS<sub>2</sub>). Such conditions result in the biological generation of acid mine drainage (AMD), ultimately causing many orders of magnitude greater metal dissolution and environmental acidification than abiotic forces alone. Since AMD is so prevalent and potentially disastrous to U.S. inland waters associated with principle energy resources such as coal and uranium, understanding the mechanisms by which biofilm communities generate AMD is a priority with DOE's Office of Biological and Environmental Research.

In addition to the detailed geochemical and microbiological description of AMD biofilms, the foundation of our current investigation rests on both genomic and proteomic datasets for each of several dominant organisms. Characterization of abundant species in one such biofilm and the associated genome analyses resulted in reconstruction of near complete or partial genomes for two different Leptospirillum bacteria and three archaea, and further work at the JGI is now extending the sequences within this biofilm and also to ones with differing population structures. In our attempt to understand essential biofilm metabolic activities and the partitioning of functions between different organism types, proteins native to an environmental biofilm were analyzed by shotgun MS proteomics. This confirmed that many hypothetical proteins are expressed at detectable levels in the major biofilm organisms, and also indicated that a majority of abundant extracellular proteins are novel.<sup>2</sup> This groundwork has enabled us to examine proteins expressed in biofilms harvested from several neighboring locations, in carefully measured geochemical conditions, using a combination of chromatographic techniques, MS identification and computational analyses/predictions.<sup>3</sup> These methods provide us with a powerful approach to isolate many of the novel proteins along with any known proteins associated with them, and to begin assigning functions using biochemical assays. Several key enzymatic activities that are suspected to play important roles in biofilm formation and function, such as hydrolases, oxidoreductases and polysaccharide synthases, will be examined in parallel. The development of this type of systematic approach for the validation and description of hypothetical proteins, used within a defined biological system, is a test case for the full utility of proteogenomic information.

- 1. Tyson et al, 2004, Nature 428:37-43.
- 2. Ram et al, 2005, Science 308:1915-20.
- 3. Banfield et al, 2005, OMICS 9:301-33.

#### 79. The Hox Cluster of a Model Chelicerate Arthropod— The Oribatid Mite *Archegozetes longisetosus*

**R. H. Thomas\*** (rthomas@zoology.siu.edu), <sup>1,2</sup> J. A. Mackenzie-Dodds, <sup>2</sup> H. M. Fourcade, <sup>3</sup> and J. L. Boore <sup>3</sup>

<sup>1</sup>Dept. of Zoology, Southern Illinois University, Carbondale, IL; <sup>2</sup>Dept. of Zoology, Natural History Museum, London, UK; and <sup>3</sup>Evolutionary Genomics Program, DOE Joint Genome Institute, Walnut Creek, CA

The arthropods are the most diverse group of animals and are usually divided into four major groups: insects, crustaceans, myriapods (centipedes, millipedes and others), and chelicerates (horseshoe crabs and arachnids, which contain mites, spiders, scorpions and other eight-legged animals). Of these chelicerate groups, mites are by far the most diverse, both in numbers of species and in ecological breadth. Here we report on the sequence of the first chelicerate arthropod Hox cluster from a freeliving oribatid mite, Archegozetes longisetosus. We have demonstrated that the mite homologs of the Drosophila genes zen and ftz both behave as Hox genes in this mite, lending support to the notion that the ancestral arthropod Hox cluster contained ten genes. The Archegozetes homolog of abdominal-A is not in its usual position between the mite homologs of Ultrabithorax and Abdominal-B. All attempts to locate an abdominal-A homolog elsewhere in the genome have failed. We conclude that abd-A is lost in this animal — the first verified loss of a Hox gene reported in an arthropod. There are no obvious simplifications or losses in the mite opisthosoma that suggest an immediate explanation for the loss of abdominal-A.

## 80. Mining of Switchgrass Microsatellites from EST Data for Mapping and Diversity Analysis

**Christian M. Tobias\*** (ctobias@pw.usda.gov)

U.S. Department of Agriculture, Agricultural Research Service

### 81. Plant-Plant Interactions: Parasitic Plant Responses in Host Plant Interactions

**Alexey Tomilov**, <sup>1</sup> Natalya Tomilova, <sup>1</sup> Manuel Torres, <sup>2</sup> Russell Reagan, <sup>1</sup> Tatiana Fillapova, <sup>1</sup> Pradeepa Gunathilake, <sup>1</sup> and **John I. Yoder**\* <sup>1</sup> (jiyoder@plantsciences.ucdavis.edu)

<sup>1</sup>Dept. of Plant Sciences, University of California, Davis, CA, and <sup>2</sup>Center for Applied Genetic Technologies, University of Georgia, Athens, GA

In natural environments roots of different plants grow closely entwined and frequently contact. When roots of parasitic plants in the Orobanchaceae contact roots of neighboring plants, they develop haustoria which attach to and invade host root tissues. When mature, haustoria connect parasite and host roots and provide the physiological conduit through which host resources are translocated into the parasite. The consequences of parasitism can be dire for the host plant and some of the world's worst agricultural pests are parasitic weeds. We are identifying genes that encode early haustorium development in parasitic Orobanchaceae in order to 1) discover the genetic basis for the origin(s) of parasitism in plants; 2) determine how parasitic plants identify and discriminate hosts, and; 3) develop strategies for engineering crop plants for genetic resistance against parasitic weeds.

*Triphysaria* is a parasitic plant that grows as a common springtime wildflower in coastal bluffs, serpentine slopes and grassland fields throughout the Pacific Coast of North America. *Triphysaria* is closely related to the internationally devastating weeds *Striga* and *Orobanche* and provides an experimentally tractable model for these aggressively invasive, highly restricted, agricultural pests. *Triphysaria* develop knob shaped haustoria at their root tips when seedlings are treated with host (*Arabidopsis*) roots, host root exudates, or purified root factors. Under these in vitro conditions haustorium development is robust, rapid and highly synchronous.

We used suppression subtractive hybridization (SSH) to enrich cDNA libraries for transcripts regulated in *Triphysaria* roots within a few hours after exposure to different haustorium inducing treatments. About ten thousand SSH products have been sequenced to date with an addition twenty thousand SSH products and thirty thousand full length cDNAs in the JGI sequencing pipeline. Using a combination of bioinformatics and cDNA arrays we showed that many of the genes expressed early in haustorium development are similarly regulated when exposed to cytotoxins released by allelopathic plants, even when the allelochemicals are inactive as haustoria inducers. Protein annotations suggest that many of these genes encode xenobiotic detoxification enzymes. A significant fraction of the parasite transcriptome regulated by host contact is predicted to function in electron or chemical transport. Two distinct quinone oxidoreductases regulated in *Triphysaria* roots by host contact have been expressed in E. coli, purified and characterized. Gene transfer technologies have been developed for Triphysaria roots and these are being used to determine the function of these and other candidate parasite genes. Our results suggest that host recognition involves a redox associated signal transduction system recruited in evolution from genes functioning in rhizosphere chemical detoxification.

## 82. Genome Sequence Comparisons of the Eukaryotic Plant Pathogens *Phytophthora sojae* and *Phytophthora ramorum*

**Sucheta Tripathy\*** (stripath@vt.edu)

Virginia Polytechnic University and State University, Blacksburg, VA

#### 83. Progress on the Human Gut Microbiota Project

**Parag Vaishampayan\*** (pavaishampayan@lbl.gov), <sup>1</sup> Jenna Morgan, <sup>1</sup> Jeff Froula, <sup>1</sup> Yogesh Shouche, <sup>2</sup> Howard Ochman, <sup>3</sup> and M. Pilar Francino <sup>1</sup> Evolutionary Genomics Department, DOE Joint Genome Institute, Walnut Creek, CA; <sup>2</sup>National Centre for Cell Science, University of Pune, India; and <sup>3</sup>Departments of Biochemistry and Molecular Biophysics, and Ecology and Evolutionary Biology, University of Arizona, Tucson, AZ

The diverse community of gut microbes plays an essential role in human health, contributing to the digestive process, promoting gut maturation, modulating the immune system, and interacting with pathogens in several complex ways. We have initiated efforts to study complex gut microbial community structure and their coding capabilities.

Human fecal samples from two healthy anonymous volunteers (mother and infant) were collected at the University of Arizona in 2004 and 2005, in collaboration with the laboratory of Howard Ochman. High molecular weight DNA for large insert library construction was prepared from these samples. From these preparations, we have produced 4 different fosmid libraries, two from adult samples (2004 and 2005) and two infant samples (2004, 1 month old and 2005, 11 months old), each containing 50,000 to 70,000 clones (40 kb insert size). We have also prepared 16s libraries from the DNA agarose plugs from which the fosmid libraries were prepared.

We have initiated screenings of the fosmid libraries with 16s rRNA primers as well as a set of 11 primer sets directed towards universally conserved bacterial proteins (Santos and Ochman 2004) for phylogenetic analysis. Currently we are optimizing protocols to avoid amplification of the host *E. coli* strain genes. The different alternative protocols include fosmid induction, effective fosmid purification, use of DNA exonucleases and use of terminator primers to abort PCR extension of *E. coli* genes.

We aim to further characterize these libraries by screening different types of genes involved in microbe-host interactions. We will perform phenotypic screenings to identify clones producing antibiotic molecules potentially involved in competition between the microbial community members. The information on phylogenetic diversity, coding capabilities related to commensalism and pathogenicity and antibiotic production will provide an ample biological background describing the GI microbiotic community.

In future we have plans to study succession and development of gut microbiota of new born infant as collaborative efforts with a research institution in India. To clarify the initial acquisition and subsequent colonization of bacteria in an infant within the few months after birth, phylogenetic analysis will be performed using 16S rDNA sequences from the DNA isolated from feces. In addition, fosmid libraries will be prepared from these DNA samples and comparative analyses will be performed between the American and Indian microbial succession data.

## 84. A Unique Diversity of Cytochrome C-Dependent Dehydrogenases Found in the *Pantoea citrea* Genome

**Fernando Valle\*** (fvalle@genencor.com), Mai Du, Bob Caldwell, Harun Rashid, and Tim Dodge

Genencor International/Danisco, Palo Alto, CA

Some bacteria utilize the extracellular oxidation of substrates as a metabolic strategy to generate membrane potential. The best known microorganism with this type of metabolism is *Gluconobacter oxidans*, who is capable of oxidizing several carbohydrates using six periplasmic dehydrogenases. Two of these dehydrogenases contain a cytochrome C subunit.

We have developed a very efficient fermentation process to produce 2-keto-L-gulonate (2-KLG) from glucose using the bacteria *Pantoea citrea* as the production host. 2-KLG is an intermediate for the synthesis of Ascorbic acid.

To understand the potentials and limitations of *P. citrea* as a production host, and to directed some of our Metabolic Engineering efforts, its genome was sequenced.

Genome analysis showed that *P. citrea* possesses an array of genes to modify carbohydrates extracellularly. Among them, 19 operons containing cytochrome c-dependent extracellular dehydrogenases were found. So far, *P. citrea* is the only bacteria possessing this astonishing number of paralogs. An analysis of these 19 operons will be presented.

# 85. Dinoroseobacter shibae, A New Aerobic Phototrophic Bacterium from the Roseobacter Lineage with a Complex Genome Structure Producing Long-Chain Acylated Homoserine Lactones

**Irene Wagner-Döbler\*** (iwd@gbf.de), <sup>1</sup> Silke Pradella, <sup>2</sup> Brian Tindall, <sup>2</sup> Rüdiger Pukall, <sup>2</sup> Stefan Schulz, <sup>3</sup> Hanno Biebl, <sup>1</sup> and Martin Allgaier <sup>4</sup> <sup>1</sup>GBF, Braunschweig, Germany; <sup>2</sup>German Collection for Microorganisms and Cell Cultures, Braunschweig, Germany; <sup>3</sup>Technical University, Braunschweig, Germany; and <sup>4</sup>Leibniz-Institute for Aquatic Ecology, Neu-Globsow, Germany

First described 15 years ago, the genus *Roseobacter* has become the core of a group of marine bacteria which are the focus of intense research because of their taxonomic diversity and abundance in the world's oceans as well as their important contribution to the global carbon and sulfur cycles and thus to climate change. Dinoroseobacter shibae is a novel genus within this group that was isolated from toxic marine dinoflagellates<sup>2</sup> and is able to perform aerobic anoxygenic photosynthesis, a way of generating energy using bacteriochlorophyll a which contributes up to 10% to the primary production in the ocean and is presently poorly understood. Dinoroseobacter shibae carries the pufLM genes of the photosynthesis reaction center and has fully functional light harvesting complexes I and II including bacteriochlorophyll a synthesis<sup>4</sup>. Dinoroseobacter shibae has a complex genome, harbouring seven linear plasmids which are most probably conjugative and together comprise 861 kb of extrachromosomal information<sup>3</sup>. Finally, *Dinoroseobacter shibae* has been found to produce highly unusual quorum sensing signalling compounds, i.e. acylated homoserine lactones (AHLs) with side chain lengths of up to 18 carbon atoms, the longest AHL side chain length presently known. Three different AHL signals have been discovered by their ability to stimulate a luxR-type reporter strain, as well as by GC-MS analysis of concentrated culture supernatants resulting in structure elucidation, namely C8-HSL, C18-en-HSL and C18-dien-HSL<sup>1</sup>. It is conceivable that some of the genetic determinants, e.g. the luxR type receptor and the luxI type synthase genes, might be located on the many linear plasmids discovered in this organism, from where they could easily be transferred to related species.

The genome sequence of *Dinoroseobacter shibae* will make it possible to unravel the mechanisms of aerobic anoxygenic photosynthesis and cell density dependent gene expression and thus help to understand the adaptations of this organism to its ecological niche in the ocean.

- 1. Wagner-Döbler et al. (2005) Discovery of complex mixtures of novel longchain quorum sensing signals in free-living and host-associated marine Alphaproteobacteria. ChemBioChem 6(12):2195-206.
- 2. Biebl et al. (2005) *Dinoroseobacter shibae*, gen. nov., sp. nov., a new aerobic phototrophic bacterium isolated from dinoflagellates. Int. J. System. Evol. Microbiol. **55**: 1089 1096.

- 3. Pradella et al. (2004) Genome Organization and Localization of the *pufLM* Genes of the Photosynthesis Reaction Center in Phylogenetically Diverse Marine *Alphaproteobacteria*. Appl. Environ. Microbiol. **70**:3360-3369.
- 4. Allgaier et al. (2003) Aerobic anoxygenic photosynthesis in *Roseobacter* clade bacteria from diverse marine habitats. Appl. Environ. Microbiol. **69**:5051-5059.

## 86. Genome Sequence of the Cellulolytic Gliding Bacterium *Cytophaga hutchinsonii*

**G. Xie**<sup>1,2</sup>, D. C. Bruce<sup>1,2</sup>, J. F. Challacombe<sup>1,2</sup>, O. Chertkov<sup>1,2</sup>, P. Gilna<sup>1,2</sup>, C. S. Han<sup>1,2</sup>, B. Henrissat<sup>3</sup>, S. Lucas<sup>2,4</sup>, M. Misra<sup>1</sup>, G. L. Myers<sup>1</sup>, P. Richardson<sup>2,5</sup>, R. Tapia<sup>1,2</sup>, N. N. Thayer<sup>1</sup>, L. S. Thompson<sup>1,2</sup>, T. S. Brettin<sup>1,2</sup>, and **M. J. McBride\*** (mcbride@uwm.edu)<sup>6</sup>

<sup>1</sup>Los Alamos National Laboratory, Los Alamos, NM; <sup>2</sup>DOE Joint Genome Institute, Walnut Creek, CA; <sup>3</sup>CNRS and Universites Aix-Marseille I & II, Marseille, France; <sup>4</sup>Lawrence Livermore National Laboratory, Livermore, CA; <sup>5</sup>Lawrence Berkeley National Laboratory, Berkeley, CA; <sup>6</sup> University of Wisconsin, Milwaukee, WI

We present the complete DNA sequence and metabolic analysis of the aerobic cellulolytic Gram-negative soil bacterium Cytophaga hutchinsonii, which belongs to the phylum *Bacteroidetes* (also known as the Cytophaga-Flavobacterium-Bacteroides group). The genome consists of a single circular 4.43 Mb chromosome containing 3805 open reading frames (ORFs), with 37% GC content, of which 1969 ORFs (52%) have been assigned a tentative function. Two of the most striking characteristics of C. hutchinsonii are its rapid gliding motility over surfaces and its contact dependent digestion of crystalline cellulose. The mechanism of C. hutchinsonii motility is not known but its genome contains homologs for each of the gld genes that are required for gliding of the distantly related bacteroidete Flavobacterium johnsoniae. Cytophaga-Flavobacterium gliding appears to be novel and does not involve wellstudied motility organelles such as flagella or type IV pili. Many genes thought to encode proteins involved in cellulose utilization were identified. These include putative endo-1, 4-β-glucanases and βglucosidases. Surprisingly, genes encoding obvious exoglucanases were not detected. Since such enzymes are usually needed for efficient cellulose digestion C. hutchinsonii either has novel exoglucanases or has an unusual method of cellulose utilization. Genes encoding typical cellulosome components were absent but the genome appears to encode 74 very-highmolecular-weight proteins that may be part of the cell-bound cellulolytic machinery. Most of these are predicted to be extracellular or cell surface proteins and many have putative polysaccharide lytic D5 domains. 184 transporters have been identified, but PTS systems are absent. Biosynthetic pathways for all common amino acids, nucleotides, vitamins and coenzymes appear to be present. Pathways needed for oxidative respiration of glucose appear intact and a modified oxidative branch of the pentose phosphate pathway is also predicted.

#### 87. Origin and Evolution of Vertebrate Neural Crest Gene Network—Insights from Comparative Genomics Analysis of *Amphioxus* ESTs

**Jr-Kai Sky Yu\*** (jkyu@caltech.edu), <sup>1</sup> Yutaka Satou, <sup>2</sup> Tadasu Shin-I, <sup>3</sup> Yuji Kohara, <sup>3</sup> Nori Satoh, <sup>2</sup> Linda Z. Holland, <sup>4</sup> and Marianne Bronner-Fraser<sup>1</sup>

<sup>1</sup>Division of Biology, California Institute of Technology, Pasadena, CA; <sup>2</sup>Dept. of Zoology, Graduate School of Science, Kyoto University, Kyoto, Japan; <sup>3</sup>National Institute of Genetics, Shizuoka, Japan; and <sup>4</sup>MBRD, Scripps Institution of Oceanography, University of California, San Diego, CA

The neural crest is a cell type unique to vertebrates that arises in the embryo from the border of the neural plate and adjacent non-neural ectoderm. Neural crest cells undergo an epithelial/mesenchymal transition to become a migratory population of cells that gives rise to numerous differentiated cell types, including the craniofacial skeleton and much of the peripheral nervous system. Evolution of the neural crest is thought to be responsible for formation of many of the complex structures of the vertebrate head that are lacking in invertebrate chordates. The cephalochordate, *amphioxus*, is an evolutionary model for understanding the origin of vertebrates since its embryonic body plan is very vertebratelike. We have taken a comparative approach using a large scale EST analysis to identify *amphioxus* homologs of vertebrate genes involved expressed at the neural plate border in a putative gene regulatory network. Our preliminary survey shows that while the early stages of neural plate border patterning genes appear conserved between amphioxus and vertebrates, later deployment of some transcription factors at the neural plate border and presumptive neural crest region seems to be a vertebrate innovation. Such an understanding of the evolution of gene regulatory events gives important insights into how developmental genes may have been co-opted for new roles in patterning the neural crest as a novel vertebrate innovation.

## 88. Metagenomic Analysis of Microbial Communities in Uranium-Contaminated Groundwaters

**Jizhong Zhou\*** (jzhou@rccc.ou.edu),<sup>1,6</sup> Terry Gentry,<sup>1</sup> Chris Hemme,<sup>1,6</sup> Liyou Wu,<sup>1</sup> Matthew W. Fields,<sup>2</sup> Chris Detter,<sup>3</sup> Kerrie Barry,<sup>3</sup> David Watson,<sup>1</sup> Christopher W. Schadt,<sup>1</sup> Paul Richardson,<sup>3</sup> James Bristow,<sup>3</sup> Terry Hazen,<sup>4</sup> James Tiedje,<sup>5</sup> and Eddy Rubin<sup>3</sup>

<sup>1</sup>Environmental Sciences Division, Oak Ridge National Laboratory, Oak Ridge, TN; <sup>2</sup>Dept. of Microbiology, Miami University, Oxford, OH; <sup>3</sup>DOE Joint Genome Institute, Walnut Creek, CA; <sup>4</sup>Earth Science Division, Lawrence Berkeley National Laboratory, Berkeley, CA; <sup>5</sup>Center for Microbial Ecology, Michigan State University, East Lansing, MI; and

<sup>6</sup>Institute for Environmental Genomics, Dept. of Botany and Microbiology, University of Oklahoma, Norman, OK

Due to the uncultivated status of the majority of microorganisms in nature, little is known about their genetic properties, biochemical functions, and metabolic characteristics. Although sequence determination of the microbial community 'genome' is now possible with high throughput sequencing technology, the complexity and magnitude of most microbial communities make meaningful data acquisition and interpretation difficult. Therefore, we are sequencing groundwater microbial communities with manageable diversity and complexity (10-400 phylotypes) at the U.S Department of Energy's Natural and Accelerated Bioremediation Research (NABIR)-Field Research Center (FRC), Oak Ridge, TN. The microbial community has been sequenced from a groundwater sample contaminated with very high levels of nitrate, uranium and other heavy metals and pH 3.7. Sequence analysis of this groundwater sample based on a 16S rDNA library revealed 10 operational taxonomic units (OTUs) at the 99.6% cutoff with >90% of the OTUs represented by an unidentified y-proteobacterial species related to the genus Frateuria. Additional OTUs were related to a J3-proteobacterial species of the genus Azoarcus. Three clone libraries with different DNA fragment sizes (3, 8 and 40 kb) were constructed, and 70 Mb raw sequences were obtained using a shotgun sequencing approach. The raw sequences were assembled into 2770 contigs totaling 6 Mb which were further assembled into 224 scaffolds (1.8 kb-2.4 Mb). Preliminary binning of the scaffolds suggests 4 primary phylotypes (FRC Gamma Groups I & II (Frateuria), FRC Beta Group I (Burkholderia) and FRC Beta Group II (Herbaspirillum), with the FRC Gamma Group I representing the dominant phylotype (1 IX genome coverage). Genes identified from the sequences were consistent with the geochemistry of the site, including multiple nitrate reductase and metal resistance genes. Despite the low *species* diversity of the samples. evidence of strain diversity within the identified species was observed. Analysis with functional gene arrays containing 23,000 probes designed based on these community sequences as well as genes important for biogeochemical cycling of C, N, and S, along with metal resistance and contaminant degradation suggested that the dominant species could be biostimulated during *in situ* uranium reduction experiments at the FRC. These results also suggest that the dominant species could play a direct or indirect role in the bioremediation of uranium.

#### 89. "Omics" of Dehalococcoides spp.

Stephen H. Zinder\* (shz1@cornell.edu)

Dept. of Microbiology, Cornell University, Ithaca, NY

Dehalococcoides spp. are members of the Chloroflexi that can reductively dechlorinate diverse chloroorganics including the solvents tetrachloroethene (PCE) and trichloroethene (TCE), chlorobenzenes, and PCBs. The genome of the PCE-dechlorinating D. ethenogenes strain 195 was 1.47 Mb and encoded 17 predicted reductive dehalogenases (RDHs). The genome of the chlorobenzene dechlorinating *Dehalococcoides* strain CBDB1 was 1.40 Mb and encoded 32 predicted RDHs. There is near complete synteny between the two strains in regions encoding "housekeeping" genes, whereas regions encoding RDHs show signs of genetic exchange and rearrangement. Two more *Dehalococcoides* genome sequences are being determined by JGI. Transcriptional analyses of RDH expression in D. ethenogenes grown on PCE or TCE by qPCR showed high expression of DET0079 and DET0318, predicted to encode TCE RDH and PCE RDH respectively. Highly expressed oxidoreductases included the Hup, Vhu, and Hym hydrogenases and an ORF predicted to encode a formate dehydrogenase (FDH). Tandem mass spectroscopy of tryptic peptides generated from membrane enriched fractions from D. ethenogenes showed high coverage of the RDHs and oxidoreductases described above. The high expression of the predicted FDH was surprising since D. ethenogenes does not utilize formate. Examination of the predicted amino acid sequence revealed a serine encoded in place of cysteine or selenocysteine at a critical residue. Also showing high peptide coverage was DET1407, and analysis of this protein suggests it is part of the S-layer cell wall in D. ethenogenes. These studies show that Dehalococcoides spp. are highly evolved to use halogenated compounds and demonstrate the ability of "omics" to shed light on organisms difficult to study in the laboratory.

Current as of March 10, 2006

Catherine C. Adley University of Limerick catherine.adley@ul.ie

Andrea Aerts

DOE Joint Genome Institute

alaerts@lbl.gov

Nina Agabian

University of California, San Francisco

nina.agabian@ucsf.edu

Eric E. Allen

University of California, Berkeley eallen@nature.berkeley.edu

Carl W. Anderson

**Brookhaven National Laboratory** 

cwa@bnl.gov

Anne J. Anderson Utah State University anderson@biology.usu.edu

Iain J. Anderson

DOE Joint Genome Institute

IJAnderson@lbl.gov

Brett J. Baker

University of California, Berkeley

bbaker@eps.berkeley.edu

Scott E. Baker

Pacific Northwest National Laboratory

scott.baker@pnl.gov

Jody Banks Purdue University banksj@purdue.edu

Sebastian F. Behrens Stanford University sbehrens@stanford.edu

Harry R. Beller

Lawrence Livermore National Laboratory

beller2@llnl.gov Randy M. Berka

Novozymes, Inc.

ramb@novozymes.com

Stephan R. Berosik Applied Biosystems

berosisr@appliedbiosystems.com

Alison M. Berry

University of California, Davis

amberry@ucdavis.edu

Diana L. Berry

Stony Brook University diberry@optonline.net

Ilka Bischofs

University of California, Berkeley

ibbischofs@lbl.gov

Hans J. Bohnert

University of Illinois, Urbana-Champaign

bohnerth@life.uiuce.edu

Harvey Bolton

Pacific Northwest National Laboratory

harvey.bolton@pnl.gov

Jeffrey L. Boore

DOE JGI, Lawrence Berkeley National

Laboratory

JLBoore@Berkeley.edu

Chris Bowler

Ecole Normale Superieure cbowler@biologie.ens.fr

Elbert W. Branscomb

Lawrence Livermore National Laboratory

branscomb1@llnl.gov

Lambert Brau

Murdoch University

lbrau@murdoch.edu.au

Jim Bristow

DOE Joint Genome Institute

JBristow@lbl.gov

Fred Brockman

Pacific Northwest National Laboratory

Fred.Brockman@pnl.gov

Eoin Brodie

Lawrence Berkeley National Laboratory

elbrodie@lbl.gov

Shane Brubaker

DOE Joint Genome Institute shanebrubaker@sbcglobal.net

Donald A. Bryant

The Pennsylvania State University

dab14@psu.edu

Andrew Cameron

California Institute of Technology

acameron@caltech.edu

Patrick Chain

Lawrence Livermore National Laboratory

chain2@llnl.gov

Cathy Chang Diversa Corporation cchang@diversa.com

Quen J. Cheng

Lawrence Berkeley National Lab

QJCheng@lbl.gov Penny Chisholm

Massachusetts Institute of Technology (MIT)

chisholm@MIT.edu
Ludmila Chistoserdova
University of Washington

milachis@u.washington.edu

Dylan Chivian

Lawrence Berkeley National Laboratory

DCChivian@lbl.gov

Richard M. Clark

Max-Planck-Institute for Developmental

Biology

richard.clark@tuebingen.mpg.de

John K. Colbourne Indiana University jcolbour@cgb.indiana.edu

Matthew Coleman

Lawrence Livermore National Laboratory

coleman16@llnl.gov

Lorenza Conterno

Cornell University, N.Y. State Agriculture

Experiment Station lc98@cornell.edu

Luis M. Corrochano Universidad de Sevilla corrochano@us.es

Adrienne M. Cottrell University of Georgia acottrel@uga.edu

Daniel Cullen

University of Wisconsin, Madison dcullen@facstaff.wisc.edu

Christopher Daum

DOE Joint Genome Institute

daum1@llnl.gov

Charles N. David

Ludwig Maximilian University, Munich david@zi.biologie.uni-muenchen.de

Seana K. Davidson University of Washington skdavid@u.washington.edu Scott C. Dawson

University of California, Berkeley sdawson@nature.berkeley.edu

Anthony De Tomaso Stanford University tdet@stanford.edu

Paramvir S. Dehal

DOE Joint Genome Institute

psdehal@lbl.gov

Edward F. DeLong

Massachusetts Institute of Technology

delong@mit.edu

Vincent J. Denef

University of California, Berkeley

vdenef@berkeley.edu

Michael K. DeSalvo

University of California, Merced mdesalvo@ucmerced.edu

Patrik D'haeseleer

Lawrence Livermore National Laboratory

patrikd@llnl.gov

Ilenys Diaz-Muniz USDA-ARS at NCSU idiazmu@ncsu.edu

Larry J. Dishaw

All Children's Hospital, H. Lee Moffitt Cancer

Center and Research Institute

ldishaw@hsc.usf.edu

Daniel W. Drell

U.S. Department of Energy daniel.drell@science.doe.gov

Nicole Dubilier

Max Planck Institute for Marine Microbiology

ndubilie@mpi-bremen.de

Jennifer L. DuBois University of Notre Dame jdubois@nd.edu

Serge Dusheyko

DOE Joint Genome Institute

sdusheyko@lbl.gov

Joseph Ecker

University of California, San Diego

ecker@salk.edu

Eric Edsinger-Gonzales

University of California, Berkeley

eegonzales@berkeley.edu

Michael Egholm 454 Life Sciences megholm@454.com

Jonathan A. Eisen

The Institute for Genomic Research

jonathan eisen@mac.com

Drew Endy

Massachusetts Institute of Technology (MIT)

endy@mit.edu

Marsha Fenner

DOE Joint Genome Institute

MWFenner@lbl.gov

Matthew W. Fields Miami University fieldsmw@muohio.edu

Peg Folta

Lawrence Livermore National Laboratory

pfolta@llnl.gov

Thomas Fowler

Southern Illinois University, Edwardsville

tfowler@siue.edu

Maria Pilar Francino DOE Joint Genome Institute

mpfrancino@lbl.gov

Marvin E. Frazier
J. Craig Venter Institute
mfrazier@venterinstitute.org

Lillian Fritz-Laylin

University of California, Berkeley

Cande Laboratory fritz-laylin@berkeley.edu

Jeff L. Froula

DOE Joint Genome Institute

jlfroula@lbl.gov

Jill O. Fuss

Lawrence Berkeley National Laboratory

jfuss@lbl.gov

Eric Gaidos

University of Hawaii gaidos@hawaii.edu

David E. Gilbert

DOE Joint Genome Institute

gilbert21@llnl.gov

Paul Gilna

Los Alamos National Laboratory

pgil@lanl.gov

Tijana Glavina Del Rio DOE Joint Genome Institute

glavinadelrio1@llnl.gov

Frank O. Gloeckner

Max Planck Institute for Marine Microbiology

fog@mpi-bremen.de

Bernardo Gonzalez

Facultad de Ciencias Biologicas. P. Universidad

Catolica de Chile bgonzalez@bio.puc.cl

Stephen B. Goodwin

USDA-ARS and Purdue University

sgoodwin@purdue.edu

Joe W. Gray

Lawrence Berkeley National Laboratory

JWGray@lbl.gov

Igor V. Grigoriev

DOE Joint Genome Institute

ivgrigoriev@lbl.gov

Arthur Grossman

Carnegie Institution of Washington

arthurg@stanford.edu

Steven J. Hallam

Bacterial Adaptation and Response Network

shallam@mit.edu

Cliff S. Han

Los Alamos National Laboratory

han\_cliff@lanl.gov

Emily E. Hare

U.C. Berkeley and LBNL ehare@berkeley.edu

Richard Harland U.C. Berkeley

harland@berkeley.edu

Jonathan K. Harris University of Colorado jjharris@colorado.edu

Loren J. Hauser

Oak Ridge National Laboratory

hauserlj@ornl.gov

Dennis Hedgecock

University of Southern California

dhedge@usc.edu

Christopher L. Hemme University of Oklahoma hemmecl@ou.edu

Thomas Henick-Kling Cornell University th12@cornell.edu

Isaac Y. Ho

DOE Joint Genome Institute

iyho@lbl.gov

Roger Hohnsbeen

Researcher

roger@hohnsbeen.com

Dawn E. Holmes

University of Massachusetts dholmes@microbio.umass.edu

Amy J. Horneman

University of Maryland School of Medicine

ahornema@epi.umaryland.edu

Benjamin Horowitz

JGI PGF

horowitz2@llnl.gov Stephen J. Horton

Union College hortons@union.edu

Ping Hu

Lawrence Berkeley National Lab

phu@lbl.gov

Phil Hugenholtz

DOE Joint Genome Institute PHugenholtz@lbl.gov

Lonnie O. Ingram

University of Florida-IFAS

ingram@ufl.edu

Natalia Ivanova

DOE Joint Genome Institute

NNIvanova@lbl.gov

Thomas W. Jeffries USDA, FS, FPL twjeffri@wisc.edu

Paul R. Jensen

Scripps Institution of Oceanography

pjensen@ucsd.edu

Jhaveri Jinal

DOE Joint Genome Institute

jajhaveri@lbl.gov David R. Johnson

University of California, Berkeley

daverj@ce.berkeley.edu

Kou-San Ju

University of California, Davis

kju@ucdavis.edu Patricia I. Kale

DOE Joint Genome Institute

kale1@llnl.gov

Marina Kalyuzhnaya University of Washington mkalyuzh@u.washington.edu

Staci Kane

Lawrence Livermore National Laboratory

kane11@llnl.gov

Samuel Kaplan University of Texas

Health Science Center at Houston samuel.kaplan@uth.tmc.edu

Kenneth G. Karol

University of Washington kkarol@u.washington.edu

Gerrit H. Kema

Plant Research International B.V. GERT.KEMA@WUR.NL

Cheryl A. Kerfeld

University of California, Los Angles

kerfeld@mbi.ucla.edu

Arshi Khan

DOE Joint Genome Institute

khan5@llnl.gov

Nicole King

University of California, Berkeley

nking@berkeley.edu

Stephen Kingsmore

NCGR sfk@ncgr.org

Martin G. Klotz University of Louisville mgk@mgklotz.com

Michael L. Knotek

Knotek Scientific Consulting m.knotek@verizon.net

Arthur Kobayashi

DOE Joint Genome Institute kobayashi1@llnl.gov

Thomas D. Kocher

University of New Hampshire

tdk@cisunix.unh.edu

Alexander Kozik

University of California, Davis

akozik@atgc.org

Jerome J. Kukor Rutgers University kukor@aesop.rutgers.edu

Alan Kuo

DOE Joint Genome Institute

akuo@lbl.gov

Nikos Kyrpides

DOE Joint Genome Institute NCKyrpides@lbl.gov

Alla L. Lapidus

DOE Joint Genome Institute

alapidus@lbl.gov

Gerard R. Lazo

USDA-ARS Western Regional Research Center

lazo@pw.usda.gov

Jared R. Leadbetter

California Institute of Technology

jleadbetter@caltech.edu

Patrick Lemaire

**CNRS** 

lemaire@ibdm.univ-mrs.fr

Johan Leveau

Netherlands Institute of Ecology

(NIOO-KNAW) j.leveau@nioo.knaw.nl

Todd M. Lowe

University of California, Santa Cruz

lowe@soe.ucsc.edu

Susan M. Lucas

DOE Joint Genome Institute

lucas11@llnl.gov

Athanasios Lykidis

DOE Joint Genome Institute

alykidis@lbl.gov

Ronald C. Mackenzie

University of Texas Health Science Center at

Houston

ronald.c.mackenzie@uth.tmc.edu

Jon K. Magnuson

Pacific Northwest National Laboratory

jon.magnuson@pnl.gov

Dina F. Mandoli

University of Washington mandoli@u.washington.edu

Reinhold C. Mann

Oak Ridge National Laboratory

mannrc@ornl.gov

Betty K. Mansfield

Oak Ridge National Laboratory

mansfieldbk@ornl.gov

Meng Markillie

Pacific Northwest National Laboratory

meng.markillie@pnl.gov

Eric J. Mathur Diversa Corporation eric.mathur@diversa.com

Julie A. Maupin-Furlow University of Florida jmaupin@ufl.edu

Mark J. McBride

University of Wisconsin, Milwaukee

mcbride@uwm.edu

Sandra McFarland

DOE Joint Genome Institute Somcfarland@lbl.gov

J. Mitchell McGrath

**USDA-ARS** 

mitchmcg@msu.edu

Michael J. McInerney University of Oklahoma mcinerney@ou.edu

David Mead

Lucigen

dmead@lucigen.com

Monica Medina

University of California, Merced

mmedina@ucmerced.edu

Galit Meshulam-Simon

Stanford University

meshulam@stanford.edu

Axel Meyer

University of Konstanz axel.meyer@uni-konstanz.de

Charles D. Miller Utah State University

Utah State University cdmiller@biology.usu.edu

David A. Mills

University of California damills@ucdavis.edu

Marissa Mills

Oak Ridge National Laboratory MILLSMD@ORNL.gov

Brent D. Mishler

University of California, Berkeley bmishler@calmail.berkeley.edu

Herve Moreau

Oceanological Observatory of Banyuls

h.moreau@obs-banyuls.fr

Jenna L. Morgan

DOE Joint Genome Institute

ilmorgan@lbl.gov

Kenneth H. Nealson

University of Southern California

knealson@usc.edu

Irene L. Newton Harvard University garcia@fas.harvard.edu

Shirley F. Nishino

Georgia Institute of Technology

sn81@ce.gatech.edu

Kenneth M. Noll

University of Connecticut kenneth.noll@uconn.edu

Howard Ochman University of Arizona hochman@email.arizona.edu

Graham W. O'Hara Murdoch University gohara@murdoch.edu.au

Regina A. O'Neil

University of Massachusetts rtarallo@microbio.umass.edu

**Bobby Otillar** 

DOE Joint Genome Institute

RPO till ar@lbl.gov

Norman R. Pace

University of Colorado-Boulder

nrpace@colorado.edu

Jasmyn Pangilinan

DOE Joint Genome Institute

jlpangilinan@lbl.gov

Rebecca E. Parales

University of California, Davis

reparales@ucdavis.edu

Juan V. Parales

University of California, Davis

jvparales@ucdavis.edu

Andrew H. Paterson University of Georgia paterson@uga.edu

Jayant Patil

DOE Joint Genome Institute

impatil@lbl.gov

Renè A. Perrier

DOE Joint Genome Institute

RAPerrier@lbl.gov

Yvette M. Piceno

Lawrence Berkeley National Laboratory

ympiceno@lbl.gov

Nicolas Pinel

University of Washington dc4el@u.washington.edu

Samuel Pitluck

DOE Joint Genome Institute

s\_pitluck@lbl.gov

Darren Platt

DOE Joint Genome Institute

dmplatt@lbl.gov

Martha L. Posada-Buitrago DOE Joint Genome Institute mlposada-buitrago@lbl.gov

Silke Pradella

German Collection of Microorganisms

and Cell Cultures (DSMZ) silke\_pradella@t-online.de

Flavia A. Ramirez

University of Illinois, Urbana-Champaign

framirez@uiuc.edu

Jason Raymond

Lawrence Livermore National Laboratory

raymond20@llnl.gov

Wayne G. Reeve Murdoch University reeve@murdoch.edu.au

Paul M. Richardson

DOE Joint Genome Institute pmrichardson@lbl.gov

Aaron O. Richardson Indiana University arichard@bio.indiana.edu

Kirsti M. Ritalahti

Georgia Institute of Technology

krita@ce.gatech.edu

Dan Rokhsar

DOE Joint Genome Institute

DSRokhsar@lbl.gov

Noah Rosenzweig University of California noahr@nature.berkeley.edu

Eddy Rubin

DOE Joint Genome Institute

EMRubin@lbl.gov

Asaf A. Salamov

DOE Joint Genome Institute

aasalamov@lbl.gov

Nori Satoh

**Kyoto University** 

satoh@ascidian.zool.kyoto-u.ac.jp

Thomas M. Schmidt Michigan State University tschmidt@msu.edu

Johannes C. Scholten

Pacific Northwest National Laboratory

johannes.scholten@pnl.gov

Gary P. Schroth

Solexa

gschroth@solexa.com

David A. Sela

University of California dasela@ucdavis.edu

Keelnatham T. Shanmugam University of Florida shan@ufl.edu

Harris Shapiro

DOE Joint Genome Institute

hjshapiro@lbl.gov

Thomas J. Sharpton

University of California, Berkeley

sharpton@berkeley.edu

Zhen Shi

University of Illinois, Urbana-Champaign

zshi@uiuc.edu David S. Shin

Scripps Research Institute davess@scripps.edu

Mark Silby

Tufts University School of Medicine

mark.silby@tufts.edu

Hauke Smidt

Wageningen University hauke.smidt@wur.nl

Alexei V. Sorokin

Gènètique Microbienne CRJ INRA

sorokine@jouy.inra.fr

Kevin R. Sowers University of Maryland Biotechnology Institute sowers@umbi.umd.edu

Shawn R. Starkenburg Oregon State University starkens@onid.orst.edu

Rob Steele

University of California, Irvine

resteele@uci.edu
Craig Stephens
Santa Clara University
cstephens@scu.edu

Jonathon Stillman

San Francisco State University

stillmaj@sfsu.edu Wesley D. Swingley Arizona State University wswingley@msn.com

John A. Tainer

Scripps Research Institute

jat@scripps.edu

Hanno Teeling

Max Planck Institute for Marine Microbiology

hteeling@mpi-bremen.de

Michael P. Thelen

Lawrence Livermore National Laboratory

mthelen@llnl.gov

Richard H. Thomas

Southern Illinois University rthomas@zoology.siu.edu

Hope N. Tice

DOE Joint Genome Institute

tice1@llnl.gov Ravi P. Tiwari Murdoch University

Murdoch University r.tiwari@murdoch.edu.au

Christian M. Tobias USDA, ARS

ctobias@pw.usda.gov

Tamas Torok

Lawrence Berkeley National Laboratory

ttorok@lbl.gov

Sucheta Tripathy

Virginia Polytechnic University and

State University sutripa@vbi.vt.edu

Stephan Trong

DOE Joint Genome Institute

trong1@llnl.gov

Hank Tu

DOE Joint Genome Institute

hctu@lbl.gov

Jerry Tuskan

Oak Ridge National Laboratory

tuskanga@ornl.gov

Tina L. Tyler-Gresham Idaho State University tyletina@isu.edu

Gene W. Tyson

University of California, Berkeley gtyson@nature.berkeley.edu

Parag A. Vaishampayan parag\_vai@yahoo.co.in

Fernando Valle Genencor International fvalle@danisco.com

Claire Vieille

Michigan State University

vieille@msu.edu

Irene Wagner-Dobler

German Biotechnology Research Centre (GBF)

iwd@gbf.de

Jeffrey J. Walker University of Colorado jeffrey.walker@colorado.edu

Jens Walter

University of Otago

jens.walter@stonebow.otago.ac.nz

Ronald A. Walters

Intelligence Technology Innovation Center

ra.walters@pnl.gov

Boris Wawrik Rutgers University bwawrik@yahoo.com

Sharlene C. Weatherwax U.S. Department of Energy

sharlene.weatherwax@ science.doe.gov

David A. Weisblat

University of California, Berkeley

weisblat@berkeley.edu

Greg Werner

DOE Joint Genome Institute

gmwerner@lbl.gov

Rachel Whitaker

University of California, Berkeley rwhitaker@nature.berkeley.edu

Larry J. Wilhelm Oregon State University

wilhelml@science.oregonstate.edu

John Willis Duke University jwillis@duke.edu

Alexandra Z. Worden

Rosenstiel School of Marine and Atmospheric

Science

aworden@rsmas.miami.edu

Martin Wu

The Institute for Genomic Research

mwu@tigr.org

Dongying Wu

The Institute for Genomic Reasearch

dwu@tigr.org

John I. Yoder

University of California, Davis

jiyoder@ucdavis.edu

Jr-Kai Sky Yu

California Institute of Technology

jkyu@caltech.edu

Xueling Zhao

DOE Joint Genome Institute

xzhao@lbl.gov

Jizhong Zhou

University of Oklahoma jzhou@rccc.ou.edu

Stephen H. Zinder Cornell University shz1@cornell.edu

Gerben J. Zylstra Rutgers University

zylstra@aesop.rutgers.edu

## **Author Index**

Abbott, Eric	20	Caldwell, David G 41	Eisen, J. A50
Acharya, Chaitanya R	64	Cameron, R. Andrew 16	Elmore, Brad39
Ackerman, Eric		Campbell, John46	El-Sheikh, Amal F39
Adams, Lorrie		Cande, W. Z	Everett, Karin44
Adley, Catherine C		Carlier, Jean37	Falciatore, Angela12
Adney, William S		Cary, S. Craig	Fields, Matthew W 26, 30, 72
Aerts, Andrea		Cavanaugh, C. M 50	Fillapova, Tatiana
Allen, Andrew		Cerdeño-Tárraga, Ana M 59	Fourcade, H. M. 66
Allen, Eric E		Chain, Patrick S. G39	Fowler, Tom32
Allgaier, Martin		Chain, Patrick S.G36	Francino, M. Pilar27, 68
		Chakicherla, Anu36	Frederick, Reid D53
Almana Fixind			
Almaas, Eivind		Challacombe, J. F	Fritz-Laylin, Lillian21
Altman, Tomer		Chan, Clara	Froula, Jeff27, 68
Andersen, Gary L9, 23		Chapman, Jarrod 13, 54	Furman, Craig54
Anderson, Anne J 10	-	Chen, Fallon41	Fuss, Jill O28
Anderson, Olin D		Cheng, Quen J28	Galleron, Nathalie61
Andrews, Justen		Chertkov, O	Garbelotto, Matteo55
Arcaina, Marlon	20	Chew, Aline Gomez Maqueo 14	Gathman, Allen32
Argyris, Jason M	41	Chisholm, Penny1	Gaut, Brandon S18
Arkin, Adam P	17	Chistoserdova, Ludmila 16, 35	Gentry, Terry72
Arp, Daniel J	39	Chivian, Dylan 17	Gerards, Saskia43
Auger, Sandrine		Churchill, Alice C. L37	Gholba, Sumedha64
Bailey, Vanessa		Clark, Richard M 18	Gihring, Thomas17
Baker, Brett J		Coesel, Sacha12	Gilna, Paul16, 71
Baker, Scott		Coffroth, Mary Alice25	Glavina del Rio, Tijana14, 29
Banfield, Jillian F 10		Colbourne, John	Gogarten, J. Peter49
Banks, Jody		Coleman-Derr, Devin	Goltsman, Eugene61
Barabote, Ravi D.		Conrad, Amber L	Gonzalez, B53
Barondeau, David P		Copeland, Alex 14, 20	Gonzalez-Pastor, J. E53
Barry, Kerrie W 14, 35, 50		Crossman, Curt C42	Goodwin, Lynne
Beatty, J. Thomas		Crous, Pedro37	Goodwin, Eyillic
		Cullen, Dan3	Gregory, Kelvin51
Beller, Harry			
Bergelson, Joy M		Culley, David E	Grigoriev, Igor V30, 33
Berglund, J. Andrew		D'Haeseleer, Patrik	Grimwood, Jane33
Bergmann, David J.		Daum, Christopher20	Gu, Yong Q
Berry, Alison M		Davidson, Eric	Gunathilake, Pradeepa66
Berube, Paul		Dawson, Scott	Gunsalus, Robert46
Biebl, Hanno		De la Iglesia, R53	Guo, Yalong18
Blackall, Linda L		de Lorenzo, V 53	Hall, Anne E18
Blankenship, Robert E		De Martino, Alessandra 12	Hammon, Nancy M14
Bohnert, Hans J	12	de Vos, Willem M60	Han, Cliff S 16, 26, 30, 71
Boore, Jeffrey L., 19, 22, 31,	41,	DeFrancesco, Robert6	Hao, Jicheng64
44, 53, 66		Dehal, Paramvir S 22, 27	Harris, J. Kirk52
Borevitz, Justin O	18	Dejesa, Liza C64	Hauser, Loren J30, 39
Bottomley, Peter J		DeLong, Edward F4	Hazen, Terry C17, 72
Bowler, Chris		Denef, Vincent64	Heijde, Marc12
Bradford, Kent J		Deole, Ratnakar H23	Hemme, Chris L30, 72
Branscomb, Elbert		Derrien, Muriel60	Henrissat, B71
Brettin, Tom S16		DeSalvo, Michael25	Hettich, Robert64
Bristow, James37		DeSantis, Todd 9, 23, 24	Hickey, William J39
Brockman, Fred J17		Detter, J. Chris 14, 50, 72	Hitomi, Chiharu58
Brodie, Eoin L 9, 17, 23		Dhodda, Vinay6	Ho, Isaac54
Brokstein, Peter	25	DiDonato, Michael58	Hoegg, Simone31
Bronner-Fraser, Marianne		Dodge, Tim	Hoff, Wouter D23
Bruce, David C 30, 35, 50			
		Donoghue, Michael J 44	Holland, Linda Z
Brumm, Phillip		Du, Mai	Holmes, Dawn
Bryant, Donald A		Dunkle, Larry D	Hommes, Norman G39
Busby, Jennifer A. C		Dusheyko, Serge	Hooper, Alan B
Caldwell, Bob	09	Ehrlich, S. Dusko61	Horton, Stephen32

#### Authors

Hristova, Krassimira36	Lucas, Susan M 14, 16, 20, 36,	Peterson, S. Brook4
Hu, P9	71	Pett-Ridge, Jennifer13
Hugenholtz, Phillip 4, 10, 24, 52	Lynch, Michael19	Pfrender, Michael19
Hura, Greg58	Mackenzie-Dodds, J. A 66	Phan, Richard24
Hwang, Mona64	Magnuson, Jon 32	Philip, Lena20
Israni, Sanjay14	Maheswari, Uma12	Philips, Michael29
Ivanova, Natalia4	Malfatti, Stephanie A 39	Piceno, Yvette9, 23, 24
Jabbari, Kamel12	Mandoli, Dina F44	Pieper, D. H53
Jackson, Terri29	Mangogna, Manuela12	Pintor, Michael29
James, Andy37	Manohar, Chitra25	Pitluck, Sam14
Janssen, Peter H60	Marcobal, Angela48	Plugge, Caroline M60
Jeans, Christopher64	Maresca, Julia A14	Poret-Peterson, Amisha T39
Jeffries, Thomas W33	Markillie, Meng L45	Posada-Buitrago, Martha L53
Jensen, Paul R34	Martín, Héctor García4	Pradella, Silke70
Jin, Yong-Su33	Mastrian, Steven D64	Pukall, Rüdiger70
Kalyuzhnaya, Marina G35	Mathrakott, Smitha 41	Putnam, Nik54
Kane, Staci36	Matthies, Heather J64	Ramirez, Flavia A54
Kaplan, Samuel2	Matvienko, Marta41	Ramos, Hector64
Karol, Kenneth G44	Maumus, Florian	Rashid, Harun69
Keller, Keith24	Maupin-Furlow, Julie A 38	Rayko, Edda12
Kema, Gert H. J37	Mayer, Klaus F. X18	Raymond, Jason13
Kennedy, Megan29	McBride, M. J71	Reagan, Russell66
Kesseli, Rick41	McGrath, Mitch	Renzaglia, Karen44
Khan, Arshi29		Richardson, Aaron O55
Kim, UnMi46	McHale, Leah K	Richardson, Paul M 6, 14, 16,
Kingsmore, Stephen38	McInerney, Michael J 46	17, 26, 30, 33, 35, 36, 50,
Kirkland, P. Aaron38	McMahon, Katherine D 4	61, 71, 72
Klotz, Martin G39	Mead, David	Rieseberg, Loren41
Knapp, Steve J41	Medina, Mónica	Rio-Hernandez, Luis46
Kohara, Yuji72	Meyer, Axel	Robinson, David29
Kothe, Erika32	Michelmore, Richard W 41	Rogers-Lowery, Constance25
Kozik, Alexander41	Mihalkanin, Danielle	Rohlin, Lars46
Krupp, Rebecca46	Miller, Charles	Rokhsar, Dan54
Kueh, Jennifer44	Miller, Don	Rosenzweig, Noah55
Kunin, Victor4	Mills, David A	Roux, Nicolas
Lafrades, Melanie20	Mishler, Brent D 44, 48	Rubin, Eddy72
	Misra, M	
Lal, Shruti	Moberg, Jordan	Salamov, Asaf
	Montsant, Anton	Satoh, Nori
Langley, Charles H	Moore, Bradley S 34	Satou, Yutaka
Lapidus, Alla L 14, 16, 17, 26,	Morgan, Jenna	Saunders, E. H
30, 61 Lanjagra Pagael 40	Moser, Duane P	Saunders, Liz
Lapierre, Pascal	Moutakki, Housna46	Sayavedra-Soto, Luis A39
Laplaza, José M	Myers, G. L71	•
Larimer, Frank W 14, 36, 39	N'Guessan, Lucie	Schadt, Christopher W72
Larrahando, Juliana51	Nanavati, Dhaval M	Schmidt, Radomir
Larsen, Niels24	Neuffer, Barbara 18	Schoenfeld, Thomas
Lavelle, Dean O41	Nevin, Kelly51	Scholten, Johannes C. M56
Lazo, Gerard R	Newton, I. L. G 50	Schroth, Gary P
Leadbetter, Jared	Noll, Kenneth M	Schulz, Stefan
Lemaire, Patrick	Nordborg, Magnus 18	Schwarz, Jodi
Leveau, Johan	Normand, Philippe 11	Scow, Kate
Levy, Stuart B59	Norton, Jeanette M39	Shah, Manesh64
Lewis, Cindy25	O'Huallachain, Maeve E 64	Shah, Maulik K64
Ley, Ruth E52	O'Neil, Regina51	Shapiro, Harris57
Li, Tao14	Ochman, Howard 68	Shin, David S28, 58
Lidstrom, Mary E 16, 35	Ochoa, Oswaldo E41	Shin, Maria36
Lilly, Walt32	O'Kelley, Charlie44	Shin-I, Tadasu72
Lin, Li-Hung17	Olmstead, Richard44	Shouche, Yogesh
Lince, Michael T64	Onstott, Tullis C17	Siaut, Magali12
Lindquist, Erika 13, 33, 37	Overmann, Jörg14	Sieber, Jessica
Linkowski, Albert20	Pace, Norman R 52	Silby, Mark W59
Loero-Pequignot, Adrienne 20	Pangilinan, Jasmyn 52, 57	Sims, Ronald47
Lovley, Derek51	Passoth, Volkmar33	Singer, Steven
Lowe, Todd M44	Patterson, Melodee6	Smidt, Hauke60
	Perez-Pantoja, D 53	Smith, Alan R44

#### **Authors**

Sorokin, Alexei61	Tiedje, James 72	Ward, Bess B	39
Souza, Jr., Manoel T37	Tighe, Damon20	Ward, Joy	
Sowers, Kevin R62	Tindall, Brian70	Warnecke, Falk	4, 52
Spear, John R52	Tobias, Christian M 66	Watson, David	72
Spurrell, Cailyn20	Tomilov, Alexey 66	Webb, Richard	10
Starkenburg, Shawn R39	Tomilova, Natalya66	Wei, Xueming	39
Steele, Rob62	Torok, T 9	Weigel, Detlef	18
Stein, Lisa Y39	Torres, Manuel66	Wolf, Paul	44
Stephens, Craig63	Touchman, Jeffrey W 64	Wosten, Han	32
Stevens, Jr., Stanley38	Tripathy, Sucheta68	Woyke, T	
Stewart, F. J50	Trivedi, Zalak29	Wright, Stephen I	18
Still, David W41	Truco, María José41	Wu, Liyou	72
Stillman, Jonathon63	Tu, Hank 57	Xie, Gary	.11, 71
Struchtemyer, Christopher G. 46	Tyson, Gene W 10, 64	Yoder, John I	66
Sullivan, S50	Vaishampayan, Parag 68	You, Frank M	42
Swingley, Wesley D64	Valle, Fernando69	Young, Mark	6
Szmant, Alina25	van de Peer, Yves18	Yu, Jr-Kai Sky	72
Tainer, John A28, 58	van der Lee, Theo A.J 37	Yuen, Andy	20
Tang, Carol25	Van Deynze, Allen41	Zane, Mathew	20
Tapia, Roxanne16, 71	Vardi, Assaf 12	Zhang, Weiwen	56
Taylor, Heather L64	VerBerkmoes, Nathan 64	Zhou, Jizhong26,	30, 72
Thayer, N. N71	Vergez, Lisa M 39	Zinder, Stephen H	74
Thelen, Michael P13, 64	Vrionis, Helen51	Zoetendal, Erwin	60
Thomas, Kelley19	Waalwijk, Cees37	Zubietta, Ingrid	23
Thomas, R. H66	Wagner-Döbler, Irene 70		
Thompson, L. S71	Walker, Jeffrey J 52		

## **Notes**

Notes